TITLE

Environmental Impact of Ether Carboxylic Derivative Surfactants

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ABSTRACT

The ultimate aerobic biodegradability and the toxicity of three ether carboxylic derivative surfactants having different alkyl chains and degrees of ethoxylation have been investigated. Ultimate aerobic biodegradation was examined in biodegradability screening tests by means of dissolved organic carbon determinations. The ultimate biodegradation was studied at different initial surfactant concentrations. For comparison, the characteristic parameters of the biodegradation process, such as half-life, mean biodegradation rate, and residual-surfactant concentration were determined. Increased surfactant concentrations decreased mineralization and lengthened the estimated half-life. The results demonstrate that the ultimate aerobic biodegradability is higher for the surfactants with the shortest alkyl chain and longest degree of ethoxylation.

Toxicity values of ether carboxylic derivative surfactants, and their binary mixtures, have been determined using three test organisms, the freshwater crustacea *Daphnia magna*, the luminescent bacterium *Vibrio fisheri* and the microalgae *Selenastrum capricornutum*. The influence of the surfactant structure is in the sense that the toxicity is lower for the surfactant with the shortest alkyl chain, and the higher degree of ethoxylation the smaller toxicity. The toxicity of binary mixtures of the three ether carboxylate surfactants at a 1:1 weight ratio was also measured. The less toxic mixture is formed by the surfactants having lower individual toxicity.

KEY WORDS

Anionic surfactants, Biodegradability, *Daphnia magna*, Ecotoxicity, *Microalgae, Vibrio fischeri*

1. INTRODUCTION

Surfactants are one of the most important components in laundry and household cleaning products, comprising from 15% to 40% of the total detergent formulation [1]. The class of anionic surfactants is very important, accounting for 60% of the world production [2]. Several of these compounds are biologically not degradable and present a threat to the environment [3]. The massive use of surfactants in detergents and cosmetic formulations and their subsequent disposal in aquatic systems require surfactants to be as environmentally friendly as possible. This implies the need for low toxicity and biodegradable surfactants. The environmental impact of chemicals is often determined by the ecotoxicity, which is relatively high in the case of surfactants as a result of surface activity and the action against biological membranes [4]. Surfactants have different behaviour and fate in the environment. Non-ionic and cationic surfactants have much higher sorption on soil and sediment than anionic surfactants such as lineal alkyl benzene sulphonate (LAS) [5, 6]. Most surfactants can be degraded by microbes in the environment although some surfactants such as LAS and dihydrogenated tallow dimethyl ammonium chloride (DTDMAC) as well as alkylphenols may be persistent under anaerobic conditions [7, 8, 9]. LAS were found to degrade in sludge amended soils with half-lives of 7 to 33 days [10]. Most surfactants are not acutely toxic to organisms at environmental concentrations and aquatic chronic toxicity of surfactants occurred at concentrations usually greater than 0.1 mg/L [11]. Many studies concerning biodegradability and toxicity of surfactants have been performed. The majority of the works are concerned with toxicity of surfactants to small crustaceans as *Daphnia magna* [12]. Great number of surfactants are not easily biodegradable, consequently many physicochemical methods of pre-treatment such the ozonation and other techniques of advanced chemical oxidation were developed to eliminate surfactants [13]. There has been an emphasis over the past few years on the development of surfactants and builders with improved biodegradability and also non-polluting [14]. This growing concern has led to the development of new surfactants, such as the ether carboxylic derivative surfactants.

The ether carboxylic derivative surfactants tested in the present work are anionic surfactants, under the commercial name AKYPO®, supplied by Kao corporation and with the general formula $R\text{-}O(CH_2\text{-}CH_2O)_E\text{-}CH_2\text{-}COO^-X$, where R is the alkyl chain and X= H^+ or Na. These surfactants improve the foaming quality of the detergent,

reducing the irritation level, and therefore they are used as co-surfactants in detergents which have to be in contact with the skin. These surfactants are marketed in concentrated acid form. The ultimate aerobic biodegradability of three ether carboxylic derivative surfactants with different alkyl chain and degree of ethoxylation has been investigated.

For continued advancement in the search for relationships between toxicity and structural parameters in the field of surfactants, in the present work the ecotoxicity assay with luminescent bacteria, *Daphnia magna*, and Microalgae is applied to different ether carboxylic derivative surfactants.

The purpose of this paper is to find the relationship between the ultimate biodegradation and the structure of different ether carboxylic derivative surfactants, and the influence of the initial surfactant concentration. Also the objective of this study is to determine the toxicity of the ether carboxylic derivative surfactants, and their binary mixtures (1:1 weight), to investigate the toxicological interactions between the surfactants, which take place in natural environments, and how they can affect the toxicity of the mixture, especially when acting in synergism.

2. EXPERIMENTAL PROCEDURES

2.1. Surfactants

The surfactants used in this study are the commercial ether carboxylic derivative surfactants EC-R_8E_8, EC-$R_{12\text{-}14}E_3$, EC-$R_{12\text{-}14}E_{10}$ and LAS supplied by Kao Corporation S.A. (Tokyo, Japan). Table 1 shows the degree of ethoxylation (E), the alkyl chain length (R), the % of active matter, and the critical micelle concentration (CMC) of the surfactants. The rest of the reagents used were grade chemical quality and supplied by Panreac.

Table 1

2.2. Surface Tension Measurements

The CMC values were established by measuring the surface tension of surfactant solutions with different concentrations at 25ºC, using a tensiometer model Tensiometer K11 (KRÜSS GmbH) equipped with a 2 cm platinum plate.

2.3. Biodegradation Tests

The biodegradation tests were carried out according to the OECD 301 E test, that is based on the removal of organic compounds measured as dissolved organic carbon DOC [15]. A solution of the surfactant, representing the sole carbon source for the

microorganisms, is tested in a mineral medium, inoculated and incubated under aerobic conditions in the dark for 21 days. The surfactant solution (for which the biodegradability is to be determined) is inoculated with 0.5 mL of water from a secondary treatment of a sewage-treatment plant (STP) that operates with active sludges. The biodegradation process is monitored by means of the residual surfactant concentration over time by DOC measurements, determined in filtered samples with 0.45-μm millipore. Reference assays were made with an easily biodegradable surfactant (LAS) in order to determine the activity of the microbial population present in the test medium. One flask was used for the blank, one for the reference surfactant, one for abiotic assay, and one for each surfactant concentration tested.

2.4. Toxicity Tests

Three toxicity tests were undertaken: the LumiStox® 300 test which employs the luminescent bacterium *Photobacterium phosphoreum*, the 24-h immobilization test with *Daphnia magna* (freshwater crustacea), and the 72-h algal growth inhibition test with *Selenastrum capricornutum*. In the first one, measurements were taken with the measuring system LumiStox® 300, which consists of an instrument for measuring bioluminescence and an incubation unit according to the UNE-EN ISO 11348-2 guideline [16]. The toxicity measurement is based on the luminous intensity of the marine bacteria of the strain *Vibrio fisheri* NRRL-B-11177 after a certain exposure time to a toxic substance. The luminescent bacteria, dehydrated and frozen at −18ºC, were reactivated with the suspension supplied by Dr. Lange (Dr. Bruno Lange GmbH & Co., Düsseldorf). The assay conditions were pH 7.0, NaCl concentration of 2%, all the measurements duplicated for incubation time of 15 min. When necessary, the sample was filtered prior to the assay. The toxicity values were measured as $EC_{50,}$ which is the surfactant concentration that inhibits 50% after 15 min of exposure.

Acute toxicity tests with *Daphnia magna* were performed in Standard Reference Water (SRW) according to the UNE-EN ISO 6341 guideline [17]. The tests were performed in 100 mL polystyrene vessels, with 50 mL of SRW in each one. 20 neonates (<24 h) were transferred to vessels containing different concentrations of the test chemical, and the vessels were closed with a polyethylene cap. The neonates were separated from adults every day. There was no feeding and no aeration during the tests and the tests were run at 20±1ºC. Immobility was determined visually after 24 h. For each surfactant, controls and at least five concentrations were used for the determination of the mobility

inhibition of 50% of *Daphnia* population (IC_{50}). The 72-h algal growth-inhibition test with the microalga *Selenastrum capricornutum* was administered according to the OECD 201 guideline [18]. The procedure consists of filling culture vials with appropriate volumes of nutrient medium and solutions of the surfactant being tested. At the beginning of the test, inoculums of algae were added to the vials to be tested and to the vials of control, and were kept under stable and predetermined incubation conditions.

Inocula were cultivated at 23±1ºC and constant uniform illumination (8000 lux). After 24, 48, and 72 h the algal density was determined to establish whether growth had been inhibited or stimulated with respect to control. Cell density was estimated by the optical density of the culture at 670 nm.

For all the tests, the surfactant concentration and one control were performed in triplicate for each organism tested. The surfactant concentration in the aquatic bioassays, at the beginning and at the end of the tests, was measured using a TOC analyzer for ether carboxylic derivative surfactants and a simplified spectrophotometric method using methylene blue for LAS [19].

3. RESULTS AND DISCUSSION

3.1. Biodegradability of ether carboxylic derivative surfactants

The ultimate biodegradation of the surfactants has been established under aerobic conditions in OECD tests for ready biodegradability [15]. The biodegradation process was monitored by means of the residual surfactant concentration over time by DOC measurements. Duplicate DOC measurements for each sample were made. It is known that sorption may significantly influence the resulting environmental effects of surfactants and this fact has been studied by some authors [20, 21]. In the biodegradation assays presented here, the sorption could be considered negligible, given the scant biomass formation. Abiotic assays were made in the presence of $HgCl_2$ to confirm this, and it was found that the values of the residual surfactant remained around 100% over the biodegradation period. These results indicate that the contribution of abiotic processes to the degradation of the surfactants in the biodegradation tests can be dismissed.

For the surfactant EC-$R_{12\text{-}14}E_{10}$, Figure 1 shows the surface-tension data vs surfactant concentration. The surface-tension data plotted on a semi-log plot for a surfactant will

have an approximately linear drop in surface tension followed by a plateau. The concentration at which this discontinuous change in slope occurs is the CMC. CMC data for the ether carboxylic derivative surfactants are shown in Table 1.

Figure 1

Figure 2 shows the time course of the ultimate biodegradation of the surfactants over the degradation period. The initial concentrations in the assays were 25, and 50 mg/L.

Figure 2

For the comparison and quantification of the different biodegradation assays, the characteristic parameters of the biodegradation profiles were evaluated [22]: half-life ($t_{1/2}$), mean biodegradation rate (V_M), and the residual-surfactant concentration at the end of the assay (S_R), which is calculated with the final DOC measurements average. $t_{1/2}$ is the time for which the substrate concentration diminishes to half from the beginning of the biodegradation process. The half-life is calculated by graphic methods on the biodegradation profile. V_M has been defined as the mean velocity of biodegradation reached until achieving 50% biodegradation of the surfactant, and it has been calculated as the quotient between the percentage of biodegradation reached and the time needed to reach this biodegradation value. This parameter provides the speed of the biodegradation process.

Table 1 shows the characteristic parameters of the biodegradation profiles for the ether carboxylic derivative surfactants for all the concentrations assayed. S_0 is the initial concentration of the biodegradation assay in mg/L and Min is the final % of mineralization reached at the end of the assay calculated with this expression:

$$Min(\%) = \frac{[TOC]_i - [TOC]_f}{[TOC]_i} \cdot 100 \qquad \text{Eq. (1)}$$

Table 1

An analysis of the influence of the initial concentration is presented in Table 1, reflecting that biodegradation process is slower when the initial concentration increases, which is the half-life increases and the mean biodegradation rate decreases. This may be due to the long adaptation time needed by the microorganisms for these surfactants, which are generally not included in conventional detergent formulas.

For the EC-$R_{12\text{-}14}E_3$ and EC-$R_{12\text{-}14}E_{10}$, the residual-surfactant concentration at the end of the assay, S_R, notably augmented with the increasing surfactant concentration. However, for EC-R_8E_8, the one with the shortest alkyl chain and the highest CMC, the

residual surfactant concentration was independent of the initial concentration, and the mineralization percentage rises with the initial concentration.

Current legislation requires a minimum level of 60% of ultimate biodegradation to be reached when applying one of the methods listed in Annex III of Regulation (EC) No 648/2004 [23]. If this condition is met the surfactant can be considered biodegradable. The surfactant EC-R_8E_8 fulfils this requirement, yielding 91.9% DOC removal. The surfactants with greater alkyl-chain lengths (EC-$R_{12\text{-}14}E_3$ and EC-$R_{12\text{-}14}E_{10}$) satisfy this requirement only with an initial surfactant concentration of 25 mg/L (62.13 and 81.42% DOC removal, respectively).

To analyse the influence of the degree of ethoxylation and the size of the alkyl chain on the final biodegradation process, the results for different surfactants at the initial concentrations of 25 mg/L and 50 mg/L are compared (Figure 3).

The surfactant that achieved the greatest biodegradation was EC-R_8E_8, with the shortest alkyl chain. In comparisons of the surfactants with the same alkyl length, EC-$R_{12\text{-}14}E_3$ and EC-$R_{12\text{-}14}E_{10}$, (C12-C14) and different degree of ethoxylation (3 and 10, respectively), it was found that there was no significant differences.

Figure 3

3.2. Toxicity of ether carboxylic derivates surfactants

The toxicity of the ether carboxylic derivative surfactants, and their binary mixtures, was measured. Toxicity values of the surfactants were determined by applying the 24-h immobilization test with *Daphnia magna*, the LumiStox® 300 test which employs the luminescent bacteria *Photobacterium phosphoreum* and the 72-h algal growth-inhibition test. These results show that *Vibrio fischeri*, *Daphnia magna* and Microalgae do not use the surfactants as sources of carbon. Therefore, the surfactant concentrations remained stable over the time period used in the bioassays. Table 2 shows the toxicity values for the tests with *Vibrio fischeri*, *Daphnia magna* and Microalgae, for the different surfactants assayed.

Table 2

The acute toxicity values of the surfactants ranged from 3.58 mg/L to 7.08 mg/L for the surfactant EC-$R_{12\text{-}14}E_3$, from 14.18 mg/L to 26.01 mg/L for the EC-$R_{12\text{-}14}E_{10}$ and from 76.26 mg/L to 134.59 mg/L for the EC-R_8E_8. According to the European Union Directive No. 67/548/EEC [24] with the respective amendment No. 7, the above results classify the surfactant EC-$R_{12\text{-}14}E_3$ as the second toxicity class (R51), which is regarded as toxic against aquatic organisms. Meanwhile, the surfactants EC-$R_{12\text{-}14}E_{10}$ and EC-

R_8E_8 are classified as harmful (third toxicity class (R52)) and safe, respectively. According to the literature, anionic and non-ionic surfactants are toxic to various aquatic organisms at the concentrations from 0.0025 to 300 mg/L and from 0.3 to 200 mg/L, respectively [25]

For ecological safety, it is further assumed that the theoretically calculated concentration of surfactant in the natural environment should be 100-fold lower than the values of IC_{50} and EC_{5O} determined experimentally. In this case, no negative environmental impact of the surfactant would be expected. The results of the toxicity tests are typically much higher compared to values that might be found in the environment [26].

The results presented in Table 2 show that *Vibrio fischeri* was more sensitive to toxic effects from ether carboxylic derivative surfactants than was *Daphnia magna* and Microalgae. The toxicity is lower for the surfactant with the shortest alkyl chain. The degree of ethoxylation (E) has the reverse effect: the higher degree of ethoxylation the smaller toxicity.

Surfactants are often used as co-surfactants in detergent formulas, so the toxicological interactions of the binary mixtures of ether carboxylic derivative surfactants have been investigated. The results presented in Table 2 shows that *Daphnia magna* was more sensitive to toxic effects from binary mixtures of ether carboxylic derivative surfactants than was *Vibrio fischeri* and Microalgae. Microalgae were less sensitive to toxic effects from binary mixtures of ether carboxylic derivative surfactants than the individual surfactants. The less toxic mixture is formed by the surfactants having lower individual toxicity, surfactants EC-R_8E_8 and EC-$R_{12\text{-}14}E_{10}$. This result highlights the synergism in the co-occurrence of this class of surfactants.

Comparisons of the toxicity of these surfactants with the typical anionic surfactant LAS show that when *Vibrio fischeri* and *Daphnia magna* test are used, LAS toxicity values are intermediate between the ether carboxylic derivative surfactants assayed. The Microalgae test indicates that LAS is the least toxic surfactant, although the synergic binary mixtures improve these surfactants results, and consequently the mixture between EC-R_8N_8 and EC-$_{R12\text{-}14}N_{10}$ proves less toxic than LAS.

In conclusion, ether carboxylic derivative surfactants can be considered biodegradable. The one with the shortest alkyl chain length and the highest CMC (EC-R_8E_8) yielded

the highest % of mineralization. The influence of the initial concentration reflected that the biodegradation process was slower when the initial concentration increased, the half-life increased, the mean biodegradation rate decreased, and the residual surfactant concentration notably augmented, except for EC-R_8E_8, for which the S_R was independent of the initial concentration. The toxicity measurements of these ether carboxylic surfactants indicate that the least toxic was the most biodegradable (EC-R_8E_8). Binary mixtures measurements indicate that the least toxic mixture was formed by the surfactant having lower individual toxicity. Moreover, the Microalgae test results indicate that there was synergism in the co-occurrence of these surfactants. The results imply that these surfactants at low concentrations may be considered less damaging to the environment.

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BIOGRAPHY

Dr. E. Jurado was born in 1951 and graduated from the University of Granada in 1975. She received her Ph.D. in 1980 and became full professor of Chemical Engineering in the Science Faculty at the same University in 1996. Her main research areas are: enzymes, kinetic enzymatic, biodegradation of surfactants, emulsions, and the physical chemistry and applications of surfactants. At present, she is head of the Chemical Engineering Department at the University of Granada. She has published over 60 papers in different fields.

Dr. M Fernández-Serrano was born in 1967 and studied chemistry at the University of Granada. She was awarded a 4-year fellowship at the same University and received her PhD in 1995. She became an associate professor at the University of Granada in 1999. Her research activities include searching for new groups of biodegradable surfactants.

Dr. M. Lechuga was born in 1976. She graduated from the University of Granada in Chemical Engineering and then worked on an investigation project entitled “Formulation of Liquid Detergents Specifically for the Industrial Agrofood and Hotel Sector” directed by E. Jurado. She received her PhD in 2005 and is currently an associate professor at the University of Granada. Her research interests include surfactants and their applications.

F. Ríos, born in 1985, graduated in Chemical Engineering at the University of Granada in 2009. Currently, he works on a project entitled “Environmental Impact of commercial surfactants” for his Ph.D.

Table 1. Characteristic parameters of the biodegradation profiles for ether carboxylic derivative surfactants

EC-R_8E_8

(*Active Matter: 89 %; CMC: 243.4 mg/L)

S_0, mg/L	$t_{1/2}$, days	V_M, %/day	S_R, mg/L	Min, %
25	5.11	11.52	2.03	91.88
50	8.12	6.96	3.20	91.96

EC-$R_{12\text{-}14}E_3$

(*Active Matter: 94 %; CMC: 33.24 mg/L)

S_0, mg/L	$t_{1/2}$, days	V_M, %/day	S_R, mg/L	Min, %
25	5.23	12.72	7.38	62.13
50	7.47	8.64	16.79	52.28

EC-$R_{12\text{-}14}E_{10}$

(*Active Matter: 94 %; CMC: 70.8 mg/L)

S_0, mg/L	$t_{1/2}$, days	V_M, %/day	S_R, mg/L	Min, %
25	7.56	6.96	3.42	81.42
50	19.46	2.40	19.64	51.33

*The % of active matter is supplied by the manufacturer

$t_{1/2}$: half-life

V_M: mean biodegradation rate

S_R: residual-surfactant concentration

Min: % of mineralization

The surfactant concentration was measured using a TOC analyzer

Table 2. Toxicity values (95% CI) for the tests with *Vibrio fischeri*, *Daphnia magna* and Microalgae

	Vibrio fischeri	*Daphnia magna*	Microalgae
Surfactants	EC_{50} (15 min), mg/L	IC_{50}, mg/L	EC_{50}, mg/L
EC-$R_{12\text{-}14}E_3$	3.58 (3.19-3.97)	3.47 (2.81-4.14)	7.08 (5.08-9.08)
EC-$R_{12\text{-}14}E_{10}$	14.18 (11.35-17.02)	18.74 (17.27-20.21)	26.01 (19.38-32.64)
EC-R_8E_8	134.59 (125.26-143.93)	120.95 (93.35-148.55)	76.26 (64.85-87.67)
EC-$R_{12\text{-}14}E_3$ + EC-R_8E_8	14.96 (9.69-20.23)	8.04 (5.81-10.27)	78.14 (67.62-88.66)
EC-$R_{12\text{-}14}E_3$ + EC-$R_{12\text{-}14}E_{10}$	17.04 (13.50-20.57)	5.04 (4.57-5.51)	29.02 (24.38-33.66)
EC-R_8E_8 + EC-$R_{12\text{-}14}E_{10}$	54.70 (46.90-62.49)	39.31 (33.34-45.28)	166.57 (149.93-183.21)
LAS	27.58 (26.26-28.90)	10.09 (9.22-10.96)	151.07 (143.09-159.05)

Figure Captions

Figure 1. Surface tension data versus surfactant concentration for the surfactant EC-$R_{12-14}E_{10}$. Temperature= 25ºC

Figure 2. Time course of ultimate biodegradation over the degradation period. a) EC-R_8E_8, b) EC-$R_{12-14}E_3$, c) EC-$R_{12-14}E_{10}$

Figure 3. Effect of the surfactant structure on the ultimate biodegradation. a) 25 mg/L, b) 50 mg/L

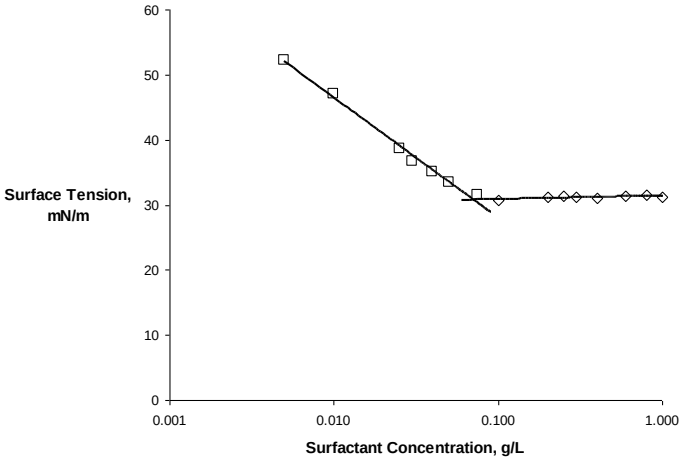

Figure 1

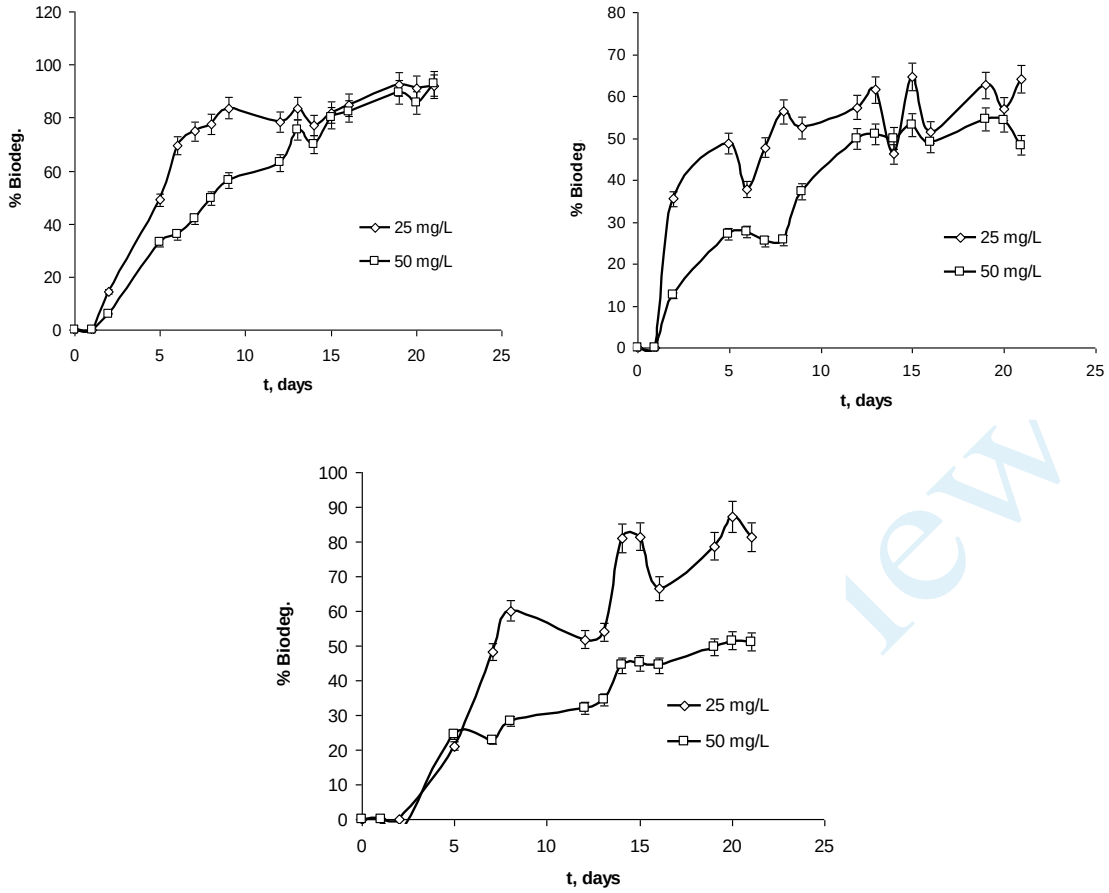

Figure 2

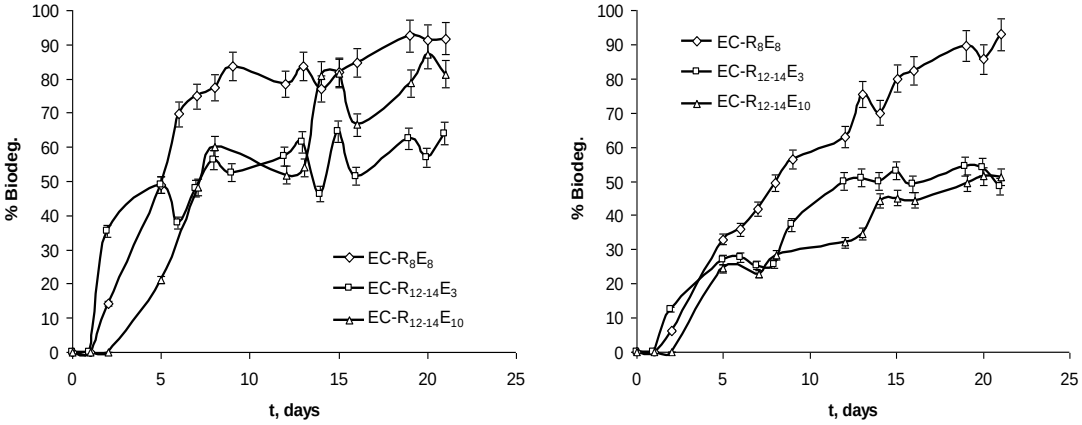

Figure 3