

# 1 GeneCodis: interpreting gene lists through enrichment analysis and integration of 2 diverse biological information

3 Ruben Nogales<sup>1,\*</sup> ([ruben.nogales@fdi.ucm.es](mailto:ruben.nogales@fdi.ucm.es)), Pedro Carmona-Saez<sup>1,\*</sup> ([pcarmona@fis.ucm.es](mailto:pcarmona@fis.ucm.es)), Miguel  
4 Vazquez<sup>2</sup> ([miguel.vazquez@fdi.ucm.es](mailto:miguel.vazquez@fdi.ucm.es)), Cesar Vicente<sup>1</sup> ([cesar.vicente@fis.ucm.es](mailto:cesar.vicente@fis.ucm.es)), Xiaoyuan Yang (),  
5 Francisco Tirado<sup>1</sup> ([ptirado@dacya.ucm.es](mailto:ptirado@dacya.ucm.es)), Jose M. Carazo<sup>3</sup> ([carazo@cnb.csic.es](mailto:carazo@cnb.csic.es)) Alberto Pascual-  
6 Montano<sup>1</sup> ([pascual@fis.ucm.es](mailto:pascual@fis.ucm.es))

7 <sup>1</sup> Computer Architecture Department, <sup>2</sup> Software Engineering Department, Complutense University.  
8 Madrid. Spain. <sup>3</sup> Biocomputing Unit. National Center for Biotechnology. CNB-CSIC. Madrid. Spain.

9 \* Both authors contributed equally to this work

10 GeneCodis: <http://genecodis.dacya.ucm.es/>

11 Keywords: functional analysis, enrichment analysis, gene ontology

## 14 ABSTRACT

15 GeneCodis is a web server application for functional analysis of gene lists able to integrate annotations  
16 from different sources and find modular patterns of interrelated annotations. This tool differs from other  
17 approaches in the way the heterogeneous information is integrated to search for annotations that  
18 frequently co-occur in a set of genes. This integrative approach has proved to be useful for the functional  
19 interpretation of high-throughput experiments and, therefore, a new version of the system has been  
20 developed to expand its functionality and scope. This new version GeneCodis not only provides  
21 functional information but also regulatory patterns with the potential to integrate both sources of  
22 information in the same analysis. In addition, user-defined annotations can also be included. To cover  
23 most of the necessities of the research activity, traditional singular enrichment has been implemented and  
24 more organisms and gene identifiers are now supported. The application has been also re-engineered to  
25 improve performance, accessibility and scalability. In addition to the web browser, GeneCodis now can  
26 be accessed through a public web services interface, enabling users to launch jobs from their own scripts  
27 and workflows. The application is freely available at <http://genecodis.dacya.ucm.es>

## 28 INTRODUCTION

29 High-throughput experiments such as DNA microarrays or proteomics techniques have been widely used  
30 during the last decade and currently they are standard technologies in many research centers. Although  
31 these methodologies generate huge amounts of data, the challenge does not lie in the analysis or data  
32 processing, in which significant advances have been done by the bioinformatics community, but rather in  
33 interpreting such datasets to get biological knowledge and meaningful information to formulate new  
34 hypothesis.

35 An essential task in this context is to translate gene signatures into information that can assist in the  
36 understanding of the biological mechanisms. In the last few years several methods and tools have been  
37 developed to interpret large lists of genes or proteins using information available in biological databases.  
38 The common idea in most of these methods is to find functional descriptors that are significantly enriched  
39 in the gene signature with respect to the entire genome or other reference list. Annotations from different  
40 resources such as Gene Ontology (GO) (Ashburner, Ball et al. 2000) or KEGG (Kanehisa, Araki et al.  
41 2008) are commonly used in this context.

42 The first type of methods that emerged in this field were focused on evaluating the frequency of  
43 individual annotations and apply an statistical test to determine what annotations are significantly  
44 enriched in a input list with respect to a reference list, usually the whole genome or all genes in the

45 microarray. Several tools were developed following this idea, and although each application introduces  
46 variations such as different statistical tests, sources of annotations or supported organisms, they all  
47 performed the same type analysis and offer slightly variations in the results. Good reviews of such  
48 methods can be found in (Khatri and Draghici 2005; Dopazo 2006). A fresh line of research appeared  
49 with the observation that the use of thresholds to select the significant genes could lead to underestimate  
50 the effect of significant biological effects during the functional analysis. This derived in a new and  
51 different analytical concept in which the distribution of annotations is evaluated in the whole list of genes,  
52 sorted by their correlation with the phenotype. Different methods followed this approach after the pioneer  
53 work of Tamayo and Subramanian published in 2007 (Subramanian, Kuehn et al. 2007), the gene set  
54 enrichment analysis (GSEA). However, both standard enrichment tools and GSEA methods evaluate each  
55 annotation independently from the others without taking into account the potential relationships between  
56 them. Nevertheless, most of the annotations in biological databases are interconnected because they are  
57 associated to common genes. Patterns that contain these relationships among annotations can provide  
58 invaluable information and extend our understanding of biologic events associated to the experimental  
59 system. It is therefore highly desirable to incorporate these relationships among annotations in the  
60 functional analysis of gene lists and there are new tools that attempt to extract this type of information  
61 (see a review in (Huang da, Sherman et al. 2009))

62 In 2007 we introduced GeneCodis (Carmona-Saez, Chagoyen et al. 2007), a tool for modular enrichment  
63 analysis oriented to integrate information from different sources and find enriched combinations of  
64 annotations in large lists of genes or proteins. This approach represented a step forward in the functional  
65 enrichment analysis because of its capacity in integrating heterogeneous annotations and discovering  
66 significant combinations among them. Since its original publication, this tool has achieved more than  
67 25,000 jobs submissions from all over the world and has been referenced in different works. Even a  
68 mirror has also been recently set up at the Center for Bioinformatics in Peking University to facilitate the  
69 access in the Asian region. In this work we present a new version of the software with an improved  
70 functionality, performance, accessibility and extended scope. Firstly, we have expanded the type of  
71 information that can be analyzed by the application incorporating new types of annotations such as  
72 microRNAs and transcription factors. In this way, the new version of GeneCodis offers the possibility to  
73 mine not only functional information but also regulatory patterns with the potential to integrate both  
74 sources of annotations in the same analysis. Moreover, the application now allows researchers to submit  
75 their own annotations and perform a joint analysis with the rest of annotations provided in the application.  
76 As in the first version, this release of the software finds concurrent annotations enriched in the input list,  
77 but we have also included the analysis of individual annotations (singular enrichment analysis). With this  
78 new feature GeneCodis covers more possible analytical scenarios than in the first version. We have also  
79 increased the type of gene identifiers that are supported including commercial platforms such as  
80 Affymetrix Probe Ids and new organisms.

81 From a technical point of view, GeneCodis has been completely reengineering making it faster, flexible  
82 and efficient. The algorithm to retrieve sets of concurrent annotations has been improved and  
83 implemented to run in a multi-grid environment that is more suitable to handle large number of jobs  
84 simultaneously, improving in this way the performance and the throughput of the system. The application  
85 now can be accessed in a programmatic way by Web Services. This allows researchers to include its  
86 functionalities in data analysis pipelines. Finally, the new friendly interface is designed to facilitate the  
87 use of the tool and now results include more useful information through different graphs and file formats.

## 88 **FEATURES AND FUNCTIONALITY**

89 GeneCodis has gone through a lot of changes since its first publication, but the tool works in a similar  
90 way. In this web-based application the users have to select the organism and the biological annotations  
91 that will be considered in the analysis and then load the gene list to be analyzed. The whole genome is  
92 used by default as a reference set but users can also load their own reference list. No more information or  
93 user's parameters are mandatory at this point. As advance options, it is possible to select the preferred  
94 statistical test among three possible alternatives: hypergeometric test (default), chi-square test or both.  
95 Annotations will be considered in the concurrent analysis only if they appear in at least a minimum  
96 number of genes. This is known as minimum support and it is set to three by default. And finally users  
97 can omit or select the multiple hypothesis correction method between two alternatives: false discovery  
98 rate method (default) or permutation-based correction. In addition, a new feature of this version allows  
99 users to submit a file with a list of user-defined annotations that can be considered in the analysis together  
100 with the rest of selected annotations.

## 101 **Functional Analysis In GeneCodis**

102 As we have commented previously, the enrichment analysis of individual annotations was the first  
103 method introduced for functional analysis of large lists of genes or proteins, being also the most popular  
104 one. There is a large collection of tools that implements this type of enrichment analysis, most of them  
105 focused on the analysis of Gene Ontology annotations. The arrival of GSEA methods turned the  
106 enrichment analysis of individual annotations from a gene-centered to a gene-set based analysis. These  
107 methods, although extremely useful for the interpretation of gene lists, do not exploit the inter-  
108 relationship that exists among gene annotations. In this context, tools such as GeneCodis provide a new  
109 way to analyze functional information by taking into account the relationships among annotations  
110 associated to common genes in the list. This analysis offers different advantages with respect to singular  
111 enrichment methods. Joint terms may contain unique biological meaning for a given study, not held by  
112 individual terms. For example the combination of two terms such as *Apoptosis* and *Mitochondria* may be  
113 enriched in a gene list while the individual annotations are not significant if evaluated independently in  
114 the same list of genes. This may indicate that mitochondria apoptotic pathways can be more significant  
115 in the system than other apoptotic processes. But probably more interesting is the possibility to integrate  
116 and jointly analyze information that covers different aspects of the biology of the genes.

117 GeneCodis is one of the few tools that offer these options for the functional analysis of gene lists by  
118 implementing an algorithm based on the extraction of frequent itemsets (see (Carmona-Saez, Chagoyen et  
119 al. 2006) for details). Nevertheless, one of the weak points of this type of analysis is that it can increase  
120 the complexity of the results because redundant patterns may be generated. Therefore, it would be  
121 beneficial for researchers to have a unified system that integrates both type of analysis, the evaluation of  
122 individual annotations and co-annotations for a better exploration of the high throughput experimental  
123 results.

124 In this new version of GeneCodis we have included the singular enrichment analysis precisely to take into  
125 account these two types of analysis. As results, besides the combinations of annotations enriched in the  
126 list of genes, in this new version GeneCodis also provides the analysis of individual annotations, showing  
127 those from the selected categories that are enriched in the input list. For example, if two categories of  
128 annotations are selected for the analysis, the results will include three different lists: one with the  
129 concurrent analysis results and two with singular enrichment information of each one of the types of  
130 annotations. Moreover, these results are provided in different formats: html tables, tabulated text files,  
131 xml files, pie charts and bar graphs.

## 132 **New Sources of Annotations: Integrating Functional and Regulatory Information**

133 Among the multiple resources of information, Gene Ontology is by far the most popular one for  
134 functional analysis of gene lists. This is reasonable due to the rich content of GO in terms that describe  
135 the functional role of genes at a molecular level and the initiative of different consortiums to annotate  
136 complete genomes with GO terms. Earlier enrichment tools were mainly based on the analysis of GO  
137 terms although annotations from other sources such as KEGG or Biocarta ([www.biocarta.org](http://www.biocarta.org)) were being  
138 incorporated in more recent applications. Although GO covers three aspects of the biology of genes;  
139 Biological Process, Cellular Component and Molecular Function, the former is the information in which  
140 researchers have been mainly interested for the functional characterization of gene lists. This is in part  
141 because biological process annotations provide explicit information to interpret the biological  
142 mechanisms that may be associated to the experimental system. Indeed, pathway information contained in  
143 KEGG or Biocarta covers similar aspects of the biology of genes than GO Biological Processes  
144 annotations.

145 Nevertheless, beyond functional information there are other properties of genes and proteins that can be  
146 also very useful to interpret biological systems, such as information related to transcriptional mechanisms.  
147 There are different sources of regulatory information that have been incorporated to enrichment tools  
148 (Dennis, Sherman et al. 2003; Al-Shahrour, Minguez et al. 2006; Abascal, Carmona-Saez et al. 2008,  
149 Guruceaga, Segura et al. 2009)). In GeneCodis we have included annotations related to transcriptional  
150 information from different sources. miRBase (Griffiths-Jones, Saini et al. 2008) contains putative targets  
151 of microRNAs, molecules that in the last few years have been shown as key regulators in many biological  
152 systems. From this database we have extracted the microRNAs associated to genes in different organisms;  
153 *B. taurus*, *C. elegans*, *D. rerio*, *D. melanogaster*, *G. gallus*, *H. sapiens*, *M. musculus* and *R. norvegicus*.

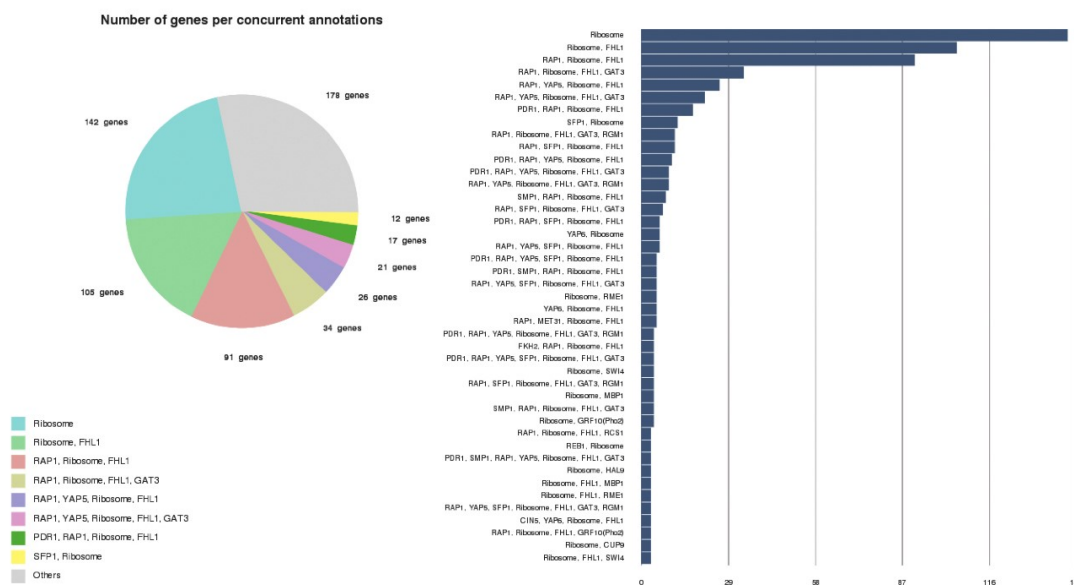
154 Using information from TransFac we have annotated genes from *H. sapiens*, *M. musculus* and *R.*  
 155 *norvegicus* with transcription factors that have binding motifs in their promoter sequences. In addition, we  
 156 have used results from chip-on-chip experiments (Carmona-Saez, Chagoyen et al. 2006) to annotate genes  
 157 in *S. cerevisiae* with transcription factors that bind to their promoter regions. This new annotations allow  
 158 users to perform enrichment analysis with regulatory information in addition to functional information.

159 Nevertheless, beyond the independent analysis of different properties of genes the integration of  
 160 heterogeneous sources of information can provide a more complete picture of the analyzed system. In this  
 161 sense, the new sources of information offer the possibility to mine gene lists to discover associations  
 162 among regulatory and functional information. This turns the potential of GeneCodis to integrate different  
 163 kinds of annotations in one of its most useful features. This is briefly illustrated in Figure 1 with a very  
 164 simple example. It shows a screenshot of the GeneCodis results from the analysis of the ribosomal gene  
 165 set in KEGG. Using also the information of transcription factors in the analysis we can see how co-  
 166 annotations of functional and regulatory information are evaluated. Indeed, enriched annotations contain  
 167 transcription factors such as RAP1, FHL1 or SFP1 that are well known to play important roles in the  
 168 regulation of ribosomal genes.

### 169 Organisms and Identifiers Supported

170 To extend the scope of GeneCodis we have also increased the number of supported organisms and gene  
 171 identifiers. GeneCodis works with most of the model organisms in biological research. The whole list of  
 172 organisms supported includes *Arabidopsis Thaliana*, *Bos Taurus*, *Caenorhabditis elegans*, *Candida*  
 173 *albicans*, *Danio rerio*, *Drosophila melanogaster*, *Escherichia coli*, *Gallus gallus*, *Homo sapiens*,  
 174 *Leishmania major*, *Mus musculus*, *Rattus norvegicus*, *Saccharomyces cerevisiae*, *Schizosaccharomyces*  
 175 *pombe* and *Vibrio cholerae*. Alternatively, to facilitate the usability, we have extended the supported gene  
 176 identifiers to cover the most common ones used in gene microarrays studies, including proprietary Ids of  
 177 commercial platforms such as Affymetrix, Agilent, Codelink and Illumina. The backbone of the system  
 178 that is now based on Ensembl gene ids has been extracted through biomart (<http://www.biomart.org>).

179



180

181 **Figure 1.** Example of the graphic generated in a typical GeneCodis analysis. Enriched combinations of  
 182 significant annotations are represented in a pie and bar graphs, where the length of the bars and size of the  
 183 slices are proportional to the number of genes supporting the significant combination of annotations.

184

## 185 Interface

186 The design of a web tool is an important part to take into account in its development. The new friendly  
187 interface has been designed to facilitate the usability of the tool. Now the application is more fresh and  
188 intuitive. Results can be explored more dynamically and are presented in different formats including pie  
189 charts and bar graphs images that allow users to explore and interpret the data in a quick look (see figure  
190 1 for an example). XML files with structured results are also provided permitting its use in data process  
191 pipelines. Finally, tabulated text files and html tables are also included in the report. These different file  
192 formats would help researches to use GeneCodis results in other applications.

193 In another context, in this new release big efforts have been done to facilitate the access of GeneCodis  
194 functionality by different ways. In addition to the classical web-server access, the tool can now be used  
195 programmatically trough the Web Services technology. Using this technology, researchers can insert  
196 functional analysis in their data mining pipelines or in other bioinformatics systems in a very  
197 straightforward manner (see figure 2 for a very simple example). There are many advantages in the use of  
198 Web Services, it is a platform-independent and language-independent technology and it is very adequate  
199 for loosely coupled systems with different architectures working together. In this way it is guaranteed that  
200 there is no any prerequisite imposed for its use. A complete tutorial can be found in the site including  
201 example scripts.

```
require 'soap/wsdlDriver'

#### Test arguments ####
#
org = "Sc" #Saccharomyces Cerevisiae
algorithm = 1 # Concurrent analysis
test = 0 # Hypergeometric p values
correction = 1 # FDR method
minsupport = 3 # Minimum support of 3 is considered in the analysis
annotations = ["GO_Biological_Process","GO_Molecular_Function"]
reference_list = [] # whole genome
input_list = ["S0000004295", "S0000005284", "S0000000598", "S0000006205",
              "S0000006183", "S0000004982", "S0000005662", "S0000001387",
              "S0000002555", "S0000005668", "S0000002585", "S0000003736"] #input list

#### Connecting to server ####
#
WSDL_URL = "http://genecodis.dacya.ucm.es/static/wsdl/genecodisWS.wsdl"
driver = SOAP::WSDLDriverFactory.new(WSDL_URL).create_rpc_driver

### Submit the analysis ###
#
job_id = driver.analyze(org,algorithm,test,correction,minsupport,input_list,annotations,reference_list)

### Check job state ###
#
status = driver.status(job_id)
while status == 1
  puts "Waiting ..."
  sleep 2
  status = driver.status(job_id)
end

# If error
if status < 0
  error_message = driver.info(job_id)
  raise "Finished with error #{ error_message }"
end

# Get results
results = driver.results(job_id)
puts results
```

202

203 Figure 2: Example of a ruby client code to invoke the GeneCodis Web Service. The access only needs  
204 three main steps: submit the analysis, ask for the job status and get the results.

## 205 **IMPLEMENTATION**

206 The algorithmic core of GeneCodis has been reviewed and drastically improved. There is a new  
207 implementation of the algorithm to extract closed itemsets based on more efficient and faster methods  
208 (Zaki and Hsiao 2005). The performance of the system depends on the number of annotations related with  
209 the genes in the input list. The new algorithm can deal with large sets of annotations in a faster and more  
210 efficient way than the algorithm implemented in the first version of GeneCodis. This is especially evident  
211 if multiple annotations are included in the same analysis and when the permutation-based test is used for  
212 the multiple hypothesis correction, in which the computing time has decreased drastically with respect to  
213 the previous version.

214 The throughput and performance of the entire system has also been improved by implementing the new  
215 algorithm and the Web Service technology in the context of a multi-grid computational environment. The  
216 new system is able to handle all submitted jobs simultaneously without queuing any of them for long  
217 periods of time. The current implementation takes advantage of one cluster with two Quad-Core Intel  
218 Xeon processors of 64 bits and two independent grid infrastructures (CyTED, <http://www.cytel.org/>, and  
219 EELA2, <http://www.eu-eela.org/>) integrated by the GridWay metascheduler (Huedo, Montero et al. 2005)

220 When users submit a query through the web site, the system launch one job for the concurrent analysis  
221 and one job for each annotation's category selected for the singular enrichment analysis. All these jobs  
222 will be executed in parallel. Users can also submit a query directly from a script using the Web Services.  
223 In all cases the flow of the system is the same. A meta-scheduler determines, depending on the  
224 computational load of the cluster and the grids, in which computational environment the jobs will be  
225 executed. When the cluster is free, then the jobs are queued and executed in the cluster. Otherwise, the  
226 jobs are sent to the less workloaded grid resource. This approach represents a cost-effective alternative to  
227 improve the throughput of the application and to guarantee its real-time performance, which is a critical  
228 step in every popular web-server application.

## 229 **CONCLUSIONS AND DISCUSSION**

230 High-throughput experimental techniques have demonstrated to be very useful allowing the study of  
231 biological systems from a global perspective. In many cases, these techniques generate huge amounts of  
232 data in the form of large gene or protein. An essential task in this context is to translate these lists to  
233 functional information that aids researchers in the interpretation of the underlying biological processes.  
234 But this interpretation is not always a trivial step; there is a lot of biological information distributed in  
235 different databases that is necessary to extract the full meaning of the data. Methods based on the  
236 enrichment analysis have proved to be very useful tools for the analysis and interpretation of such lists of  
237 genes.

238 GeneCodis is a tool designed to expand the enrichment analysis of annotations by adding the possibility  
239 of extracting not only individual annotations, but also significant combinations of them. Since its creation,  
240 this tool has proved to be a useful resource for the research activity. This encourages us to improve it by  
241 adding more functionality that expands its scope, performance and accessibility.

242 In summary, the new version of GeneCodis finds combinations of annotations but also includes in the  
243 output the traditional singular enrichment analysis. New annotations have been included such as  
244 microRNAs or transcription factors to extend the functionality of GeneCodis towards the analysis of  
245 regulatory information. Moreover, GeneCodis allows researchers to jointly analyze regulatory and  
246 functional information and to extract association patterns between both data sources. In addition, users  
247 can submit their own annotations that can be considered in the analysis together with the rest of the  
248 annotations provided in the application. The algorithm to retrieve sets of concurrent annotations has been  
249 improved and implemented to run in a multi-grid environment that is able to handle all submitted jobs  
250 simultaneously, improving in this way the performance and the throughput of the system. Finally,  
251 GeneCodis now can be accessed in a programmatic way by a web services interface; which allows  
252 researchers to include its functionalities in data analysis pipelines.

253 One of the disadvantages of the modular enrichment analysis used in GeneCodis is the intrinsic  
254 redundancy of the combined functional annotations that are usually extracted. This is due to the  
255 unavoidable natural redundancy of the information that is already known about genes and proteins and the

256 nature of the frequent itemsets mining algorithm that extract all possible combinations. Several methods  
257 still need to be implemented to filter out intrinsic repetitions and constitute a necessary future  
258 functionality. Going in the direction of creating a more self-contained application, new future releases of  
259 GeneCodis will also include Gene Set Enrichment Analysis methods to allow users the selection of all  
260 possible flavours of functional analysis in the same environment.

261  
262 The new version has been running since August 2008. Extensive tests have been carried out using  
263 synthetic and real datasets for which the outcome of the software is known. The diverse functionalities  
264 supported by this tool have been fully tested by real users who have provided feedback on issues that  
265 have helped in improving the application. We hope the renewed GeneCodis will be of interest to the  
266 scientific community.

## 267 GENECODIS AVAILABILITY

268 This application can be freely accessed through its main site at <http://genecodis.dacya.ucm.es>. A mirror in  
269 the Asian region has also been recently set up at the Center for Bioinformatics in Peking University. The  
270 mirror is available at <http://genecodis.cbi.pku.edu.cn/>.

## 271 ACKNOWLEDGEMENTS

272 This work has been partially funded by the Spanish grants BIO2007-67150-C03-02, S-Gen-0166/2006,  
273 CYTED-505PI0058, TIN2005-5619 and PS-010000-2008-1. This work also makes use of results  
274 produced by the EELA-2 project ([www.eu-eela.eu](http://www.eu-eela.eu)), co-funded by the European Commission within its  
275 Seventh Framework Programme". We would like to acknowledge Luis Canet for his technical help.  
276 Special thanks also to Prof. Jinchu Luo from the Center for Bioinformatics at Peking University for his  
277 help in setting the Asian GeneCodis mirror. A.P.M. acknowledges the support of the Spanish Ramón y  
278 Cajal program.

## 279 REFERENCES

- 280 Abascal, F., P. Carmona-Saez, et al. (2008). "ChIPCodis: mining complex regulatory  
281 systems in yeast by concurrent enrichment analysis of chip-on-chip data." *Bioinformatics* **24**(9): 1208-9.  
282  
283 Al-Shahrour, F., P. Minguez, et al. (2006). "BABELOMICS: a systems biology  
284 perspective in the functional annotation of genome-scale experiments." *Nucleic  
285 Acids Res* **34**(Web Server issue): W472-6.  
286 Ashburner, M., C. A. Ball, et al. (2000). "Gene ontology: tool for the unification of  
287 biology. The Gene Ontology Consortium." *Nat Genet* **25**(1): 25-9.  
288 Carmona-Saez, P., M. Chagoyen, et al. (2006). "Integrated analysis of gene expression  
289 by Association Rules Discovery." *BMC Bioinformatics* **7**: 54.  
290 Carmona-Saez, P., M. Chagoyen, et al. (2007). "GENECODIS: a web-based tool for  
291 finding significant concurrent annotations in gene lists." *Genome Biol* **8**(1): R3.  
292 Dennis, G., Jr., B. T. Sherman, et al. (2003). "DAVID: Database for Annotation,  
293 Visualization, and Integrated Discovery." *Genome Biol* **4**(5): P3.  
294 Dopazo, J. (2006). "Functional interpretation of microarray experiments." *OMICS*  
295 **10**(3): 398-410.  
296 Griffiths-Jones, S., H. K. Saini, et al. (2008). "miRBase: tools for microRNA  
297 genomics." *Nucleic Acids Res* **36**(Database issue): D154-8.  
298 Guruceaga, E., V. Segura, et al. (2009). "FactorY, a bioinformatic resource for genome-  
299 wide promoter analysis." *Computers in Biology and Medicine* (in press).  
300 Huang da, W., B. T. Sherman, et al. (2009). "Bioinformatics enrichment tools: paths  
301 toward the comprehensive functional analysis of large gene lists." *Nucleic Acids  
302 Res* **37**(1): 1-13.

303 Huedo, E., R. S. Montero, et al. (2005). "The GridWay Framework for Adaptive  
304 Scheduling and Execution on Grids." Scalable Computing - Practice and  
305 Experience **6**(3): 8.

306 Kanehisa, M., M. Araki, et al. (2008). "KEGG for linking genomes to life and the  
307 environment." Nucleic Acids Res **36**(Database issue): D480-4.

308 Khatri, P. and S. Draghici (2005). "Ontological analysis of gene expression data: current  
309 tools, limitations, and open problems." Bioinformatics **21**(18): 3587-95.

310 Subramanian, A., H. Kuehn, et al. (2007). "GSEA-P: a desktop application for Gene Set  
311 Enrichment Analysis." Bioinformatics **23**(23): 3251-3.

312 Zaki, M. J. and C.-J. Hsiao (2005). "Efficient Algorithms for Mining Closed Itemsets  
313 and Their Lattice Structure." IEEE Transaction on Knowledge and Data  
314 Engineering **17**(4): 12.

315

316