

1           **ONE INPUT-CLASS AND TWO INPUT-CLASS CLASSIFICATIONS FOR**  
2           **DIFFERENTIATING OLIVE OIL FROM OTHER EDIBLE VEGETABLE OILS BY**  
3           **USE OF THE NORMAL-PHASE LIQUID CHROMATOGRAPHY FINGERPRINT OF**  
4           **THE METHYL-TRANSESTERIFIED FRACTION**

5  
6           Ana M. JIMÉNEZ-CARVELO <sup>a,✉</sup>, Estefanía PÉREZ-CASTAÑO<sup>a</sup>, Antonio GONZÁLEZ-  
7           CASADO <sup>a</sup>, Luis CUADROS-RODRÍGUEZ <sup>a</sup>

8           <sup>a</sup> Department of Analytical Chemistry, University of Granada, c/ Fuentenueva, s.n. E-18071  
9           Granada, Spain.

10  
11  
12  
13           **Abstract**

14           A new method for differentiation of olive oil (independently of the quality category) from other  
15           vegetable oils (canola, safflower, corn, peanut, seeds, grapeseed, palm, linseed, sesame  
16           and soybean) has been developed. The analytical procedure for chromatographic  
17           fingerprinting of the methyl-transesterified fraction of each vegetable oil, using normal-phase  
18           liquid chromatography, is described and the chemometric strategies applied and discussed.  
19           Some chemometric methods, such as k-nearest neighbours (kNN), partial least squared-  
20           discriminant analysis (PLS-DA), support vector machine classification analysis (SVM-C), and  
21           soft independent modelling of class analogies (SIMCA), were applied to build classification  
22           models. Performance of the classification was evaluated and ranked using several  
23           classification quality metrics. The discriminant analysis, based on the use of one input-class,  
24           (plus a dummy class) was applied for the first time in this study.

25  
26  
27           **Keywords**

28           Olive oil authentication, methyl-transesterified fraction, chromatographic fingerprinting, one  
29           input-class and two input-class classification

30  
31  
32           ✉ Corresponding author: phone: +34 958240797; fax: +34 958243328; email: [amariaic@ugr.es](mailto:amariaic@ugr.es)  
33

## 34 1. INTRODUCTION

35 Edible vegetable oils are important worldwide products, which are used as raw materials  
36 and/or ingredients in several foodstuffs. Although most vegetable oils are extracted from  
37 oilseeds, some are obtained directly from the fruit as a juice. This is the case for virgin olive  
38 oil, which is collected directly from olive fruits by mechanical procedures (grinding followed  
39 by centrifugation and/or decantation). Furthermore, in contrast to other vegetable oils, virgin  
40 olive oil is not refined for human consumption. Extra virgin olive oil is more expensive than  
41 other vegetable oils, owing to the specific process required for extraction [Jabeur, Zribi,  
42 Makni, Rebai, Abdelheidi, & Bouaziz, 2014]. For this reason, olive oils are susceptible to  
43 adulteration, with cheaper vegetable oils, to achieve an illicit profit. Unauthorized blends or  
44 adulteration of olive oil of any quality category with oils obtained from seeds is a particular  
45 problem in Spain, as well as other Mediterranean countries, which have specific legislation  
46 prohibiting the marketing of such blends. Therefore, it is desirable to develop rapid and  
47 simple methods to monitor the authenticity of olive oil.

48 The analytical methodologies applied to authenticate the olive oil are, generally, based on  
49 the quantification of certain chemical markers, which constitute a characteristic fraction of the  
50 oils [Arvanitoyannis & Vlachos, 2007; Aparicio, Morales, Aparicio-Ruiz, Tena & García-  
51 González, 2013]. Thus, families of compound, such as fatty acids, triacylglycerols (or  
52 triglycerides) or sterols, have been proposed. Other chemical fractions, such as volatile  
53 compounds or phenols, have also been used but they are not stable enough to give reliable  
54 results.

55 Triglycerides represent 95-99% of the chemical composition of vegetable oils. The  
56 compositional characterization of these compounds, determined by gas chromatography  
57 (GC) or high-performance liquid chromatography (HPLC), has been proposed for the  
58 detection of other oils due to their specific compositional profiles [Aparicio & Aparicio-Ruiz,  
59 2000; Ruiz-Samblás, Marini, Cuadros-Rodríguez, & González-Casado, 2012; Lerma-Garcia,  
60 Simó-Alfonso, Méndez, Lliberia & Herrero-Martínez, 2011]. The content of some fatty acids,  
61 such as linolenic and oleic acids, has also been used to detect blending in olive oils [Aparicio  
62 & Aparicio-Ruiz, 2000].

63 Fatty acids are quantified using GC, following derivatization to increase the volatility of the  
64 compounds, as necessary [Sánchez de Medina, El Riachy, Priego-Capote & Luque de  
65 Castro, 2014; Fernandes, Fernandes, Simas, Barrera-Arellano, Eberlin, & Alberici, 2013].  
66 Moreover, sterols are applied as markers of authenticity in vegetable oils. In order to  
67 characterize the compositional profile of these compounds, firstly, it is necessary to carry out  
68 saponification of the oil followed by isolation of free sterols by means preparative  
69 chromatography or solid phase extraction, and silanization. Then, GC analysis is performed

70 [Gázquez-Evangelista, Pérez-Castaño, Sánchez-Viñas & Bagur-González, 2013].  
71 Consequently, this methodology is difficult, tedious and time-consuming.

72 In 1993, Bierdermann et al. [Biedermann, Grob & Mariani, 1993] developed a new strategy  
73 that replaced the conventional saponification/ isolation process with a methyl-  
74 transesterification reaction. This approach, which inexplicably has been underused by the  
75 analytical community, requires less vegetable oil and facilitates extraction since soaps are  
76 not produced and the process is faster. The breakdown of molecules during  
77 transesterification leads to the formation of methyl esters from fatty acids and the liberation of  
78 sterols. Two fractions are obtained during this process: (1) the water soluble fraction, which  
79 contains the polar compounds, and (2) the organic fraction (transesterified fraction) in which  
80 fatty acid methyl esters, sterols, alcohol, monoglycerides, diglycerides and other molecules  
81 can be found. In the latter fraction, Bierdermann et al. [1993] identified methyl sterols,  
82 dimethyl sterols and linear alcohols. The methyl-transesterified fraction can be analysed by  
83 liquid chromatography to obtain a characteristic fingerprint of each vegetable oil, which might  
84 also be used to detect potential adulteration. The fingerprinting methodology is based on  
85 treating the entire or a part of the chromatogram as a whole, without identifying or quantifying  
86 each compound [Ellis et al., 2012; Cuadros-Rodríguez, Ruiz-Samblás, Valverde-Som, Pérez-  
87 Castaño, & González-Casado, 2016]. Effective implementation of fingerprinting requires the  
88 use of chemometric tools. Chromatograms are exported as data vectors and treated with  
89 pattern recognition methods to develop multivariate classification or regression models,  
90 which are suitable to differentiate among vegetable oils.

91 The chemometric methods fall in to two groups: supervised and non-supervised [Naes,  
92 Isaksson, Fearn & Davies, 2002; Marini, 2010]. In the first group, the category or class  
93 membership of each data vector is known and used to build the multivariate model. In  
94 contrast, the model from non-supervised methods does not consider this information  
95 [Correia & Ferreira, 2007]. Supervised classification methods are used to categorize objects  
96 (samples) in two or more classes according to a set of characteristic features of each class.  
97 Such features are extracted previously from information supplied for standard objects and  
98 selected during the model-training step. In order to perform the classification process, two  
99 approaches could be applied: discriminant analysis methods and class-modelling methods  
100 [Bevilacqua, Nescatelli, Bucci, Magrì, Magrì & Marini, 2014]. A discriminant method works by  
101 finding the borders between groups of objects from different classes, while a class-modelling  
102 method defines a particular enclosed space for all the objects from the same class.

103 Sometimes [class-modelling methods are described](#) as 'one-class classifier' (e.g. SIMCA)  
104 [where](#) each class is modelled independently [Brereton, 2011] [and as](#) many model as classes  
105 are built. [Classification](#) is performed considering all the models simultaneously. In our

106 opinion, [however](#), this term should not be used as synonym [for](#) class-modelling since [the two](#)  
107 [might](#) be confused.

108 This study proposes a multivariate method to differentiate olive oil from other edible  
109 vegetable oils. For this, the methyl-transesterified fraction from each oil class (olive and non-  
110 olive) was analysed using normal-phase conventional high-performance liquid  
111 chromatography. The chromatograms (chromatographic fingerprints), acquired by means of  
112 a corona charged aerosol detector (CAD), were used as a source of analytical information to  
113 set up the classification models. Some common and well-established classification methods  
114 were applied, such as k-nearest neighbours (kNN), partial least squares discriminant  
115 analysis (PLS-DA), support vector machine classification (SVM-C) and soft independent  
116 modelling of class analogies (SIMCA). Two classification strategies were tried for each  
117 classification method according to the number of class used for model training: two input-  
118 class and one input-class classifications. In addition, the use of a 'dummy' class was  
119 proposed for applying discrimination methods with a one input-class strategy. The  
120 classification results from each method and strategy were compared and ranked on the basis  
121 of several classification performance metrics [Cuadros-Rodríguez, Pérez-Castaño & Ruiz-  
122 Samblás, 2016].

123

## 124 **2. MATERIALS AND METHODS**

125

### 126 **2.1. Chemicals**

127 All solvents used were HPLC grade. Isopropanol, n-hexane, methanol and tert-butyl methyl  
128 ether (TBME) were provided by the VWR International Eurolab, S.L. (Barcelona, Spain).  
129 Sodium methoxide (MeONa), citric acid monohydrate, and anhydride sodium sulphate were  
130 purchased from Merck (Darmstadt, Germany). The nitrogen (99.9999 %) used was provided  
131 by Air Liquid (Madrid, Spain).

132

### 133 **2.2. Chromatography**

134 The analyses were carried out with an Agilent 1100 series liquid chromatograph (Santa  
135 Clara, USA) equipped with a column thermostat (Eppendorf CH30), a quaternary pump and  
136 degasser auto sampler. Detection was performed with a corona charged aerosol detector  
137 (CAD) (ESA [Biosciences](#) Inc., Chemsford, MA, USA). Agilent ChemStation software (rev.  
138 B.02.01-SR1) for LC systems was used to collect and process data.

139 The HPLC analysis was carried out on a (250 × 4 mm i.d, 5 µm) column Lichrospher® 100  
140 CN. The column temperature was set at 30 °C during the entire operation. The composition  
141 of the mobile phase was n-hexane/isopropanol (96:4, v/v) at a flow rate of 1.2 mL min<sup>-1</sup>. The  
142 injection volume was 20 µL and the run time was only 8 min.

143

### 144 **2.3. Samples**

145 A total of 127 vegetable oil samples of different types were analysed. The samples were  
146 obtained directly from local providers. More specifically, 66 samples were different categories  
147 of marketed olive oil (virgin extra, virgin, refined+virgin, and pomace+virgin), and the other 61  
148 were canola, safflower, corn, peanut, sunflower, (no-specified) seed, grapeseed, palm,  
149 linseed, sesame, and soybean oils. Table 1 summarizes the different vegetable oils and the  
150 number of samples analysed for each.

151

TABLE 1
---------

152

### 153 **2.4. Sample preparation**

154 Previous to the chromatographic analysis, a transesterification reaction was applied. A  
155 modification of the procedure described by Biedermann et al [Biedermann, Grob & Mariani,  
156 1993] was used. For this, 0.1 g of oil was weighed into a centrifuge tube. 1 mL of extracting  
157 agent (MeONa at 10 % in methanol in TBME, 4:6 (v/v)) was added and mixed with the oil.  
158 The mixture was stirred for 20 s and then allowed to stand for 20 min. This step was  
159 repeated twice. Then, 1 mL of water and 8 mL of hexane was added, and the mixture  
160 centrifuged for 3 min at 1,500 g. The aqueous phase was removed with a Pasteur pipette  
161 and 1 mL of 1 % citric acid in water added to the residual. Again, the aqueous phase was  
162 eliminated before 2 g of anhydrous sodium sulphate added and the mixture allowed to stand  
163 for 20 min. The methyl-transesterified organic fraction was passed through a  
164 polytetrafluoroethylene (PTFE) membrane syringe filter (0.22 µm) and the solution stored at  
165 -20°C until analysis. For the chromatographic analysis, 200 µL of transesterified solution was  
166 added to a 2 mL HPLC vial before 450 µL of n-hexane was added and 20 µL injected.

167

### 168 **2.5. Chemometrics**

169 The raw data files from each chromatogram were obtained in a CSV file and exported to  
170 MATLAB (version R2013a). In this way, a data vector composed of 839 variables defined

171 each chromatogram. The data pre-processing was done with a home-programmed MATLAB  
172 function, "Medina" (version 10) [Pérez Castaño et al., 2015]. This function implemented  
173 several algorithms from the MATLAB Bioinformatics Toolbox™ and 'icoshift' (*interval*  
174 *correlation optimized shifting*) algorithm [Tomasi, Savorani & Engelsen, 2011] to align the  
175 peaks of the chromatograms. The steps for pre-processing the data were: (1) raw  
176 chromatograms data grouping and overlay; (2) selection of interval of interest in  
177 chromatograms; (3) filtered of the raw chromatograms data to eliminate noise of signal  
178 analytical; (4) correction of the baseline using the 'msbackadj' function (included in the  
179 Bioinformatics Toolbox™); (5) alignment of the peaks with the function 'icoshift'; and finally  
180 (6) mean centring of the data set.

181 The original dataset was divided in two groups: (1) the training set, which was made up of 84  
182 oil samples (44 olive oil, 40 non-olive oil), and (2) the validation (or test) set composed of the  
183 remaining oil samples (25 olive oil, 18 non-olive oil). Selection was carried out **ensuring** that  
184 a sample from each class of oil was **allocated** to one vegetable oil group or the other. Within  
185 each group, the samples were **selected** randomly.

186 Classification of the vegetable oils was achieved using multivariate chemometric pattern  
187 recognition in the PLS\_Toolbox (version 7.5.2, Eigenvector Research, Wenatchee, WA).

188

### 189 *Principal Component Analysis (PCA)*

190 The main aim of PCA is to reduce the number of variables to evaluate which components  
191 contain essential information. Each principal component (PC) is a lineal combination  
192 between original variables (chromatographic intensities) of each object, which are described  
193 as:  $X=T \times P^T$  where X is the original data matrix, T is the score matrix and P is the transposed  
194 loading matrix [Bro & Smilde, 2014].

195

### 196 *k-Nearest Neighbours (kNN)*

197 kNN is a based-similarity classification method that uses distance measures between  
198 objects. The classification is carried out as follows: first, a multidimensional hyperspace is  
199 defined with the training set and, then, the prediction is performed. The assigned class of  
200 each new object will be one where the number of k-neighbours is largest [Correia & Ferreira,  
201 2007; Alsberg, Goodacre, Rowland & Kell, 1997] and k is an odd integer that could be  
202 selected previously. Each sample is classified based on the most represented classes of the  
203 k-nearest samples.

204

205 *Partial Least Squares Regression-Discriminant Analysis (PLS-DA)*

206 PLS-DA is a latent variable-based method that builds a PLS regression model on latent  
207 variables (LV) to establish limits of the class and, then, carries out a discriminant analysis  
208 (DA) to classify the samples [Bevilacqua, Nescatelli, Bucci, Magrì, Magrì & Marini, 2014;  
209 Ballabio & Consonni, 2013]. In order to develop the best PLS model, it is necessary to  
210 optimize the number of LVs to be used in advance.

211

212 *Support Vector Machine Classification (SVM-C)*

213 SVM is a based-machine learning method. As with PLS-DA, SVM-C works by carrying out a  
214 SVM regression model for building hyperplanes in a multidimensional space that separates  
215 the different classes of objects [Xu, Zomer, Brereton, 2006; Luts, Ojeda, Van de Plas, De  
216 Moor, Huffel & Suykends, 2010]. SVM can be optimized with 'nu' and 'C' parameters. The  
217 former optimizes a model with an adjustable parameter Nu [ $0 \rightarrow 1$ ], which indicates the upper  
218 boundary for the number of misclassifications allowed, and the latter optimizes a model with  
219 an adjustable cost function C [ $0 \rightarrow \infty$ ], which indicates how strongly misclassifications should  
220 be penalized [SVM Function Settings, Eigenvector Documentation wiki. URL  
221 [http://wiki.eigenvector.com/index.php?title=SVM\\_Function\\_Settings](http://wiki.eigenvector.com/index.php?title=SVM_Function_Settings). Accessed 29.06.15].

222

223 *Soft Independent Modelling of Class Analogies (SIMCA)*

224 This chemometric technique performs as many principal component (PC) models as input-  
225 classes in study and, then, the classification is carried out from the distance of the object to  
226 the centre of each principal component score space [Bevilacqua, Nescatelli, Bucci, Magrì,  
227 Magrì & Marini, 2014]. The assignment of each unknown sample to a particular class is  
228 based on the nearest distance to the corresponding regions established by the PC model.

229

230 *Two input-class (2iC) and one input-class (1iC) classification*

231 Usually a two-class classification method (or more properly, two output-class classification)  
232 requires using two input-classes, the target class and the non-target class (in this paper,  
233 olive and non-olive classes). The term 'output' is related to the classes to which objects or  
234 samples will be assigned as result of the classification while the term 'input' refers to the  
235 class that is used to train the classification model [Cuadros-Rodríguez, Pérez-Castaño, &  
236 Ruiz-Samblás, 2016]. It is also possible to perform the same classification method by training  
237 the model with a single input-class, *i.e.* the target class.

238 Working with one input-class classification has significant advantages. For example, in food  
239 authentication, the model can be built with data from only genuine foods (target class) and it  
240 is not necessary to have other foods (non-target class) to train the model. Consequently, the  
241 necessary experimental work is halved. When this model is applied on unknown foods, only  
242 those recognized by the model will be declared as "true" whereas the remaining food will be  
243 refused and they are candidate to be considered as "false". The greater the training set of  
244 genuine representative samples, the better the quality classification performance. Obviously,  
245 this strategy can be applied to differentiate olive oils from other edible vegetable oils.

246 This is a very easy task when a class-modelling method is applied because each class is  
247 modelled independently. This approach has been used already with SIMCA [López, Trullós,  
248 Callao & Ruisánchez, 2014]. However, the discriminant methods, such as PLS-DA or SVM-  
249 C, usually require two input-classes to define the discrimination model. Although some  
250 proposals have been reported as one-class PLS (OCPLS) [Xu, Yan, Cai & Yu, 2013], in fact,  
251 this is a class-modelling method. To resolve this drawback, a fictitious class or 'dummy' class  
252 could be used as a substitute for the second class (the non-target class). The dummy class  
253 should be defined from inactive objects that do not have analytical information of interest for  
254 the target class, e.g. analytical blank.

255 In this study, both 2iC and 1iC strategies were applied to devise a classification model for  
256 differentiating olive oil from non-olive oil. When the 1iC was applied, a dummy class was  
257 from the dataset provided using 30 chromatograms for the solvent blank.

258

### 259 **3. RESULTS AND DISCUSSION**

260 A chromatogram was recorded for each vegetable oil sample. Figure 1 shows the  
261 superposed chromatograms for all vegetable oil samples. Two regions could be easily  
262 differentiated: (1) region A shows a major peak, which was essentially composed of methyl  
263 esters of fatty chains derived from triglycerides, phospholipids, waxes, esterified sterols and  
264 free fatty acids, and (2) region B that was composed of several minor peaks and contained  
265 information about the families of free sterols and terpenic alcohols.

266

FIGURE 1

267

#### 268 *Exploratory Analysis*

269 A principal component analysis (PCA) was carried out considering the dataset composed of  
270 the whole chromatogram from each vegetable oil sample. Four PCs were enough to explain



271 87.16% of the variance. Figure 2a shows the biplot for scores on the PC2-PC1 plane. PC1  
272 and PC2 explained 56.2% and 17.3% of the variance, respectively. Three groups of  
273 vegetable oils could be distinguished easily, which corresponded with olive oil (centre left),  
274 palm oil (top left) and other vegetable oils (right).

275 Two additional PCA were carried out, one for each of the regions of the chromatograms to  
276 check if both regions grouped the oil samples in the same way. Figure 2b and 2c show the  
277 biplot for scores on the PC2-PC1 plane, corresponding to the data subset from regions A and  
278 B, respectively.

279

FIGURE 2

280

281 The three scores biplots allowed differentiation in similar ways to the three sample groups  
282 and, in principle, there was no conclusive reason –from a chemometric point of view– to  
283 select one dataset or the others. However, looking the chromatographic retention time,  
284 region A was preferred to minimize the analysis time.

285

#### 286 *Two input-class (2iC) classification*

287 In order to differentiate olive oils from other vegetable oils, a two input-class (2iC)  
288 classification strategy was applied where the target class was 'olive oil' and the alternative  
289 class was, generally, denoted as 'non-olive oil'. Four well-established classification methods  
290 were tried: kNN, PLS-DA, SVM-C and SIMCA.

291 To differentiate the two vegetable oils classes,  $k=3$  was enough to decide the neighbour  
292 distance in the kNN model. The olive class was defined by a class predicted probability value  
293 equal to 1, while the non-olive class was defined by a probability of 0. Classification of the  
294 samples contained in the validation set was carried out directly by the software. All of olive oil  
295 samples were well classified (probability=1) and the non-olive oil samples were also  
296 classified correctly (probability=0), with exception of palm oil samples, which had an  
297 assigned probability of 0.5; in this case, we also classified these samples as non-olive oil.

298 The PLS-DA model was built using four LVs, with 92.91% of the variance explained. Each  
299 class was characterized by a predicted value around 1 for olive oil and 0 for non-olive oil.  
300 The classification threshold established by the software from the corresponding probability  
301 curves was a predicted value of 0.6 for the olive oil class.

302 The SVM-C model was optimized with 'C-svc' and 'nu-svc' parameters, and the results  
303 obtained in both cases were similar. As in the kNN method, the olive class was assigned to  
304 samples with a predicted probability value equal to 1 and the non-olive class was defined by  
305 samples with a probability of 0. The software also carried out the class assignment for the  
306 validation samples. Both olive and non-olive oil samples were classified correctly.

307 Figure 3 (a) and (b) show the classification plots obtained from both 2iC PLS-DA and 2iC  
308 SVM-C methods.

309

FIGURE 3 (a) (b)

310

311 The application of SIMCA implies building of two PC models. The number of PCs chosen for  
312 each model was four for 'olive oil' and five for 'non-olive oil'. The software carried out  
313 classification of the validation dataset based on the Q-residual values for each olive oil  
314 sample. Samples with a normalized Q-residual (95% confidence) value less than  $\sqrt{2}$  were  
315 classified as olive oil.

316 Table 2 shows the different quality performance features for the 2iC classification method,  
317 calculated according to the olive oil samples classification. These show that, in this  
318 classification scenario, 2iC kNN and SVM-C were faultless and, in contrast, 2iC SIMCA  
319 performed poorly.

TABLE 2

320

### 321 *One input-class (1iC) classification*

322 Since the aim of this study was differentiation of olive oil from other vegetable oils, the  
323 classification model could be trained using objects from the olive oil class. In this way, the  
324 objects recognized by the model should be assigned as olive oil whereas the remainder,  
325 regardless of their botanical origin, should be classified as non-olive oils. The same  
326 classification methods, kNN, PLS-DA, SVM-C and SIMCA, were applied. For each, a  
327 confidence interval-based classification criterion was established because the default  
328 classification threshold defined by the software was not applicable.

329 The kNN model conformed with  $k=3$ , but did not generate good results and all the non-olive  
330 oil samples were misclassified because they were considered to be "nearest neighbours" to  
331 the target class (olive oil). Thus, the 1iC strategy was not applicable for the kNN method.

332 Two strategies were applied for 1iC PLS. In a first step, a PLS-DA classification with dummy  
333 class was performed using the PLS\_Toolbox. Next, a one-class PLS without dummy class  
334 (OCPLS) was performed using software provided by Xu [Xu, Yan, Cai, & Yu, 2013].

335 A conventional PLS-DA was built with only two LVs explaining 99.74% of the variance. A  
336 confidence interval was established centred on 1, which was the value assigned for the olive  
337 oil class. The width of the interval was calculated as plus/minus 2.33-times the standard  
338 deviation (s) from the predicted values for the olive oil samples in the training set. The  
339 expression 2.33×s is an "ad-hoc" application, recommended by the EC for estimating the  
340 decision limit (DL), formally termed as CC $\alpha$ , concerning the performance of analytical  
341 methods in the case of substances for which no permitted limit has been established [EU  
342 Commission Decision, 2002]. This decision limit defines the limit at and above which it can  
343 be concluded with an error probability of  $\alpha$  that a sample is non-compliant. Strictly speaking,  
344 the correct expression would be:  $DL = 1.645 \times \sqrt{2} s$ , where 1.645 is the critical value for the  
345 standardized normal distribution ( $\alpha = 1\%$ ) and s the whitening-batch standard distribution of the  
346 difference between the predicted values of both the target and the non-target samples, which  
347 are considered equal, and consequently:  $s = \sqrt{s^2(\text{targ}) + s^2(\text{non-targ})} = \sqrt{2} s(\text{targ})$ . The  
348 coefficient 2.33 is the result of multiplying  $1.645 \times \sqrt{2}$  (or 1.414). The confidence band is  
349 calculated from an estimated standard deviation of 0.026.

350 Figures 4(a) and 4(b) show the classification plots obtained from the 1iC PLS-DA method.

351

FIGURE 4 (a) (b) (c) (d)

352

353 Most olive oil samples were included within the confidence interval while the no-olive oils  
354 were not. However, samples in the non-olive oil class were separate into two subclasses on  
355 both sides of the interval. The seed oils were located in the upper region whereas the palm  
356 oils were in the lower region. This surprising outcome implies the classification scenario is  
357 suitable for implementing a three output-class classification (olive oil, palm oil, and  
358 generically seed oil) from a one input-class strategy, making it possible to distinguish palm oil  
359 from a classification model trained only with olive oils. Currently, the authors are working to  
360 develop and apply this approach.

361 OCPLS was built with seven LVs. For classification purpose, the regions pre-established by  
362 the software were used. The results are showed in Table 3.

363 SVM-C classification was carried out by optimization of 'C-svc' and 'nu-svc' parameters, and  
364 the results obtained in both cases were similar. All the oil samples were assigned to a

365 predicted probability close to 1 and always distant from 0, which was assigned to the dummy  
366 class. Specifically, the probability value was ca. 0.98 for the olive oil class and less but  
367 always greater than 0.92 for the non-olive class. The confidence interval was determined by  
368 means of a probability interval centred on the average olive oil class probability calculated  
369 from the training set. The width of the interval was also calculated as plus/minus 2.33 times  
370 the standard deviation from the predicted class probability. The estimated value of the  
371 probability standard deviation was 0.0015. Figures 4 (c) and (d) show the classification plots  
372 obtained from the 1iC SVM-C method.

373 Finally, the SIMCA method was also applied. Since SIMCA is a class-modelling method, two  
374 options were applied: i) a double PCA model using both the olive oil and dummy classes;  
375 and ii) a single model from the olive oil class. In both cases, five PCs were used to build the  
376 olive oil model. In both cases, a sample oil was classified as olive oil when the normalized Q-  
377 residual (95% confidence) value was less than  $\sqrt{2}$ .

378 Table 3 shows the quality performance features of the different 1iC classification methods. In  
379 contrast with the 2iC classification method, the 1iC PLS-DA provided the best classification  
380 performance and 1iC SIMCA (without dummy class) was, again, the worst.

381

TABLE 3
---------

382

#### 383 4. CONCLUSIONS

384 In this study, several classification methods were applied and the application strategy has  
385 been discussed. Four well-established classification methods were used, namely kNN,  
386 PLS-DA, SVM-C and SIMCA. Each was applied using two classification strategies  
387 designated as two input-class (2iC) and one input-class (1iC) classifications. This is the first  
388 time a dummy class has been used to perform discriminant analysis methods with a single  
389 input-class. This new approach does not require having and analysing samples from the non-  
390 target class (non-olive vegetable oil) in order to train the classification model. In order to  
391 assess and rank the different classification methods and strategies, several quality  
392 classification metrics were calculated. kNN and SVM-C, on the one hand, and PLS-DA, on  
393 the other, proved to be the best when 2iC or 1iC classification strategies were applied,  
394 respectively. Furthermore, the proposed analytical method consumed less time in sample  
395 treatment (transesterification reaction, 60 min) and chromatographic elution (8 min) than  
396 previous methods (saponification, 120 min) and chromatographic analysis (40 min).

397

- Alsberg, B.K., Goodacre, R., Rowland, J.J., & Kell, D.B. (1997). Classification of pyrolysis mass spectra by fuzzy multivariate rule induction-comparison with regression, K-nearest neighbour, neural and decision-tree methods. *Analytica Chimica Acta*, *348*, 389–407.
- Aparicio, R., & Aparicio Ruíz, R. (2000). Authentication of vegetable oils by chromatographic techniques. *Journal of Chromatography A*, *881*, 93–104.
- Aparicio, R., Morales, M.T., Aparicio-Ruiz, R., Tena N., & García-González D.L. (2013). Authenticity of olive oil: Mapping and comparing official methods and promising alternatives. *Food Research International*, *54*, 2025–2038.
- Arvanitoyannis, I.S., & Vlachos, A. (2007). Implementation of physicochemical and sensory analysis in conjunction with multivariate analysis towards assessing olive oil authentication/adulteration. *Critical Reviews in Food Science and Nutrition*, *47*, 441–498.
- Ballabio, D., & Consonni, V. (2013). Classification tools in chemistry. Part 1: linear models. PLS-DA. *Analytical Methods*, *5*, 3790–3798.
- Bevilacqua, M., Nescatelli, R., Bucci, R., Magrì, A.D., Magrì, A.L., & Marini, F. (2014). Chemometric Classification Techniques as a Tool for Solving Problems in Analytical Chemistry. *Journal of AOAC International*, *97*, 19–28.
- Biedermann, M., Grob, K., & Mariani, C. (1993). Transesterification and on-line LC-GC for determining the sum of free and esterified sterols in edible oils and fats. *Fett Wissenschaft Technologie - Fat Science Technology*, *95*(4), 127–133.
- Brereton, R.G. (2011). One-class classifiers. *Journal of Chemometrics*, *25*, 225–246.
- Bro, R., & Smilde, A. (2014). Principal component analysis. *Analytical Methods*, *6*, 2812–2831.
- Correia, M.M., & Ferreira, P.R.M. (2007). Non-supervised pattern recognition methods: Exploring chemometrical procedures for evaluating analytical data. *Quimica Nova*, *30*, 481–487.
- Cuadros Rodríguez, L., Pérez Castaño, E., & Ruiz Samblás, C. (2016). Quality performance metrics in multivariate classification methods for qualitative analysis. *Trends in Analytical Chemistry*, *80*, 612–624.
- Cuadros Rodríguez, L., Ruiz Samblás, C., Valverde Som, L., Pérez Castaño, E., & González Casado, A. (2016). Chromatographic fingerprinting: An innovative approach for food 'identification' and food authentication - A tutorial. *Analytica Chimica Acta*, *909*, 9–23.
- Ellis, D.I., Brewster, V.L., Dunn, W.B., Allwood, J.W., Golovanov, A.P., & Goodacre R. (2012). Fingerprinting food: current technologies for the detection of food adulteration and contamination. *Chemical Society Reviews*, *41*, 5706-5727.
- EU Commission Decision (2002/657/EC) implementing Council Directive 96/23/EC concerning the performance of analytical methods and the interpretation of results, Official Journal of the European Communities, L 221/8-36.
- Fernandes, A.M.A.P., Fernandes, G.D., Simas, R.C., Barrera-Arellano, D., Eberlin, M.N., & Alberici, R.M. (2013). Quantitation of triacylglycerols in vegetable oils and fats by easy ambient sonic-spray ionization mass spectrometry. *Analytical Methods*, *5*, 6969-6975.
- Gázquez Evangelista, D., Pérez Castaño, E., Sánchez Viñas, M., & Bagur González, M.G. (2013). Using offline HPLC-GC-FID 4-Desmethylsterols Concentration Profiles, Combined with Chemometric Tools, to Discriminate Different Vegetable Oils. *Food Analytical Method*, *7*, 912–925.
- Jabeur, H., Zribi, A., Makni, J., Rebai, A., Abdelheidi, R., & Bouaziz, M. (2014). Detection of chemlali extra-virgin olive oil adulteration mixed with soybean oil, corn oil, and sunflower oil by using GC and HPLC. *Journal of Agricultural and Food Chemistry*, *62*, 4893–4904.
- Lerma García, M.J., Simó Alfonso, E.F., Méndez, A., Lliberia, J.L., & Herrero Martínez, J.M. (2011). Classification of extra virgin olive oils according to their genetic variety using linear discriminant analysis of sterol profiles established by ultra-performance liquid chromatography with mass spectrometry detection. *Food Research International*, *44*, 103–108.

- López, M.I., Trullós, E., Callao, M.P., & Ruisánchez, I. (2014). Multivariate screening in food adulteration: untargeted versus targeted modelling. *Food Chemistry*, *147*, 177-181.
- Luts, J., Ojeda, F., Van de Plas, R., De Moor, B., Van Huffel, S., & Suykens, J.A.K.(2010). A tutorial on support vector machine-based methods for classification problems in chemometrics. *Analytica Chimica Acta*, *665*,129–145.
- Marini, F. (2010). Classification Methods in Chemometrics. *Current Analytical Chemistry*, *6*, 72–79.
- Naes, T., Isaksson, T., Fearn T., & Davies, T. (2002). A user-friendly guide to multivariate calibration and classification, NIR Publications, Chichester.
- Pérez Castaño, E., Ruiz Samblás, C., Medina Rodríguez, S., Quirós Rodríguez, V., Jiménez Carvelo, A.M., Valverde Som, L., González Casado, A., & Cuadros Rodríguez, L. (2015) Comparison of different analytical classification scenarios: application for the geographical origin of edible palm oil by sterolic (NP) HPLC fingerprinting. *Analytical Methods*, *7*, 4192–4201.
- Ruiz Samblás, C., Marini, F., Cuadros Rodríguez, L., & González Casado, A. (2012). Quantification of blending of olive oils and edible vegetable oils by triacylglycerol fingerprint gas chromatography and chemometric tools. *Journal of Chromatography B: Analytical Technologies in the Biomedical and Life Sciences*, *910*, 71–77.
- Sánchez de Medina, V., El Riachy, M., Priego-Capote, F., & Luque de Castro, M.D.(2014). Composition of fatty acids in virgin olive oils from cross breeding segregating populations by gas chromatography separation with flame ionization detection. *Journal of the Science of Food and Agriculture*, *In press*, doi:10.1002/jsfa.7030.
- SVM Function Settings, Eigenvector Documentation Wiki [accessed 09 November 2015], [http://wiki.eigenvector.com/index.php?title=SVM\\_Function\\_Settings](http://wiki.eigenvector.com/index.php?title=SVM_Function_Settings)
- Tomasi, G., Savorani, F., & Engelsen, S.B. (2011). Icoshift: An effective tool for the alignment of chromatographic data. *Journal of Chromatography A*, *1218*, 7832–7840.
- Xu, L., Yan, S-M., Cai, C-B., Yu, X-P. (2013). One-class partial least squares (OCPLS) classifier. *Chemometrics and Intelligent Laboratory Systems*, *126*, 1-5.
- Xu, Y., Zomer, S., & Brereton, R.G. (2006). Support Vector Machines: A recent method for classification in chemometrics. *Critical Reviews in Analytical Chemistry*, *36*, 177–188.

**Table 1.** Class and types of vegetable oils analysed.

<b>Class</b>	<b>Category/type</b>	<b>Nº samples</b>
Olive oil (66 samples)	Virgin extra	50
	Virgin	4
	"Refined" <sup>a</sup>	6
	"Pomace" <sup>b</sup>	6
Non-olive oil (61 samples)	Canola	4
	Safflower	4
	Corn	5
	Peanut	5
	Sunflower <sup>c</sup>	13
	Seeds	6
	Grapeseed	4
	Palm	7
	Linseed	3
	Sesame	3
Soybean	7	

<sup>a</sup> A marketed blend of refined and virgin olive oil (5-10 %).

<sup>b</sup> A marketed blend of pomace and virgin olive oil (5-10 %).

<sup>c</sup> Two samples of high-oleic sunflower oils are included.

**Table 2.** Values of the quality performance features of the different 2iC classification methods.

<b>Performance features</b>	<b>kNN</b>	<b>PLS-DA</b>	<b>SVM-C</b>	<b>SIMCA</b>
Sensibility (or Recall)	1.00	1.00	1.00	0.48
Specificity	1.00	0.94	1.00	1.00
Positive predictive value (Precision)	1.00	0.96	1.00	1.00
Negative predictive value	1.00	1.00	1.00	0.58
Youden index	1.00	0.94	1.00	0.48
Positive likelihood rate	–	18.00	–	–
Negative likelihood rate	0.00	0.00	0.00	0.52
F-measure	1.00	0.98	1.00	0.65
Discriminant power	–	–	–	–
Efficiency (or Accuracy)	1.00	0.98	1.00	0.70
AUC (Correctly classified rate)	1.00	0.97	1.00	0.74
Matthews correlation coefficient	1.00	0.95	1.00	0.53
Kappa coefficient	1.00	0.95	1.00	0.44

*The hyphen "-" is signifying that the performance feature cannot be determined*



**Table 3.** Values of the quality performance features of the different 1iC classification methods.

Performance features	With a dummy class				Without dummy class	
	kNN	PLS-DA	SVM-C	SIMCA	OCPLS	SIMCA
Sensibility (or Recall)	1.00	0.96	0.88	0.88	0.80	0.80
Specificity	0.00	1.00	1.00	0.83	0.89	1.00
Positive predictive value (Precision)	0.58	1.00	1.00	0.88	0.91	1.00
Negative predictive value	–	0.95	0.86	0.83	0.76	0.78
Youden index	0.00	0.96	0.88	0.71	0.69	0.80
Positive likelihood rate	1.00	–	–	5.28	7.20	–
Negative likelihood rate	–	0.04	0.12	0.14	0.23	0.20
F-measure	0.74	0.98	0.94	0.88	0.85	0.89
Discriminant power	–	–	–	0.86	0.83	–
Efficiency (or Accuracy)	0.58	0.98	0.93	0.86	0.84	0.88
AUC (Correctly classified rate)	0.50	0.98	0.94	0.86	0.84	0.90
Matthews correlation coefficient	–	0.95	0.87	0.71	0.68	0.79
Kappa coefficient	0.00	0.95	0.86	0.71	0.67	0.77

*The hyphen "–" is signifying that the performance feature cannot be determined*

## FIGURE CAPTIONS

**Figure 1.** Superposed chromatograms of the 127 vegetable oil samples showing the two characteristic regions (see text for additional explanations). The chromatograms have been previously pre-processed with the exception of the mean centring step.

**Figure 2.** PCA scores biplot obtained from the fingerprint data of the methyl-transesterified fraction of the 127 vegetable oil samples: **(a)** PC2-PC1 plane of the whole chromatogram; **(b)** PC2-PC1 plane from region A; **(c)** PC2-PC1 plane from region B.

**Figure 3.** Classification plots on the 2iC classification strategy: **(a)** PLS-DA; **(b)** SVM-C.

**Figure 4.** Classification plots on the 1iC classification strategy: **(a)** and **(b)** PLS-DA full plot and zoomed plot, respectively; **(c)** and **(d)** SVM-C full plot and zoomed plot, respectively. In addition, the confidence bands are superposed on **(b)** and **(d)** plots.

Figure 1

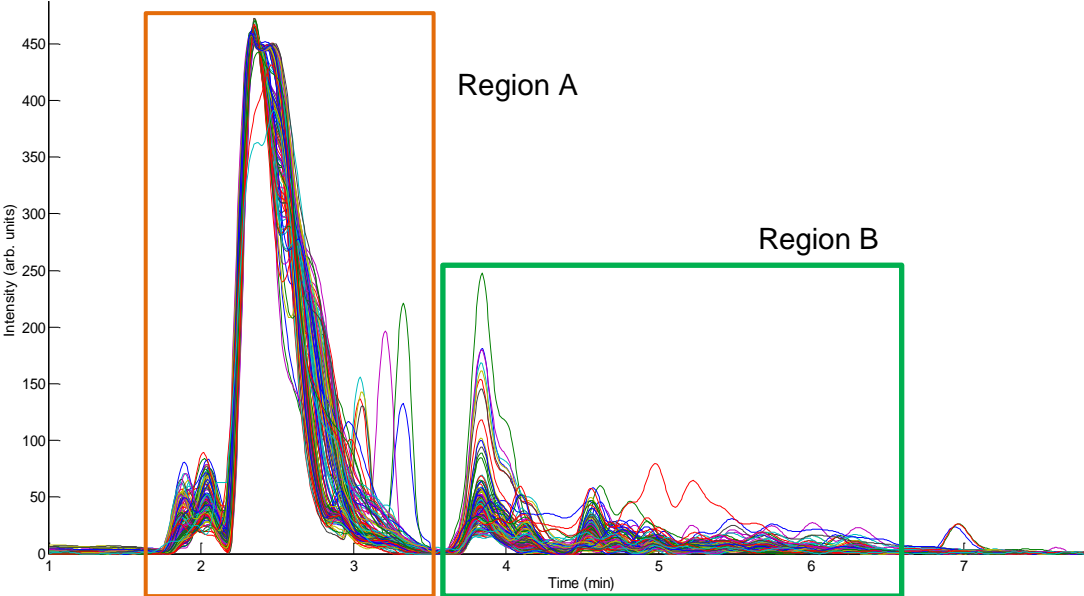


Figure 2

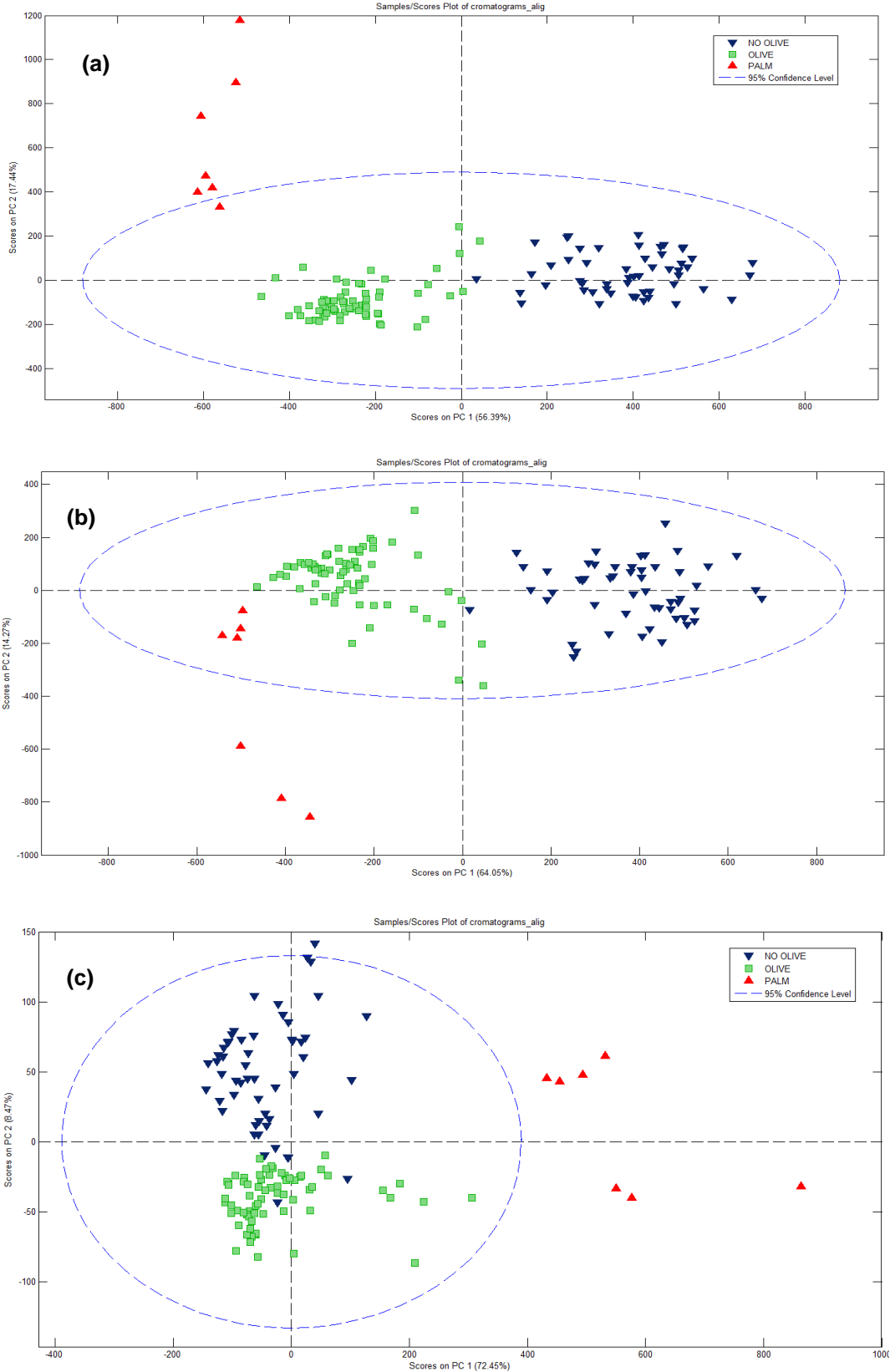


Figure 3

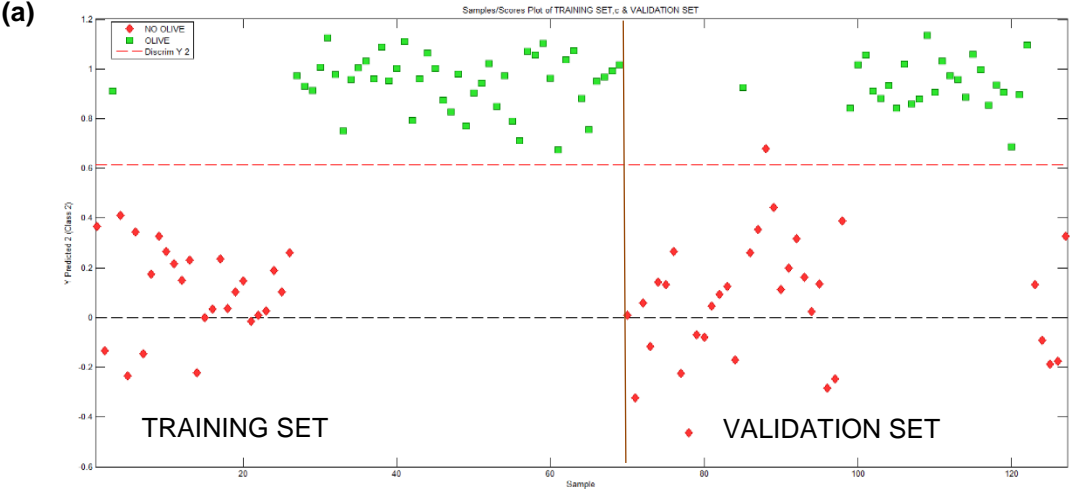


Figure 4

