

Review

Antioxidant Functions of Vitamin D and CYP11A1-Derived Vitamin D, Tachysterol, and Lumisterol Metabolites: Mechanisms, Clinical Implications, and Future Directions

Héctor Vázquez-Lorente 1,2,† [,](https://orcid.org/0000-0003-0183-6902) Lourdes Herrera-Quintana 1,2,† [,](https://orcid.org/0000-0003-1407-322X) Laura Jiménez-Sánchez 1,2,3 , Beatriz Fernández-Perea ⁴ and Julio Plaza-Diaz 3,5,6,[*](https://orcid.org/0000-0002-5171-9408)

- ¹ Department of Physiology, Schools of Pharmacy and Medicine, University of Granada, 18071 Granada, Spain; hectorvazquez@ugr.es (H.V.-L.); lourdesherrera@ugr.es (L.H.-Q.); laurajimsan@gmail.com (L.J.-S.)
- ² Biomedical Research Center, Health Sciences Technology Park, University of Granada, 18016 Granada, Spain 3 Instituto de Investigación Biosanitaria IBS.GRANADA, Complejo Hospitalario Universitario de Granada, 18014 Granada, Spain
- 4 Immunology and Clinical Analysis Service, Virgen de las Nieves University Hospital, 18014 Granada, Spain; beafdezperea@correo.ugr.es
- ⁵ Department of Biochemistry and Molecular Biology II, School of Pharmacy, University of Granada, Campus de Cartuja s/n, 18071 Granada, Spain
- ⁶ Children's Hospital of Eastern Ontario Research Institute, Ottawa, ON K1H 8L1, Canada
- ***** Correspondence: jrplaza@ugr.es; Tel.: +34-958241000 (ext. 41599)
- These authors contributed equally to this work.

Abstract: Evidence is increasing that vitamin D and CYP11A1-derived vitamin D, tachysterol, and lumisterol metabolites play a significant antioxidant role beyond its classical functions in bone health and calcium metabolism. Several recent studies have linked these elements to reduced oxidative stress as well as improved immune, cardiovascular, and neurological functions as a result of chronic kidney disease and cancer. Additionally, supplementation with this vitamin has been shown to be one of the most cost-effective micronutrient interventions worldwide, highlighting its potential as a therapeutic approach. The underlying mechanisms and implications of this antioxidant function of vitamin D or CYP11A1-derived vitamin D, tachysterol, and lumisterol metabolites are not well understood. This comprehensive and narrative review is aimed at summarizing the current evidence regarding the molecular mechanisms implicated in this antioxidant function of vitamin D, as well as to provide a general overview and to identify key research areas for the future, offering an extensive perspective that can guide both researchers and clinicians in the management of diseases associated with oxidative stress and/or insufficient vitamin D status.

Keywords: vitamin D; antioxidant; oxidative stress; reactive oxygen species; neurodegenerative diseases

1. Introduction

Vitamin D, historically considered an essential nutrient derived from various dietary sources, is now defined as a steroid hormone. Its precursor molecule is synthesized in the skin when exposed to ultraviolet radiation (UVR) [\[1\]](#page-11-0), which further generates electrical, chemical, and biological signals that are transmitted to the brain, endocrine, and immune systems, as well as other central organs. These systems work in concert to regulate body homeostasis, thereby providing a photo-neuro-immuno-endocrinological perspective [\[2\]](#page-11-1). Among its classical functions, vitamin D has been recognized as an important factor in the regulation of neuro-musculoskeletal activities, tightly controlling circulating Ca^{2+} levels. However, in recent years, numerous other physiological processes have been found to be influenced by vitamin D, with significant implications in the prevention and reduction of severity of various conditions such as autoimmune, cardiovascular, and metabolic

Citation: Vázquez-Lorente, H.; Herrera-Quintana, L.; Jiménez-Sánchez, L.; Fernández-Perea, B.; Plaza-Diaz, J. Antioxidant Functions of Vitamin D and CYP11A1-Derived Vitamin D, Tachysterol, and Lumisterol Metabolites: Mechanisms, Clinical Implications, and Future Directions. *Antioxidants* **2024**, *13*, 996. [https://doi.org/10.3390/](https://doi.org/10.3390/antiox13080996) [antiox13080996](https://doi.org/10.3390/antiox13080996)

Academic Editor: Alessandra Napolitano

Received: 31 July 2024 Revised: 11 August 2024 Accepted: 17 August 2024 Published: 17 August 2024

Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license [\(https://](https://creativecommons.org/licenses/by/4.0/) [creativecommons.org/licenses/by/](https://creativecommons.org/licenses/by/4.0/) $4.0/$).

disorders $[1,3]$ $[1,3]$. Due to the pleiotropic vitamin D effects and the high prevalence of its insufficient status worldwide, supplementation with this vitamin has been shown to be one of the most cost-effective micronutrient interventions [\[4\]](#page-11-3). Furthermore, among its various beneficial effects, vitamin D has been observed to reduce levels of oxidative stress (OS) parameters in different diseases, such as polycystic ovary syndrome [\[5\]](#page-11-4) and mild cognitive impairment [\[6\]](#page-11-5).

As a result of reactive oxygen species (ROS), reactive nitrogen species (RNS), and other free radicals, OS contributes to the onset and progression of various diseases, such as diabetes, obesity, neurological disorders, and cardiovascular conditions [\[7\]](#page-11-6). The imbalance between pro-oxidant and antioxidant molecules leads to cellular damage, which manifests as inflammation, DNA damage, protein cross-linking, lipid membrane peroxidation, mitochondrial dysfunction, and necrosis/apoptosis [\[8\]](#page-11-7). Therefore, maintaining an adequate concentration of prooxidants and antioxidants, which constitute the redox systems, is crucial. Disruption of this equilibrium can contribute to the development of pathological conditions such as cancers and chronic diseases [\[9\]](#page-11-8). In this context, the use of pharmacological antioxidants for health protection has emerged as a promising therapeutic target in recent decades. Consequently, dietary recommendations emphasizing the consumption of plant foods rich in antioxidants and the use of antioxidant supplements have garnered increasing interest [\[10\]](#page-11-9). Regarding vitamin D supplementation and its effect on antioxidant status, while several studies indicate that vitamin D decreases OS parameters, the results in human studies remain controversial. Thus, the antioxidant function of vitamin D and its underlying mechanisms are not yet well established [\[11\]](#page-11-10).

Based on the above considerations, the aim of the present study is to review the current evidence regarding the main antioxidant functions of vitamin D and CYP11A1 derived vitamin D, tachysterol, and lumisterol metabolites, and their role in OS. This review provides novel insights into its mechanisms, clinical implications, and future perspectives.

2. Oxidative Stress and Antioxidant Status

OS is defined as a dysregulation between the production of RNS and ROS, and the endogenous antioxidant defense mechanisms, collectively known as the redox state [\[12\]](#page-11-11). OS indicates a disparity between ROS generation and the organism's capacity to detoxify reactive intermediates or repair the resulting damage through an adequate antioxidant defense system. High levels of ROS can damage nucleic acids, proteins, membranes, nucleic acids, organelles, and lipids, eventually leading to irreversible cell damage and death [\[13\]](#page-11-12). This macromolecular damage is often considered in terms of oxidative mechanisms linked to free radicals, which are reactive and can initiate chain reactions that propagate to damage multiple molecules [\[14\]](#page-11-13). Free radicals or active oxidants, such as superoxide radicals (O2^{•−}), hydroxyl radicals ([•]OH), and hydrogen peroxide (H₂O₂), which are originated from oxygen, are collectively termed ROS [\[15\]](#page-11-14). It is widely accepted that the principal source of free radicals in the cell is the mitochondrial respiratory chain [\[16\]](#page-12-0). The production of reactive species can result from biochemical processes, exposure to damaging agents (e.g., environmental pollutants and radiation), or limited capabilities of endogenous antioxidant systems [\[17\]](#page-12-1).

ROS are implicated in various biological functions, including the regulation of physiological processes and the pathogenesis of numerous human diseases [\[18\]](#page-12-2). Under normal conditions, there is a balance between oxidant and antioxidant activity due to the action of the antioxidant defense system. RNS and ROS are neutralized by both extrinsic and intrinsic methods. Intrinsic mechanisms mainly involve enzymatic processes [\[19\]](#page-12-3), including antioxidant enzymes such as superoxide dismutase (SOD), peroxidases, and glutathione reductase (GR). These enzymes are distributed through the cell to protect DNA integrity, hormones, enzymes, cell membranes, and other organelles and structures from ROS-induced damage [\[20](#page-12-4)[,21\]](#page-12-5).

3. Background and Significance of Vitamin D in Human Health

3.1. Sources, Synthesis, and Metabolism

Vitamin D is a fat-soluble secosteroid that can be obtained exogenously through diet or supplements. The following foods are good sources of vitamin D: fatty fish (such as salmon, mackerel, and sardines), fish liver oils, fortified foods (such as milk, orange juice, cereals, and egg yolks), and fortified foods (such as milk, orange juice, and cereals [\[22\]](#page-12-6). Vitamin D supplements are also an effective means of ensuring adequate intake, particularly for individuals at deficiency risk; in particular, those with limited sun exposure, the elderly, and people with darker skin should avoid excessive exposure to the sun [\[23\]](#page-12-7).

Endogenously, pre-vitamin D_3 can be synthesized in the skin following the absorption of ultraviolet-B (UVB) energy by 7-dehydrocholesterol (7DHC) [\[24](#page-12-8)[,25\]](#page-12-9). Prolonged exposure of 7-DHC to UVB results in the phototransformation of pre-vitamin D_3 into tachysterol and lumisterol [\[26\]](#page-12-10). Specifically, in the skin, the absorption of UVB energy leads to the breakage of the unsaturated B ring in 7DHC, causing its transformation into corresponding secosteroids, with or without a full-length side chain. This results in the formation of vitamin D-like, tachysterol-like, and lumisterol-like compounds when the broken B ring reseals in a different stereochemical configuration [\[27\]](#page-12-11). Although the role of vitamin D_3 as a prohormone is well-established, tachysterol and lumisterol have been reported to exhibit similar functions [\[28\]](#page-12-12).

Vitamin D activation occurs through two distinct pathways: the canonical and noncanonical mechanisms. Concerning the canonical pathway [\[29\]](#page-12-13), vitamin D, in its inactive form, undergoes specific metabolic processes to become active and is transported to the liver, bound to vitamin D-binding proteins, where it undergoes hydroxylation to form 25-hydroxyvitamin D. This metabolite is used to assess a person's vitamin D status [\[30\]](#page-12-14). Various tissues, including prostate, pancreatic islets, breast, colon, malignant and immune cells, macrophages, vascular smooth muscle cells, and especially the kidneys, possess $1-\alpha$ hydroxylase [\[31\]](#page-12-15). In these tissues, 25-hydroxyvitamin D undergoes a second hydroxylation to produce 1,25-dihydroxycholecalciferol, the physiologically active form of vitamin D. This active form exerts its effects by interacting with vitamin D receptors (VDR) [\[32\]](#page-12-16), which then enter the nucleus and heterodimerize with the retinoic acid X receptor [\[33\]](#page-12-17). This receptor complex acts on vitamin D-response elements to exert transcriptional control over a set of genes [\[34\]](#page-12-18), thereby increasing the transcription of vitamin D-dependent genes, which are essential for its various functions.

The recently identified as non-canonical pathways of vitamin D activation are initiated by CYP11A1, an essential enzyme in steroidogenesis present in various organs and tissues, including the skin, peripheral tissues, adrenal glands, placenta, gastrointestinal tract, and diverse cancer cells [\[35\]](#page-12-19). CYP11A1, the first enzyme in steroidogenesis or the non-canonical pathway, is responsible for hydroxylating and cleaving the side chain of cholesterol to produce pregnenolone [\[36\]](#page-12-20). This enzyme also has the capability to hydroxylate and cleave the side chain of 7DHC and hydroxylate the side chains of vitamin D_3 and D_2 without cleavage, involving a complex process with several cytochrome P450 (CYP) enzymes and generating numerous hydroxyderivatives. Additionally, CYP11A1 can activate tachysterol and lumisterol through sequential hydroxylation, resulting in multiple hydroxylated forms [\[37](#page-12-21)[–39\]](#page-12-22). By binding to the VDR genomic site, these compounds can modulate gene expression like 1,25-dihydroxyvitamin D_3 . Furthermore, hydroxyderivatives of vitamin D_3 , including those derived from tachysterol and lumisterol, interact with other nuclear receptors such as the aryl hydrocarbon receptor (AhR), retinoic acid orphan receptors (ROR) α and γ, and liver X receptors (LXR) α and β , thereby influencing their expression and activity [\[40\]](#page-12-23). Vitamin D sources, synthesis, and metabolism are summarized in Figure [1.](#page-3-0)

Figure 1. Vitamin D sources, synthesis, metabolism, and antioxidant mechanisms. Abbreviations. **Figure 1.** Vitamin D sources, synthesis, metabolism, and antioxidant mechanisms. Abbreviations. UV, ultraviolet. UV, ultraviolet.

3.2. Vitamin D Deficiency and Its Implications 3.2. Vitamin D Deficiency and Its Implications

Vitamin D deficiency has emerged as a significant global health concern, affecting Vitamin D deficiency has emerged as a significant global health concern, affecting approximately 1 billion individuals worldwide and presenting substantial health risks approximately 1 billion individuals worldwide and presenting substantial health risks [\[41\]](#page-12-24).
The contract of the contract of the substantial health risks [41]. The prevalence of vitamin D deficiency is estimated to range between 60% and 80% , with even higher rates observed among the elderly population [\[42\]](#page-13-0). The circulating concentration of 25-hydroxyvitamin D serves as a comprehensive indicator of overall vitamin D status, as it reflects the levels derived from various sources and has a half-life of approximately two months, making it the most reliable measure of vitamin D levels in the body [\[43\]](#page-13-1). Several factors contribute to the risk of 25-hydroxyvitamin D deficiency, including race, elevated body mass index, seasonal variations (particularly during winter months with reduced daylight), residence in higher latitudes, and dietary insufficiencies in vitamin D [\[44\]](#page-13-2). Notably, obesity is associated with lower circulating levels of vitamin D due to factors such as volumetric dilution and sequestration in adipose tissue [\[45\]](#page-13-3). Consequently, $\frac{1}{2}$ conditions $\frac{1}{2}$. $\$ vitamin D deficiency is linked to an increased risk of numerous diseases and clinical

vitamin D deficiency is linked to an increased risk of numerous diseases and clinical ing of vitamin D's potential roles in various biological processes [47]. Vitamin D is known conditions [\[46\]](#page-13-4).

The identification of VDRs in virtually all human cells has expanded our understand-ing of vitamin D's potential roles in various biological processes [\[47\]](#page-13-5). Vitamin D is known to regulate the expression of over 1000 genes in the human genome, accounting for approx-imately 5% of all protein-coding genes [\[48\]](#page-13-6). Its active form acts as a potent inhibitor of cell proliferation while promoting cell differentiation and apoptosis in cancer cells [49]. VDRs are not confined to tissues involved in bone metabolism but are also present in numerous other tissues, including the brain, prostate, breasts, and immune cells. This widespread distribution of receptors associates vitamin D with a range of chronic diseases [\[50\]](#page-13-8). Vitamin D deficiency has been implicated in a variety of clinical conditions. For example, it is linked glucose metabolism and endocrease metabolism and endocrease conditions. The endingless is and endocreased to cardiovascular disease by diminishing pro-inflammatory cytokines production [\[51\]](#page-13-9), type t_{total} and t_{total} a 2 diabetes through its role in stimulating insulin production and regulating genes involved in glucose metabolism [\[52\]](#page-13-10), neuropsychiatric disorders, and endocrinopathies [\[53\]](#page-13-11). Additionally, it affects tumor development and immune diseases by modulating the number of embryonic hematopoietic stem cells [\[54\]](#page-13-12), influencing the differentiation of myeloid progenitor cells into monocytes and granulocytes [\[55\]](#page-13-13), and affecting the differentiation of monocytes into dendritic cells and macrophages [\[56\]](#page-13-14). Furthermore, vitamin D has recently

been recognized for its antioxidant properties [\[57\]](#page-13-15). Similar to 1,25-dihydroxyvitamin D_3 , the principal hormonally active form of vitamin D_3 , hydroxymetabolites derived from CYP11A1 also demonstrate anticancer properties, including antiproliferative and prodifferentiation effects, as well as immunomodulatory functions [\[58\]](#page-13-16). Additionally, these CYP11A1-derived vitamin D_3 hydroxyderivatives have been found to offer protection against skin damage by mitigating UVB-induced DNA damage and reducing levels of ROS [\[59\]](#page-13-17).

4. Antioxidant Functions of Vitamin D

In 1993, Helen Wiseman demonstrated that vitamin D is an antioxidant vitamin [\[60\]](#page-13-18). Since then, many more studies have been conducted, and vitamin D mechanisms have become increasingly clear [\[61\]](#page-13-19). Namely, vitamin D is known to enhance antioxidant gene expression via binding to the vitamin D response element [\[62\]](#page-13-20). Specifically, vitamin D deactivates the nuclear transcription factor kB (NF-KB) by increasing the expression of IkB and decreasing the phosphorylation of IkB- α . It suppresses the expression of the nicotinamide adenine dinucleotide phosphate (NADP) enzyme and prevents the accumulation of advanced glycation end-products, which may contribute to the suppression of OS [\[63\]](#page-13-21). However, one of the main mechanisms that explain vitamin D effects on antioxidant enzymes is the indirect regulation of the expression of nuclear factor erythroid 2-related factor 2 (Nrf2)-regulating also the expression of other relevant genes to the ROS signaling pathway function, such as klotho [\[64\]](#page-13-22), which in turn increases the production of antioxidant enzymes [\[65\]](#page-13-23). Mainly, the Nrf2 induces the expression of vitagenes, which are cytoprotective genes [\[66\]](#page-14-0); these are essential for cellular defense mechanisms such as redox equilibrium and detoxification. Furthermore, these proteins control the expression of a variety of proteins, including heat-shock proteins, which have been shown to enhance cytoprotection in a number of diseases and processes, including cancer, inflammation, neurodegenerative diseases, and aging. [\[67\]](#page-14-1). Consequently, vitamin D has been seen to affect different parameters related to antioxidant status, thus impacting on the total antioxidant capacity (TAC) [\[68\]](#page-14-2). In this context, vitamin D scavenges physiologically relevant free radicals by increasing SOD levels which react with oxygen radicals [\[69\]](#page-14-3). Furthermore, an increase in the intracellular pool of reduced GSH has been also observed, partially through upstream regulation of the GR and glutamate-cysteine ligase (GCL) genes, GCL being a vital enzyme in GSH production [\[70\]](#page-14-4). The expression of thioredoxin reductase 1 (TXNRD1) is also induced by vitamin D. As well as reducing thioredoxin, TXNRD1 reduces glucose-6-phosphate dehydrogenase (G6PD), which generates NADPH to regenerate glutathione (GSH) [\[71\]](#page-14-5). These functions of vitamin D may be observed in different cells. For instance, vitamin D exerts an antioxidant effect on monocytes by upregulating GR and GCL, thereby reducing oxygen radicals, including ROS [\[72\]](#page-14-6).

Additionally, vitamin D appears to play an essential role in protecting mitochondria and preventing lipid peroxidation through the regulation of inflammatory processes, ROS concentrations, and mitochondrial antioxidant secretion through the cell signaling pathway [\[70\]](#page-14-4). ROS generation may be boosted by changes in mitochondrial energy metabolism during vitamin D deficiency, along with decreased antioxidant defenses [\[73\]](#page-14-7). Moreover, hyperglycemia disturbs electron transfer and mitochondria activation, causing excessive ROS production via the polyol pathway [\[51\]](#page-13-9). Having a suboptimal vitamin D status fails to subdue OS conditions, augmenting intracellular oxidative damage and the apoptosis rate. Indeed, intracellular Nrf2 is inversely correlated with mitochondrial ROS accumulation and OS escalation [\[71\]](#page-14-5). In addition to improving OS, cytokine production, and proinflammatory status, vitamin D can normalize mitochondrial dynamics [\[74\]](#page-14-8). A further benefit of vitamin D is that it inhibits lipid peroxidation and exerts antioxidant effects through the alteration of antioxidant enzymes in the cell membrane [\[51\]](#page-13-9). Malondialdehyde (MDA) levels, which are considered a marker of lipid peroxidation, could be lowered via vitamin D supplementation [\[75\]](#page-14-9). As a result of their structural similarity, vitamin D and cholesterol, as well as ergosterol, are known to possess antioxidant properties in the membrane [\[60\]](#page-13-18). Consequently, vitamin D could serve as an antioxidant for the cellular membrane as it Euroepartity, vitaming the case of the distribution of the choles-
inhibits the oxidation of lipids induced by iron in brain liposomes [\[76\]](#page-14-10).

Antioxidants **2024**, *13*, 996 6 of 19

Hydroxymetabolites of vitamin D_3 derived from CYP11A1, such as 20-hydroxyvitamin D_3 derived from CYP11A1, such as 20-hydroxyvitamin D_3 (20(OH) D_3) and 20,23-dihydroxyvitamin D_3 (20,23(OH)₂D₃), have been observed to exhibit anti-oxidative properties comparable to those of the classical active form of vitamin D_3 . The anti-oxidative effects of active vitamin D forms can be mediated through the activation of NRF2 and p53 signaling pathways, involving interaction with the VDR and inverse agonism on ROR γ , which subsequently attenuates Th17 responses [29]. Consistent with this, CYP11A1-derived vitamin D_3 hydroxyderivatives have been shown to protect cultured human keratinocytes and melanocytes from UVB-induced damage. This protection is characterized by reduced production of ROS, H_2O_2 , and nitric oxide (NO), as well as an increased expression of genes encoding enzymes involved in OS defense [\[77\]](#page-14-11). Furthermore, hydroxylated forms of lumisterol have been demonstrated to induce the Furthermore, hydroxylated forms of lumisterol have been demonstrated to induce the expression of several genes associated with protection against OS [\[35\]](#page-12-19). If y dioxyfie tabolites of vitamin D₃ defived none channel as 20-hydroxyvi as a condition of general expression of the condition of the condition of \mathbb{R}^2 .

Recently, vitamin D has garnered increased attention as a non-enzymatic antioxidant compound. Vitamin D exerts its antioxidant effects through several mechanisms, including the inhibition of free radical generation by nitric oxide synthase (NOS) and gamma-glutamyl transpeptidase (GGT) [\[78\]](#page-14-12). Nevertheless, there is variability in the literature concerning the antioxidant effects of vitamin D. A systematic review of clinical trials involving non-pregnant individuals indicated that beneficial effects on OS parameters, such as reductions in MDA levels and increases in GSH, were observed only at high doses of 100,000-200,000 IU per month [\[79\]](#page-14-13). While many studies report a reduction in MDA levels with oral vitamin D supplementation, the findings are inconsistent, with some studies fail-ing to demonstrate positive effects [\[80\]](#page-14-14). These discrepancies may be attributed to variations ing to demonstrate positive effects [80]. These discrepancies may be attributed to variations in the study design, participant characteristics, vitamin D dosage, administration route, and study duration [\[81\]](#page-14-15). The principal antioxidant mechanisms of vitamin D are reviewed
. in Figure [2.](#page-5-0) ations in the study design, participant characteristics, vitamin D dosage, administration

Figure 2. Antioxidant functions of vitamin D. Abbreviations: MDA, Malondialdehyde; Nrf2, nuclear factor erythroid 2-related factor 2; •OH, hydroxyl radicals; •O2⁻, superoxide radicals; •O2²⁻, hydrogen peroxide; ROS, reactive oxygen species.

4.1. Evidence from Clinical and Experimental Studies 4.1.1. Cell Culture Studies and Animal Models

It has been documented that vitamin D has a substantial protective role against cellular stress and may function as an antioxidant in different cell types like hepatocytes [\[82\]](#page-14-16). Vitamin D has also demonstrated antioxidant properties, notably in protecting cardiomyocytes from toxicity induced by aluminum phosphide [\[83\]](#page-14-17). Additionally, it has been shown to downregulate intracellular adhesion molecule-1 (ICAM-1) expression in peripheral blood mononuclear cells following exposure to tumor necrosis factor-alpha (TNF- α). In monocytes, vitamin D is able to reduce intracellular adhesion molecule (ICAM-1) expression after TNF- α exposure [\[84\]](#page-14-18), and improves antioxidant properties in these cells by decreasing lipid peroxidation and increasing oxidative capacity in monocytes [\[60,](#page-13-18)[85\]](#page-14-19). Furthermore, by mitigating OS in endothelial cells, vitamin D may contribute to the inhibition of atherosclerosis [\[86\]](#page-14-20). In fact, supplementation with vitamin D has been related to a significant reduction in oxidized lipid markers.

Studies conducted on animal models have investigated the antioxidant properties of vitamin D. In this line, using a rat model of chronic immobilization-induced cardiac dysfunction, supplementation with vitamin D resulted in a significant dose-dependent reduction in OS, which was achieved through an increase in the activity of antioxidant enzymes improving directly the mitochondrial function [\[87\]](#page-14-21). In a mouse model of triple-negative breast cancer, vitamin D supplementation mitigated doxorubicin-induced cardiotoxicity by reducing ROS and mitochondrial damage, while preserving the anticancer efficacy of doxorubicin [\[88\]](#page-14-22). Additionally, vitamin D has been shown to offer protection against dextran sulfate sodium-induced colitis by enhancing the activity of 3-hydroxy-3-methylglutaryl-CoA synthase 2, an enzyme implicated in vitamin D-mediated defense against OS and inflammation. These findings suggest that vitamin D may provide a potential therapeutic strategy for alleviating OS and inflammation associated with colitis [\[89\]](#page-15-0).

On the other hand, different studies have pointed out the potential of vitamin D supplementation for improving food production. In dairy cows, where there are critical moments that produce an increase in OS that can trigger a pathology, such as the change from lactation to feed, the concentration of vitamin D correlated with antioxidant potential [\[90\]](#page-15-1). Likewise, although the administration of a regular dosage of 2000 IU of vitamin D within the diet of weaned piglets did not achieve optimal antioxidant capacity or immune function [\[91\]](#page-15-2), vitamin D supplementation in growing-finishing pigs, fed with a low-phosphorus diet, resulted in increased serum levels of SOD and glutathione peroxidase (GSH-Px), alongside decreased serum levels of bone-specific alkaline phosphatase. Vitamin D supplementation also elevated mucosal GSH-Px activity in the duodenum and ileum and TAC and SOD activity in the longissimus dorsi muscle. Furthermore, vitamin D significantly improved mRNA levels of copper/zinc SOD and modified the mRNA levels of Keap1 and Nrf2 [\[92\]](#page-15-3). In a study assessing the impact of vitamin D on growth performance, antioxidant enzyme activity, and antimicrobial capability in juvenile grass carp, six supplemental levels of vitamin D $(0, 300, 600, 1200, 2400,$ and 4800 IU/kg diet) were tested. Supplementation with vitamin D at levels ranging from 300 to 2400 IU/kg significantly improved growth performance and antioxidant enzyme activity in fish livers compared to the vitamin D deficiency group [\[93\]](#page-15-4).

4.1.2. Human Studies

While the impact of vitamin D on mitigating OS in cellular and animal models has been consistently demonstrated, clinical outcomes in human subjects have yielded mixed results. This variability may be attributed to factors such as differences in dosage, treatment duration, and clinical settings [\[94\]](#page-15-5). Vitamin D has been investigated for its potential therapeutic effects across a range of diseases in humans. There have been several studies exploring the antioxidant properties of vitamin D within the context of COVID-19 [\[95–](#page-15-6)[97\]](#page-15-7); however, that is not the main focus of this review.

In order to illustrate the importance of vitamin D in OS, a wide range of pathologies are shown below.

A recent meta-analysis suggests that antioxidants could represent an effective and safe treatment for patients with atopic dermatitis, particularly when combined with oral vitamin D and topical vitamin B_{12} [\[98\]](#page-15-8). In a randomized, double-blind, placebo-controlled trial involving 75 children aged 6–12 with attention deficit hyperactivity disorder, children were allocated to receive either vitamin D (2000 IU) or a placebo for three months. The supplementation with vitamin D did not yield significant benefits on biomarkers of OS [\[99\]](#page-15-9).

In patients with Behçet's disease, an association has been observed between low levels of vitamin D and increased OS, as evidenced by elevated MDA and NO levels, alongside decreased levels of GSH, SOD, and catalase activities, as well as TAC. Therefore, fortified foods, beverages, or vitamin D supplements may potentially ameliorate the severity of Behçet's disease and reduce OS [\[70\]](#page-14-4).

A recent umbrella meta-analysis indicates that vitamin D supplementation in adults may lead to reductions in C-reactive protein (CRP), MDA, and $TNF-\alpha$ levels across various health conditions. Consequently, vitamin D could be studied as a supplementary therapy for alleviating inflammation and OS [\[62\]](#page-13-20).

Furthermore, the impact of supplementation with vitamin D on inflammatory biomarkers and OS in pregnant women has been assessed in another systematic review, which reported increased levels of vitamin D, TAC, and GSH, along with decreased MDA concentrations following supplementation [\[100\]](#page-15-10).

4.2. Antioxidant Properties of Vitamin D in Chronic Diseases

4.2.1. Cardiovascular Health and Endothelial Function

VDRs have been identified in the heart, suggesting potential intracrine or paracrine roles in the cardiovascular system [\[87\]](#page-14-21). In fact, Vitamin D use has been recommended as an adjunct therapy in the treatment of inflammatory diseases and cardiometabolic conditions [\[21\]](#page-12-5). It exerts a vasoprotective effect by reducing OS-induced endothelial dysfunction and regulating the bioavailability of NO [\[101\]](#page-15-11). OS is a key contributor to endothelial dysfunction, damaging the endothelial lining and impairing its ability to control the blood flow and preserve vascular health. The antioxidant effects of vitamin D protect endothelial cells from oxidative damage, preserving their functionality and promoting vascular health [\[102\]](#page-15-12). ROS are well-known contributors to cardiovascular disease, being the excessive release of these free radicals from various intracellular sources related to disturbances in the normal function of endothelial cells and cardiac myocytes. Hence, OS is implicated in the development of different diseases, such as cardiac hypertrophy and heart fibrosis resulting from long-term hypertension [\[103\]](#page-15-13).

Vitamin D regulates the endothelial cell function by interacting with the endothelial NOS pathway. The NOS is an enzyme responsible for producing NO in the endothelium. Vitamin D enhances NOS expression and activity, thereby promoting NO production and supporting the optimal endothelial function. The NOS pathway is essential for maintaining vascular homeostasis and preventing endothelial dysfunction [\[104\]](#page-15-14).

4.2.2. Diabetes and Insulin Resistance

Besides hyperinsulinemia and hyperglycemia, diabetes is characterized by increased free radical formation and decreased TAC [\[105\]](#page-15-15). Vitamin D supplementation protocols that normalize blood vitamin D concentrations may improve glycemic control parameters and reduce OS in diabetic patients [\[106\]](#page-15-16). In this regard, vitamin D plays a significant role in managing OS by modulating the Nrf2 gene expression and influencing the pathogenesis of diabetes through its effects on the β-cell function, peripheral insulin sensitivity, or both [\[90\]](#page-15-1). Furthermore, vitamin D has been effective in improving the inflammatory response and OS, as well as maintaining glucose metabolic homeostasis in patients with gestational diabetes mellitus [\[107\]](#page-15-17). Moreover, a decrease in serum MDA levels and an increase in TAC following vitamin D supplementation have also been reported in patients with diabetes and vitamin D deficiency [\[108\]](#page-15-18). A high dosage of vitamin D supplementation significantly elevated TAC and GSH levels in women with gestational diabetes [\[109\]](#page-15-19). Conversely, vitamin D supplementation did not affect the majority of antioxidant enzyme activities in diabetic mice. By increasing intracellular zinc concentrations, oxidative damage was indirectly mitigated [\[110\]](#page-15-20). In older people with diabetes, blood vitamin D concentrations were inversely

correlated with various circulating OS biomarkers, including advanced oxidation protein products, low-density lipoprotein oxidation susceptibility, and NO metabolites, which were indirectly mitigated by increasing intracellular zinc concentrations [\[111\]](#page-16-0). Furthermore, antioxidant supplementation may not be beneficial to all individuals with diabetes, as their response to vitamin D supplementation may vary depending on their VDR FokI genotype [\[112\]](#page-16-1).

4.2.3. Neurodegenerative Diseases

Vitamin D has a significant role in neuronal development, encompassing processes such as proliferation, differentiation, migration, and axonal growth. It stimulates GGT activity, thereby preventing hippocampal damage [\[113\]](#page-16-2). Decreased TAC and increased ROS lead to damage in DNA, proteins, and membrane fatty acids, ultimately resulting in cell neurodegeneration, apoptosis, and volumetric changes in the brain [\[114\]](#page-16-3). The brain is particularly vulnerable to OS and damage due to its high oxygen consumption, deficiency in antioxidant enzymes, and high content of polyunsaturated fatty acids, which are prone to oxidation. Vitamin D regulates the oxidant–antioxidant balance by increasing intracellular antioxidant concentrations and reducing OS by scavenging excess free radicals [\[115\]](#page-16-4). As an antioxidant, vitamin D controls brain detoxification processes through the upregulation of GGT activity [\[78\]](#page-14-12). The neuroprotective role of vitamin D is extremely important due to its role in regulating calcium homeostasis in brain cells [\[116\]](#page-16-5). By treating hippocampal cells with high doses of vitamin D, they were protected from age-related excitotoxicity, resulting in a significant downregulation of L-type voltage-sensitive calcium channels [\[117,](#page-16-6)[118\]](#page-16-7), which are normally expected to surge in aging hippocampal cells and damaged cells [\[119,](#page-16-8)[120\]](#page-16-9). A total of 74 genes and 36 proteins are regulated by vitamin D, and it also participates in the post-transcriptional control of voltage-sensitive calcium channels of the L-type [\[121,](#page-16-10)[122\]](#page-16-11).

Furthermore, after traumatic brain injury, where there is an increase in autophagy and an inhibition of Nrf2 signaling leading to uncontrolled OS, vitamin D treatment significantly reduces neurological deficits and histopathological changes [\[123\]](#page-16-12).

This study demonstrated that vitamin D effectively protected the mitochondria in a Parkinson's disease model from dysfunction and oxidative damage, suggesting that it could be used as a therapeutic agent in the treatment of neurodegenerative disease [\[124\]](#page-16-13). Vitamin D supplementation reduced MDA levels, suggesting a protective effect that is at least partially associated with the prevention of oxidative damage production [\[125\]](#page-16-14). Vitamin D could play also an important role in modulation inflammation in the hypertensive brain, where the renin-angiotensin system (RAS) exerts pro-inflammatory and pro-oxidant effects. Vitamin D modulates the RAS, and it could be responsible for a neuroprotective effect described in this pathology [\[126\]](#page-16-15). Indeed, vitamin D deficiency has been reported to downregulate antioxidant enzymes, including SOD and GSH-Px. Vitamin D also inhibits NOS, thereby reducing NO production, while increasing GSH levels in the brain [\[127\]](#page-16-16). Regarding Alzheimer's disease, vitamin D may ameliorate cognitive deficits induced by aluminum-chloride-D-galactose through the upregulation of the calcium/calmodulindependent protein kinase-kinase 2-AMPK/Sirtuin1 pathway. This mechanism restores the normal mitochondrial function and reduces inflammation and OS [\[128\]](#page-16-17). Furthermore, given that OS can initiate microglial activation, which contributes to the pathophysiology of Alzheimer's disease, vitamin D's antioxidant properties through the activation of Nrf2 may be associated with its neuroprotective properties against the development and progression of Alzheimer's disease [\[129\]](#page-16-18). Moreover, by acting as an antioxidant, it has been shown to reduce OS biomarkers in rats with Alzheimer's disease hippocampus and serum [\[130\]](#page-16-19), while vitamin D has been shown to protect the cortex from OS by increasing the levels of SOD, GSH, and TAC, as well as reducing MDA levels in mice [\[131\]](#page-16-20).

4.2.4. Chronic Kidney Disease

Dietary antioxidants, such as vitamin D, which scavenge free radicals, are garnering significant interest as potential therapeutic agents for patients with chronic kidney disease (CKD) [\[132\]](#page-16-21). CKD is a global health issue affecting 5–10% of the population worldwide, and is characterized by an elevated inflammatory state, impaired immune response, and OS [\[133\]](#page-17-0). In individuals with CKD, vitamin D has been demonstrated to mitigate OS by upregulating Nrf2 and activating the antioxidant response element, highlighting a non-traditional regulatory function of the vitamin D pathway in the context of CKD [\[94\]](#page-15-5). Specifically, the expression of Nrf2 and serum levels of antioxidant enzymes have been observed to increase in individuals with end-stage renal disease [\[134\]](#page-17-1). Furthermore, vitamin D enhances the expression of Klotho proteins, a family involved in antioxidant production that is predominantly expressed in the kidneys. Klotho proteins have been shown to reduce OS by modulating the NF-kB expression and preventing other deleterious effects of CKD [\[135\]](#page-17-2). Conversely, vitamin D deficiency was not found to play a significant role in the increased OS state observed in pre-dialysis CKD patients, indicating the multifactorial nature of this condition [\[136\]](#page-17-3).

4.2.5. Cancer

Age-related reductions in vitamin D synthesis and VDR expression are linked to age-related diseases such as cancer [\[137\]](#page-17-4). Increased vitamin D intake and higher blood vitamin D concentrations have been proposed as interventions to mitigate conditions such as cancer [\[3\]](#page-11-2). Despite this, human studies on the effects of vitamin D treatment on circulating antioxidant and OS biomarkers, as well as cancer incidence and prognosis, have produced conflicting results [\[138\]](#page-17-5). Vitamin D contributes to tumorigenesis through various pathways, including the upregulation of antioxidant enzymes and inhibition of lipid peroxidation [\[138\]](#page-17-5). VDR activation may inhibit p53 signaling pathway activation, thereby promoting the proliferation, invasion, and metastasis of esophageal squamous cell carcinoma through OS mechanisms [\[139\]](#page-17-6). Moreover, osteosarcoma, which is characterized by high ROS production, relies significantly on OS for its pathogenesis. Vitamin D has been found to inhibit osteosarcoma, suggesting therapeutic applications not only for osteosarcoma patients but also for the broader aging population by mitigating OS [\[140\]](#page-17-7). Another possible mechanism involves vitamin D deficiency impairing mitochondrial respiration by downregulating complex I expression in the electron transport chain. This downregulation reduces the formation of adenosine triphosphate, thereby increasing cancer risk. Furthermore, compromised electron transport chain activity leads to elevated ROS production and heightened OS [\[141\]](#page-17-8). Finally, CYP11A1-derived secosteroids exhibit anti-cancer activities that vary depending on the specific cell type and lineage involved [\[142\]](#page-17-9). In this context, vitamin D hydroxymetabolites and lumisterol, along with their respective receptors, play a pivotal role in the prevention of skin cancer by activating protective anti-cancer pathways, such as the Nrf2-antioxidant response and p53 phosphorylation. Furthermore, these compounds promote DNA repair mechanisms that safeguard human keratinocytes from DNA damage [\[143\]](#page-17-10). UVR enhances the production of ROS, which may have harmful effects on the skin, and induces the tumor suppressor protein p53 as part of the cellular response to DNA damage. This process increases the risk of melanoma, a risk that may be mitigated by the antimelanoma activity of CYP11A1-derived secosteroids [\[27\]](#page-12-11).

5. Future Directions

Despite the progress made in the study of molecular mechanisms related to Vitamin D's antioxidant activity, they are not completely understood at present. In this regard, investigating the precise molecular pathways through which vitamin D exerts its antioxidant effects is needed, being one of the promising markers the gene expression of the VDR and its effect on different processes [\[144\]](#page-17-11). Additionally, another interesting field would be to explore vitamin D's impact on the mitochondrial function and its role in reducing mitochondrial OS. This includes studying how vitamin D affects the mitochondrial biogenesis, dynamics, and function [\[145\]](#page-17-12).

On the other hand, it is necessary to conduct large-scale and long-term clinical trials to evaluate the efficacy of vitamin D supplementation in preventing and managing OS-related diseases, as well as to identify and validate specific biomarkers of OS that can be reliably used to measure the antioxidant effects in human subjects [\[146\]](#page-17-13). Furthermore, it would be necessary to determine the optimal dosage and form of vitamin D supplementation to achieve significant antioxidant effects, without adverse outcomes and taking into account factors like age, sex, ethnicity, genetic variations, and pre-existing health conditions, leading to more tailored supplementation recommendations [\[50\]](#page-13-8). Similarly, the development and efficacy of vitamin D analogues and derivatives that might have enhanced antioxidant properties or reduced side effects compared to natural vitamin D should be investigated [\[147\]](#page-17-14).

In the context of CYP11A1-derived vitamin D and lumisterol metabolites, exploring the therapeutic potential of vitamin D hydroxyderivatives in the treatment of chronic diseases associated with OS presents promising new avenues for medical intervention. However, the precise molecular mechanisms through which these metabolites modulate OS remain only partially understood. Future research should prioritize the identification of specific signaling pathways and target genes that are regulated by these metabolites. This includes examining their interactions with non-classical receptors and elucidating how these interactions affect cellular redox homeostasis [\[37,](#page-12-21)[148](#page-17-15)[,149\]](#page-17-16).

Finally, the relationship between vitamin D status and the progression of chronic diseases characterized by high OS levels, providing insights into potential therapeutic roles for vitamin D like cardiovascular diseases, type 2 diabetes, and CKD should be studied more in greater depth [\[150\]](#page-17-17). Likewise, vitamin D's role in modulating the immune system's response to OS, particularly in conditions where inflammation and OS are closely linked, such as autoimmune diseases, should be investigated [\[151\]](#page-17-18). Moreover, conducting studies to assess the impact of vitamin D on cognitive function and the prevention of cognitive decline through its antioxidant properties could clarify its neuroprotective effects [\[152\]](#page-17-19). An overview of vitamin D's future directions in antioxidant activity is shown in Figure [3.](#page-10-0)

Figure 3. Main future directions for vitamin D. **Figure 3.** Main future directions for vitamin D.

6. Conclusions

Vitamin D has been seen to affect different parameters related to antioxidant status. This function is mainly associated with increased levels of antioxidant enzymes and the regulation of the expression of Nrf2 and Klotho. These antioxidant properties present a compelling case for vitamin D's role in promoting cellular health and preventing oxidative damage. Future research should focus on elucidating the precise molecular pathways involved and exploring the therapeutic potential of CYP11A1-derived vitamin D and tachysterol and lumisterol metabolites in the skin and vitamin D supplementation in conditions characterized by elevated OS as chronic diseases. This expanded understanding could pave the way for novel interventions aimed at leveraging their antioxidant capabilities to enhance health outcomes and potentially reduce chronic diseases' impact on human health.

Author Contributions: Conceptualization, H.V.-L. and J.P.-D.; methodology, L.H.-Q., L.J.-S., B.F.-P. and H.V.-L.; software, J.P.-D.; validation, J.P.-D.; writing—original draft preparation, L.H.-Q., L.J.-S., B.F.-P. and H.V.-L.; writing—review and editing, J.P.-D.; visualization, J.P.-D.; supervision J.P.-D. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Acknowledgments: Julio Plaza-Diaz is part of the "UGR Plan Propio de Investigación 2016" and the "Excellence actions: Unit of Excellence on Exercise and Health (UCEES), University of Granada".

Conflicts of Interest: The authors declare no conflicts of interest.

References

- 1. Fantini, C.; Corinaldesi, C.; Lenzi, A.; Migliaccio, S.; Crescioli, C. Vitamin D as a Shield against Aging. *Int. J. Mol. Sci.* **2023**, *24*, 4546. [\[CrossRef\]](https://doi.org/10.3390/ijms24054546)
- 2. Slominski, R.M.; Chen, J.Y.; Raman, C.; Slominski, A.T. Photo-neuro-immuno-endocrinology: How the ultraviolet radiation regulates the body, brain, and immune system. *Proc. Natl. Acad. Sci. USA* **2024**, *121*, e2308374121. [\[CrossRef\]](https://doi.org/10.1073/pnas.2308374121)
- 3. Wimalawansa, S.J. Physiology of Vitamin D-Focusing on Disease Prevention. *Nutrients* **2024**, *16*, 1666. [\[CrossRef\]](https://doi.org/10.3390/nu16111666)
- 4. Pludowski, P.; Holick, M.F.; Grant, W.B.; Konstantynowicz, J.; Mascarenhas, M.R.; Haq, A.; Povoroznyuk, V.; Balatska, N.; Barbosa, A.P.; Karonova, T.; et al. Vitamin D supplementation guidelines. *J. Steroid Biochem. Mol. Biol.* **2018**, *175*, 125–135. [\[CrossRef\]](https://doi.org/10.1016/j.jsbmb.2017.01.021)
- 5. Zhao, J.F.; Li, B.X.; Zhang, Q. Vitamin D improves levels of hormonal, oxidative stress and inflammatory parameters in polycystic ovary syndrome: A meta-analysis study. *Ann. Palliat. Med.* **2021**, *10*, 169–183. [\[CrossRef\]](https://doi.org/10.21037/apm-20-2201)
- 6. Yang, T.; Wang, H.; Xiong, Y.; Chen, C.; Duan, K.; Jia, J.; Ma, F. Vitamin D Supplementation Improves Cognitive Function Through Reducing Oxidative Stress Regulated by Telomere Length in Older Adults with Mild Cognitive Impairment: A 12-Month Randomized Controlled Trial. *J. Alzheimers Dis.* **2020**, *78*, 1509–1518. [\[CrossRef\]](https://doi.org/10.3233/JAD-200926)
- 7. Reddy, V.P. Oxidative Stress in Health and Disease. *Biomedicines* **2023**, *11*, 2925. [\[CrossRef\]](https://doi.org/10.3390/biomedicines11112925)
- 8. Kıran, T.R.; Otlu, O.; Karabulut, A.B. Oxidative stress and antioxidants in health and disease. *J. Lab. Med.* **2023**, *47*, 1–11. [\[CrossRef\]](https://doi.org/10.1515/labmed-2022-0108)
- 9. Aramouni, K.; Assaf, R.; Shaito, A.; Fardoun, M.; Al-Asmakh, M.; Sahebkar, A.; Eid, A.H. Biochemical and cellular basis of oxidative stress: Implications for disease onset. *J. Cell Physiol.* **2023**, *238*, 1951–1963. [\[CrossRef\]](https://doi.org/10.1002/jcp.31071)
- 10. Tyuryaeva, I.; Lyublinskaya, O. Expected and Unexpected Effects of Pharmacological Antioxidants. *Int. J. Mol. Sci.* **2023**, *24*, 9303. [\[CrossRef\]](https://doi.org/10.3390/ijms24119303)
- 11. Miao, D.; Goltzman, D. Mechanisms of action of vitamin D in delaying aging and preventing disease by inhibiting oxidative stress. *Vitam. Horm.* **2023**, *121*, 293–318. [\[CrossRef\]](https://doi.org/10.1016/bs.vh.2022.09.004)
- 12. van der Pol, A.; van Gilst, W.H.; Voors, A.A.; van der Meer, P. Treating oxidative stress in heart failure: Past, present and future. *Eur. J. Heart Fail.* **2019**, *21*, 425–435. [\[CrossRef\]](https://doi.org/10.1002/ejhf.1320)
- 13. Pingitore, A.; Lima, G.P.; Mastorci, F.; Quinones, A.; Iervasi, G.; Vassalle, C. Exercise and oxidative stress: Potential effects of antioxidant dietary strategies in sports. *Nutrition* **2015**, *31*, 916–922. [\[CrossRef\]](https://doi.org/10.1016/j.nut.2015.02.005)
- 14. Jones, D.P. Radical-free biology of oxidative stress. *Am. J. Physiol. Cell Physiol.* **2008**, *295*, C849–C868. [\[CrossRef\]](https://doi.org/10.1152/ajpcell.00283.2008)
- 15. Bal, A.; Panda, F.; Pati, S.G.; Das, K.; Agrawal, P.K.; Paital, B. Modulation of physiological oxidative stress and antioxidant status by abiotic factors especially salinity in aquatic organisms. *Comp. Biochem. Physiol. C Toxicol. Pharmacol.* **2021**, *241*, 108971. [\[CrossRef\]](https://doi.org/10.1016/j.cbpc.2020.108971)
- 16. Filomeni, G.; De Zio, D.; Cecconi, F. Oxidative stress and autophagy: The clash between damage and metabolic needs. *Cell Death Differ.* **2015**, *22*, 377–388. [\[CrossRef\]](https://doi.org/10.1038/cdd.2014.150)
- 17. Marrocco, I.; Altieri, F.; Peluso, I. Measurement and Clinical Significance of Biomarkers of Oxidative Stress in Humans. *Oxid. Med. Cell Longev.* **2017**, *2017*, 6501046. [\[CrossRef\]](https://doi.org/10.1155/2017/6501046)
- 18. Checa, J.; Aran, J.M. Reactive Oxygen Species: Drivers of Physiological and Pathological Processes. *J. Inflamm. Res.* **2020**, *13*, 1057–1073. [\[CrossRef\]](https://doi.org/10.2147/JIR.S275595)
- 19. Kimball, J.S.; Johnson, J.P.; Carlson, D.A. Oxidative Stress and Osteoporosis. *J. Bone Jt. Surg. Am.* **2021**, *103*, 1451–1461. [\[CrossRef\]](https://doi.org/10.2106/JBJS.20.00989)
- 20. Arazi, H.; Eghbali, E.; Suzuki, K. Creatine Supplementation, Physical Exercise and Oxidative Stress Markers: A Review of the Mechanisms and Effectiveness. *Nutrients* **2021**, *13*, 869. [\[CrossRef\]](https://doi.org/10.3390/nu13030869)
- 21. Renke, G.; Starling-Soares, B.; Baesso, T.; Petronio, R.; Aguiar, D.; Paes, R. Effects of Vitamin D on Cardiovascular Risk and Oxidative Stress. *Nutrients* **2023**, *15*, 769. [\[CrossRef\]](https://doi.org/10.3390/nu15030769)
- 22. Norman, A.W. From vitamin D to hormone D: Fundamentals of the vitamin D endocrine system essential for good health. *Am. J. Clin. Nutr.* **2008**, *88*, 491S–499S. [\[CrossRef\]](https://doi.org/10.1093/ajcn/88.2.491S)
- 23. Nimitphong, H.; Holick, M.F. Vitamin D status and sun exposure in southeast Asia. *Dermato-Endocrinology* **2013**, *5*, 34–37. [\[CrossRef\]](https://doi.org/10.4161/derm.24054)
- 24. Iddir, M.; Brito, A.; Dingeo, G.; Fernandez Del Campo, S.S.; Samouda, H.; La Frano, M.R.; Bohn, T. Strengthening the Immune System and Reducing Inflammation and Oxidative Stress through Diet and Nutrition: Considerations during the COVID-19 Crisis. *Nutrients* **2020**, *12*, 1562. [\[CrossRef\]](https://doi.org/10.3390/nu12061562)
- 25. Gil, A.; Plaza-Diaz, J.; Mesa, M.D. Vitamin D: Classic and Novel Actions. *Ann. Nutr. Metab.* **2018**, *72*, 87–95. [\[CrossRef\]](https://doi.org/10.1159/000486536) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29346788)
- 26. Wacker, M.; Holick, M.F. Sunlight and Vitamin D: A global perspective for health. *Dermato-Endocrinology* **2013**, *5*, 51–108. [\[CrossRef\]](https://doi.org/10.4161/derm.24494)
- 27. Slominski, A.T.; Brozyna, A.A.; Skobowiat, C.; Zmijewski, M.A.; Kim, T.K.; Janjetovic, Z.; Oak, A.S.; Jozwicki, W.; Jetten, A.M.; Mason, R.S.; et al. On the role of classical and novel forms of vitamin D in melanoma progression and management. *J. Steroid Biochem. Mol. Biol.* **2018**, *177*, 159–170. [\[CrossRef\]](https://doi.org/10.1016/j.jsbmb.2017.06.013)
- 28. Zmijewski, M.A.; Li, W.; Chen, J.; Kim, T.K.; Zjawiony, J.K.; Sweatman, T.W.; Miller, D.D.; Slominski, A.T. Synthesis and photochemical transformation of 3beta,21-dihydroxypregna-5,7-dien-20-one to novel secosteroids that show anti-melanoma activity. *Steroids* **2011**, *76*, 193–203. [\[CrossRef\]](https://doi.org/10.1016/j.steroids.2010.10.009)
- 29. Qayyum, S.; Slominski, R.M.; Raman, C.; Slominski, A.T. Novel CYP11A1-Derived Vitamin D and Lumisterol Biometabolites for the Management of COVID-19. *Nutrients* **2022**, *14*, 4779. [\[CrossRef\]](https://doi.org/10.3390/nu14224779)
- 30. Mokhtari-Zaer, A.; Hosseini, M.; Salmani, H.; Arab, Z.; Zareian, P. Vitamin D(3) attenuates lipopolysaccharide-induced cognitive impairment in rats by inhibiting inflammation and oxidative stress. *Life Sci.* **2020**, *253*, 117703. [\[CrossRef\]](https://doi.org/10.1016/j.lfs.2020.117703)
- 31. Andress, D.L. Vitamin D in chronic kidney disease: A systemic role for selective vitamin D receptor activation. *Kidney Int.* **2006**, *69*, 33–43. [\[CrossRef\]](https://doi.org/10.1038/sj.ki.5000045)
- 32. Shaban, N.Z.; Abdel-Alnaby, M.; Atta, M.N.; Abdul-Aziz, A.A.; Megahed, F. The association between body mass index elevation and differentiation in vitamin D receptor gene expression, genetic polymorphism, and oxidative stress in adult Egyptian individuals. *Sci. Rep.* **2023**, *13*, 17696. [\[CrossRef\]](https://doi.org/10.1038/s41598-023-44607-4)
- 33. Tangpricha, V.; Spina, C.; Yao, M.; Chen, T.C.; Wolfe, M.M.; Holick, M.F. Vitamin D deficiency enhances the growth of MC-26 colon cancer xenografts in Balb/c mice. *J. Nutr.* **2005**, *135*, 2350–2354. [\[CrossRef\]](https://doi.org/10.1093/jn/135.10.2350)
- 34. Jiang, P.; Zhang, L.; Zhu, W.; Li, H.; Dang, R.; Tang, M. Chronic stress causes neuroendocrine-immune disturbances without affecting renal vitamin D metabolism in rats. *J. Endocrinol. Investig.* **2014**, *37*, 1109–1116. [\[CrossRef\]](https://doi.org/10.1007/s40618-014-0191-5)
- 35. Chaiprasongsuk, A.; Janjetovic, Z.; Kim, T.K.; Jarrett, S.G.; D'Orazio, J.A.; Holick, M.F.; Tang, E.K.Y.; Tuckey, R.C.; Panich, U.; Li, W.; et al. Protective effects of novel derivatives of vitamin D(3) and lumisterol against UVB-induced damage in human keratinocytes involve activation of Nrf2 and p53 defense mechanisms. *Redox Biol.* **2019**, *24*, 101206. [\[CrossRef\]](https://doi.org/10.1016/j.redox.2019.101206)
- 36. Slominski, A.T.; Kim, T.K.; Janjetovic, Z.; Brozyna, A.A.; Zmijewski, M.A.; Xu, H.; Sutter, T.R.; Tuckey, R.C.; Jetten, A.M.; Crossman, D.K. Differential and Overlapping Effects of 20,23(OH)(2)D3 and 1,25(OH)(2)D3 on Gene Expression in Human Epidermal Keratinocytes: Identification of AhR as an Alternative Receptor for 20,23(OH)(2)D3. *Int. J. Mol. Sci.* **2018**, *19*, 3072. [\[CrossRef\]](https://doi.org/10.3390/ijms19103072)
- 37. Slominski, A.T.; Kim, T.K.; Janjetovic, Z.; Slominski, R.M.; Li, W.; Jetten, A.M.; Indra, A.K.; Mason, R.S.; Tuckey, R.C. Biological Effects of CYP11A1-Derived Vitamin D and Lumisterol Metabolites in the Skin. *J. Investig. Dermatol.* **2024**, in press. [\[CrossRef\]](https://doi.org/10.1016/j.jid.2024.04.022)
- 38. Chaiprasongsuk, A.; Janjetovic, Z.; Kim, T.K.; Schwartz, C.J.; Tuckey, R.C.; Tang, E.K.Y.; Raman, C.; Panich, U.; Slominski, A.T. Hydroxylumisterols, Photoproducts of Pre-Vitamin D3, Protect Human Keratinocytes against UVB-Induced Damage. *Int. J. Mol. Sci.* **2020**, *21*, 9374. [\[CrossRef\]](https://doi.org/10.3390/ijms21249374)
- 39. Slominski, A.T.; Li, W.; Kim, T.K.; Semak, I.; Wang, J.; Zjawiony, J.K.; Tuckey, R.C. Novel activities of CYP11A1 and their potential physiological significance. *J. Steroid Biochem. Mol. Biol.* **2015**, *151*, 25–37. [\[CrossRef\]](https://doi.org/10.1016/j.jsbmb.2014.11.010)
- 40. Slominski, A.T.; Brozyna, A.A.; Kim, T.K.; Elsayed, M.M.; Janjetovic, Z.; Qayyum, S.; Slominski, R.M.; Oak, A.S.W.; Li, C.; Podgorska, E.; et al. CYP11A1-derived vitamin D hydroxyderivatives as candidates for therapy of basal and squamous cell carcinomas. *Int. J. Oncol.* **2022**, *61*, 96. [\[CrossRef\]](https://doi.org/10.3892/ijo.2022.5386)
- 41. Chassoux, F.; Navarro, V.; Quirins, M.; Laurent, A.; Gavaret, M.; Cousyn, L.; Crepon, B.; Landre, E.; Marchi, A.; Soufflet, C.; et al. Vitamin D deficiency and effect of treatment on seizure frequency and quality of life parameters in patients with drug-resistant epilepsy: A randomized clinical trial. *Epilepsia* **2024**. [\[CrossRef\]](https://doi.org/10.1111/epi.18050) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38980968)
- 42. Xiang, Q.; Xu, H.; Liu, Y.; Huang, W. Elevated TyG index is associated with increased risk of vitamin D deficiency among elderly patients with type 2 diabetes. *Sci. Rep.* **2024**, *14*, 16098. [\[CrossRef\]](https://doi.org/10.1038/s41598-024-67127-1) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38997409)
- 43. Swarnkar, M.; Kumar, K.; Prasad, P.; Singhal, K. Association Between Vitamin D Deficiency and Tumor Characteristics in Breast Cancer Patients. *Cureus* **2024**, *16*, e62296. [\[CrossRef\]](https://doi.org/10.7759/cureus.62296) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/39006561)
- 44. Jain, S.K.; Justin Margret, J.; Abrams, S.A.; Levine, S.N.; Bhusal, K. The Impact of Vitamin D and L-Cysteine Co-Supplementation on Upregulating Glutathione and Vitamin D-Metabolizing Genes and in the Treatment of Circulating 25-Hydroxy Vitamin D Deficiency. *Nutrients* **2024**, *16*, 2004. [\[CrossRef\]](https://doi.org/10.3390/nu16132004) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38999752)
- 45. Barrea, L.; Frias-Toral, E.; Pugliese, G.; Garcia-Velasquez, E.; Carignano M, D.L.A.; Savastano, S.; Colao, A.; Muscogiuri, G. Vitamin D in obesity and obesity-related diseases: An overview. *Minerva Endocrinol.* **2021**, *46*, 177–192. [\[CrossRef\]](https://doi.org/10.23736/S2724-6507.20.03299-X)
- 46. Ramezani-Jolfaie, N.; Eftekhar, E.; Dadinasab, M.; Hesarooeyeh, Z.G.; Pakdaman, P.; Razmpour, F.; Javedan, G.; Khayatian, M.; Azad, M.H.; Davoodian, P.; et al. The effect of vitamin D and magnesium supplementation on clinical symptoms and serum inflammatory and oxidative stress markers in patients with COVID-19: A structured summary of a study protocol for a randomized controlled trial. *Trials* **2023**, *24*, 87. [\[CrossRef\]](https://doi.org/10.1186/s13063-023-07107-4) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36747270)
- 47. Wilson, L.R.; Tripkovic, L.; Hart, K.H.; Lanham-New, S.A. Vitamin D deficiency as a public health issue: Using vitamin D2 or vitamin D3 in future fortification strategies. *Proc. Nutr. Soc.* **2017**, *76*, 392–399. [\[CrossRef\]](https://doi.org/10.1017/S0029665117000349)
- 48. Chiang, C.M.; Ismaeel, A.; Griffis, R.B.; Weems, S. Effects of Vitamin D Supplementation on Muscle Strength in Athletes: A Systematic Review. *J. Strength. Cond. Res.* **2017**, *31*, 566–574. [\[CrossRef\]](https://doi.org/10.1519/JSC.0000000000001518) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27379960)
- 49. Armas, L.A.; Heaney, R.P. Vitamin D: The iceberg nutrient. *J. Ren. Nutr.* **2011**, *21*, 134–139. [\[CrossRef\]](https://doi.org/10.1053/j.jrn.2010.09.002)
- 50. Holick, M.F. Vitamin D deficiency. *N. Engl. J. Med.* **2007**, *357*, 266–281. [\[CrossRef\]](https://doi.org/10.1056/NEJMra070553)
- 51. Fathi, F.; Sadek, K.M.; Khafaga, A.F.; Al Senosy, A.W.; Ghoniem, H.A.; Fayez, S.; Zeweil, M.F. Vitamin D regulates insulin and ameliorates apoptosis and oxidative stress in pancreatic tissues of rats with streptozotocin-induced diabetes. *Environ. Sci. Pollut. Res. Int.* **2022**, *29*, 90219–90229. [\[CrossRef\]](https://doi.org/10.1007/s11356-022-22064-2) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35864405)
- 52. Torres Dominguez, E.A.; Meza Penafiel, A.; Gomez Pedraza, A.; Martinez Leo, E.E. Molecular mechanisms from insulin-mimetic effect of vitamin D: Treatment alternative in Type 2 diabetes mellitus. *Food Funct.* **2021**, *12*, 6682–6690. [\[CrossRef\]](https://doi.org/10.1039/D0FO03230A)
- 53. Alonso, N.; Zelzer, S.; Eibinger, G.; Herrmann, M. Vitamin D Metabolites: Analytical Challenges and Clinical Relevance. *Calcif. Tissue Int.* **2023**, *112*, 158–177. [\[CrossRef\]](https://doi.org/10.1007/s00223-022-00961-5)
- 54. Cortes, M.; Chen, M.J.; Stachura, D.L.; Liu, S.Y.; Kwan, W.; Wright, F.; Vo, L.T.; Theodore, L.N.; Esain, V.; Frost, I.M.; et al. Developmental Vitamin D Availability Impacts Hematopoietic Stem Cell Production. *Cell Rep.* **2016**, *17*, 458–468. [\[CrossRef\]](https://doi.org/10.1016/j.celrep.2016.09.012)
- 55. Novershtern, N.; Subramanian, A.; Lawton, L.N.; Mak, R.H.; Haining, W.N.; McConkey, M.E.; Habib, N.; Yosef, N.; Chang, C.Y.; Shay, T.; et al. Densely interconnected transcriptional circuits control cell states in human hematopoiesis. *Cell* **2011**, *144*, 296–309. [\[CrossRef\]](https://doi.org/10.1016/j.cell.2011.01.004)
- 56. Kim, S.; Yamazaki, M.; Zella, L.A.; Meyer, M.B.; Fretz, J.A.; Shevde, N.K.; Pike, J.W. Multiple enhancer regions located at significant distances upstream of the transcriptional start site mediate RANKL gene expression in response to 1,25-dihydroxyvitamin D3. *J. Steroid Biochem. Mol. Biol.* **2007**, *103*, 430–434. [\[CrossRef\]](https://doi.org/10.1016/j.jsbmb.2006.12.020)
- 57. Elsaeed, M.Y.; Mehanna, O.M.; Abd-Allah, E.E.; Hassan, M.G.; Ahmed, W.M.S.; Moustafa, A.; Eldesoky, G.E.; Hammad, A.M.; Elgazzar, U.B.; Elnady, M.R.; et al. Combination Therapy with Enalapril and Paricalcitol Ameliorates Streptozotocin Diabetes-Induced Testicular Dysfunction in Rats via Mitigation of Inflammation, Apoptosis, and Oxidative Stress. *Pathophysiology* **2023**, *30*, 567–585. [\[CrossRef\]](https://doi.org/10.3390/pathophysiology30040041) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38133142)
- 58. Janjetovic, Z.; Postlethwaite, A.; Kang, H.S.; Kim, T.K.; Tuckey, R.C.; Crossman, D.K.; Qayyum, S.; Jetten, A.M.; Slominski, A.T. Antifibrogenic Activities of CYP11A1-derived Vitamin D3-hydroxyderivatives Are Dependent on RORgamma. *Endocrinology* **2021**, *162*, bqaa198. [\[CrossRef\]](https://doi.org/10.1210/endocr/bqaa198)
- 59. Chaiprasongsuk, A.; Janjetovic, Z.; Kim, T.K.; Tuckey, R.C.; Li, W.; Raman, C.; Panich, U.; Slominski, A.T. CYP11A1-derived vitamin D(3) products protect against UVB-induced inflammation and promote keratinocytes differentiation. *Free Radic. Biol. Med.* **2020**, *155*, 87–98. [\[CrossRef\]](https://doi.org/10.1016/j.freeradbiomed.2020.05.016)
- 60. Wiseman, H. Vitamin D is a membrane antioxidant Ability to inhibit iron-dependent lipid peroxidation in liposomes compared to cholesterol, ergosterol and tamoxifen and relevance to anticancer action. *FEBS Lett.* **1993**, *326*, 285–288. [\[CrossRef\]](https://doi.org/10.1016/0014-5793(93)81809-E) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/8325381)
- 61. Ke, C.Y.; Yang, F.L.; Wu, W.T.; Chung, C.H.; Lee, R.P.; Yang, W.T.; Subeq, Y.M.; Liao, K.W. Vitamin D3 Reduces Tissue Damage and Oxidative Stress Caused by Exhaustive Exercise. *Int. J. Med. Sci.* **2016**, *13*, 147–153. [\[CrossRef\]](https://doi.org/10.7150/ijms.13746) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26941574)
- 62. Moslemi, E.; Musazadeh, V.; Kavyani, Z.; Naghsh, N.; Shoura, S.M.S.; Dehghan, P. Efficacy of vitamin D supplementation as an adjunct therapy for improving inflammatory and oxidative stress biomarkers: An umbrella meta-analysis. *Pharmacol. Res.* **2022**, *186*, 106484. [\[CrossRef\]](https://doi.org/10.1016/j.phrs.2022.106484) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36206958)
- 63. Motamed, S.; Anari, R.; Motamed, S.; Amani, R. Vitamin D and biomarkers of inflammation and oxidative stress among pregnant women: A systematic review of observational studies. *BMC Immunol.* **2023**, *24*, 41. [\[CrossRef\]](https://doi.org/10.1186/s12865-023-00577-w) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37891486)
- 64. Camara, A.B.; Brandao, I.A. The relationship between vitamin D deficiency and oxidative stress can be independent of age and gender. *Int. J. Vitam. Nutr. Res.* **2021**, *91*, 108–123. [\[CrossRef\]](https://doi.org/10.1024/0300-9831/a000614) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31711376)
- 65. Nakai, K.; Fujii, H.; Kono, K.; Goto, S.; Kitazawa, R.; Kitazawa, S.; Hirata, M.; Shinohara, M.; Fukagawa, M.; Nishi, S. Vitamin D activates the Nrf2-Keap1 antioxidant pathway and ameliorates nephropathy in diabetic rats. *Am. J. Hypertens.* **2014**, *27*, 586–595. [\[CrossRef\]](https://doi.org/10.1093/ajh/hpt160) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24025724)
- 66. Berridge, M.J. Vitamin D cell signalling in health and disease. *Biochem. Biophys. Res. Commun.* **2015**, *460*, 53–71. [\[CrossRef\]](https://doi.org/10.1016/j.bbrc.2015.01.008) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25998734)
- 67. Calabrese, V.; Cornelius, C.; Mancuso, C.; Lentile, R.; Stella, A.G.; Butterfield, D.A. Redox homeostasis and cellular stress response in aging and neurodegeneration. In *Free Radicals and Antioxidant Protocols*; Humana Press: Totowa, NJ, USA, 2010; pp. 285–308.
- 68. Nisar, M.; Beigh, S.A.; Mir, A.Q.; Hussain, S.A.; Dar, A.A.; Yatoo, I.; Khan, A.M. Association of vitamin D status with redox balance and insulin resistance and its predicting ability for subclinical pregnancy toxemia in pregnant sheep. *Domest. Anim. Endocrinol.* **2024**, *86*, 106823. [\[CrossRef\]](https://doi.org/10.1016/j.domaniend.2023.106823) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37944202)
- 69. Talla, V.; Veerareddy, P. Oxidative stress induced by fluoroquinolones on treatment for complicated urinary tract infections in Indian patients. *J. Young Pharm.* **2011**, *3*, 304–309. [\[CrossRef\]](https://doi.org/10.4103/0975-1483.90242) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22224037)
- 70. Omar, H.S.; Taha, F.M.; Fouad, S.; Ibrahim, F.A.; El Gendy, A.; Bassyouni, I.H.; El-Shazly, R. The association between vitamin D levels and oxidative stress markers in Egyptian Behcet's disease patients. *Orphanet J. Rare Dis.* **2022**, *17*, 264. [\[CrossRef\]](https://doi.org/10.1186/s13023-022-02416-4)
- 71. Cusato, J.; Manca, A.; Palermiti, A.; Mula, J.; Costanzo, M.; Antonucci, M.; Chiara, F.; De Vivo, E.D.; Maiese, D.; Ferrara, M.; et al. COVID-19: Focusing on the Link between Inflammation, Vitamin D, MAPK Pathway and Oxidative Stress Genetics. *Antioxidants* **2023**, *12*, 1133. [\[CrossRef\]](https://doi.org/10.3390/antiox12051133)
- 72. Rizwan, M.; Cheng, K.; Gang, Y.; Hou, Y.; Wang, C. Immunomodulatory Effects of Vitamin D and Zinc on Viral Infection. *Biol. Trace Elem. Res.* **2024**. [\[CrossRef\]](https://doi.org/10.1007/s12011-024-04139-y) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38451442)
- 73. Mhaibes, A.M.; Abdul-Wahab, F.K. Nephroprotective effect of vitamin D Against Levofloxacin-induced renal injury: An observational study. *J. Med. Life* **2023**, *16*, 1032–1040. [\[CrossRef\]](https://doi.org/10.25122/jml-2023-0096) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37900077)
- 74. Atanasovska, E.; Petrusevska, M.; Zendelovska, D.; Spasovska, K.; Stevanovikj, M.; Kasapinova, K.; Gjorgjievska, K.; Labachevski, N. Vitamin D levels and oxidative stress markers in patients hospitalized with COVID-19. *Redox Rep.* **2021**, *26*, 184–189. [\[CrossRef\]](https://doi.org/10.1080/13510002.2021.1999126) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34727009)
- 75. Bao, B.Y.; Ting, H.J.; Hsu, J.W.; Lee, Y.F. Protective role of 1 alpha, 25-dihydroxyvitamin D3 against oxidative stress in nonmalignant human prostate epithelial cells. *Int. J. Cancer* **2008**, *122*, 2699–2706. [\[CrossRef\]](https://doi.org/10.1002/ijc.23460) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18348143)
- 76. Saif-Elnasr, M.; Ibrahim, I.M.; Alkady, M.M. Role of Vitamin D on glycemic control and oxidative stress in type 2 diabetes mellitus. *J. Res. Med. Sci.* **2017**, *22*, 22. [\[CrossRef\]](https://doi.org/10.4103/1735-1995.200278) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28413419)
- 77. Slominski, A.T.; Chaiprasongsuk, A.; Janjetovic, Z.; Kim, T.K.; Stefan, J.; Slominski, R.M.; Hanumanthu, V.S.; Raman, C.; Qayyum, S.; Song, Y.; et al. Photoprotective Properties of Vitamin D and Lumisterol Hydroxyderivatives. *Cell Biochem. Biophys.* **2020**, *78*, 165–180. [\[CrossRef\]](https://doi.org/10.1007/s12013-020-00913-6)
- 78. Wassif, G.A.; Alrehely, M.S.; Alharbi, D.M.; Aljohani, A.A. The Impact of Vitamin D on Neuropsychiatric Disorders. *Cureus* **2023**, *15*, e47716. [\[CrossRef\]](https://doi.org/10.7759/cureus.47716) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38022259)
- 79. Motamed, S.; Nikooyeh, B.; Kashanian, M.; Chamani, M.; Hollis, B.W.; Neyestani, T.R. Evaluation of the efficacy of two doses of vitamin D supplementation on glycemic, lipidemic and oxidative stress biomarkers during pregnancy: A randomized clinical trial. *BMC Pregnancy Childbirth* **2020**, *20*, 619. [\[CrossRef\]](https://doi.org/10.1186/s12884-020-03311-1) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33054794)
- 80. Imanparast, F.; Mashayekhi, F.J.; Kamankesh, F.; Rafiei, F.; Mohaghegh, P.; Alimoradian, A. Improving the endothelial dysfunction in type 2 diabetes with chromium and vitamin D(3) byreducing homocysteine and oxidative stress: A randomized placebocontrolled trial. *J. Trace Elem. Med. Biol.* **2020**, *62*, 126639. [\[CrossRef\]](https://doi.org/10.1016/j.jtemb.2020.126639)
- 81. Akbari, M.; Ostadmohammadi, V.; Lankarani, K.B.; Tabrizi, R.; Kolahdooz, F.; Heydari, S.T.; Kavari, S.H.; Mirhosseini, N.; Mafi, A.; Dastorani, M.; et al. The Effects of Vitamin D Supplementation on Biomarkers of Inflammation and Oxidative Stress Among Women with Polycystic Ovary Syndrome: A Systematic Review and Meta-Analysis of Randomized Controlled Trials. *Horm. Metab. Res.* **2018**, *50*, 271–279. [\[CrossRef\]](https://doi.org/10.1055/s-0043-125148)
- 82. Zhang, H.Q.; Teng, J.H.; Li, Y.; Li, X.X.; He, Y.H.; He, X.; Sun, C.H. Vitamin D status and its association with adiposity and oxidative stress in schoolchildren. *Nutrition* **2014**, *30*, 1040–1044. [\[CrossRef\]](https://doi.org/10.1016/j.nut.2014.02.024)
- 83. Hirunsai, M.; Srikuea, R. Differential effects of cholecalciferol and calcitriol on muscle proteolysis and oxidative stress in angiotensin II-induced C2C12 myotube atrophy. *Physiol. Rep.* **2024**, *12*, e16011. [\[CrossRef\]](https://doi.org/10.14814/phy2.16011)
- 84. Martinesi, M.; Treves, C.; d'Albasio, G.; Bagnoli, S.; Bonanomi, A.G.; Stio, M. Vitamin D derivatives induce apoptosis and downregulate ICAM-1 levels in peripheral blood mononuclear cells of inflammatory bowel disease patients. *Inflamm. Bowel Dis.* **2008**, *14*, 597–604. [\[CrossRef\]](https://doi.org/10.1002/ibd.20354)
- 85. Cohen, M.S.; Mesler, D.E.; Snipes, R.G.; Gray, T. 1, 25-Dihydroxyvitamin D3 activates secretion of hydrogen peroxide by human monocytes. *J. Immunol.* **1986**, *136*, 1049–1053. [\[CrossRef\]](https://doi.org/10.4049/jimmunol.136.3.1049) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/3079794)
- 86. Haas, M.J.; Jafri, M.; Wehmeier, K.R.; Onstead-Haas, L.M.; Mooradian, A.D. Inhibition of endoplasmic reticulum stress and oxidative stress by vitamin D in endothelial cells. *Free Radic. Biol. Med.* **2016**, *99*, 1–10. [\[CrossRef\]](https://doi.org/10.1016/j.freeradbiomed.2016.07.020)
- 87. Hussien, N.I.; El-Wakeel, H.S.; Souror, S.M.; Ahmed, I.A. Alleviation of cardiac mitochondrial dysfunction and oxidative stress underlies the protective effect of vitamin D in chronic stress-induced cardiac dysfunction in rats. *Gen. Physiol. Biophys.* **2019**, *38*, 51–61. [\[CrossRef\]](https://doi.org/10.4149/gpb_2018036) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30761994)
- Lee, K.J.; Wright, G.; Bryant, H.; Wiggins, L.A.; Dal Zotto, V.L.; Schuler, M.; Malozzi, C.; Cohen, M.V.; Gassman, N.R. Cytoprotective Effect of Vitamin D on Doxorubicin-Induced Cardiac Toxicity in Triple Negative Breast Cancer. *Int. J. Mol. Sci.* **2021**, *22*, 7439. [\[CrossRef\]](https://doi.org/10.3390/ijms22147439) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34299059)
- 89. Wang, H.Q.; Zhao, M.X.; Hong, S.C.; He, X.; Tao, L.; Tong, C.C.; Jing, G.; Xu, D.X.; Chen, X. 1,25(OH)(2)D(3) alleviates oxidative stress and inflammation through up-regulating HMGCS2 in DSS-induced colitis. *Int. Immunopharmacol.* **2023**, *125*, 111131. [\[CrossRef\]](https://doi.org/10.1016/j.intimp.2023.111131)
- 90. Strickland, J.M.; Wisnieski, L.; Mavangira, V.; Sordillo, L.M. Serum Vitamin D Is Associated with Antioxidant Potential in Peri-Parturient Cows. *Antioxidants* **2021**, *10*, 1420. [\[CrossRef\]](https://doi.org/10.3390/antiox10091420)
- 91. Zhou, X.; Zou, Y.; Xu, Y.; Zhang, Z.; Wu, Y.; Cao, J.; Qiu, B.; Qin, X.; Han, D.; Piao, X.; et al. Dietary Supplementation of 25-Hydroxyvitamin D(3) Improves Growth Performance, Antioxidant Capacity and Immune Function in Weaned Piglets. *Antioxidants* **2022**, *11*, 1750. [\[CrossRef\]](https://doi.org/10.3390/antiox11091750)
- 92. Zhang, L.; Long, S.; Wang, H.; Piao, X. Dietary 25-hydroxycholecalciferol modulates gut microbiota and improves the growth, meat quality, and antioxidant status of growing-finishing pigs. *Front. Microbiol.* **2022**, *13*, 1095509. [\[CrossRef\]](https://doi.org/10.3389/fmicb.2022.1095509)
- 93. Wang, Y.; Liu, J.; Xiao, H.; Sun, H.; Hu, H.; Ma, X.; Zhang, A.; Zhou, H. Dietary intakes of vitamin D promote growth performance and disease resistance in juvenile grass carp (*Ctenopharyngodon idella*). *Fish. Physiol. Biochem.* **2024**, *50*, 1189–1203. [\[CrossRef\]](https://doi.org/10.1007/s10695-024-01330-9) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38427282)
- 94. Norris, K.C.; Olabisi, O.; Barnett, M.E.; Meng, Y.X.; Martins, D.; Obialo, C.; Lee, J.E.; Nicholas, S.B. The Role of Vitamin D and Oxidative Stress in Chronic Kidney Disease. *Int. J. Environ. Res. Public Health* **2018**, *15*, 2701. [\[CrossRef\]](https://doi.org/10.3390/ijerph15122701)
- 95. Schmitt, G.; Labdouni, S.; Soulimani, R.; Delamare, C.; Bouayed, J. Oxidative stress status and vitamin D levels of asymptomatic to mild symptomatic COVID-19 infections during the third trimester of pregnancy: A retrospective study in Metz, France. *J. Med. Virol.* **2022**, *94*, 2167–2173. [\[CrossRef\]](https://doi.org/10.1002/jmv.27606)
- 96. Golabi, S.; Ghasemi, S.; Adelipour, M.; Bagheri, R.; Suzuki, K.; Wong, A.; Seyedtabib, M.; Naghashpour, M. Oxidative Stress and Inflammatory Status in COVID-19 Outpatients: A Health Center-Based Analytical Cross-Sectional Study. *Antioxidants* **2022**, *11*, 606. [\[CrossRef\]](https://doi.org/10.3390/antiox11040606)
- 97. Reino-Gelardo, S.; Palop-Cervera, M.; Aparisi-Valero, N.; Espinosa-San Miguel, I.; Lozano-Rodriguez, N.; Llop-Furquet, G.; Sanchis-Artero, L.; Cortes-Castell, E.; Rizo-Baeza, M.; Cortes-Rizo, X. Effect of an Immune-Boosting, Antioxidant and Anti-Inflammatory Food Supplement in Hospitalized COVID-19 Patients: A Prospective Randomized Pilot Study. *Nutrients* **2023**, *15*, 1736. [\[CrossRef\]](https://doi.org/10.3390/nu15071736)
- 98. Yang, H.; Chen, J.S.; Luo, X.Y.; Wang, H. Efficacy and safety profile of antioxidants in the treatment of atopic dermatitis: A systematic review and meta-analysis of randomized controlled trials. *Dermatol. Ther.* **2022**, *35*, e15549. [\[CrossRef\]](https://doi.org/10.1111/dth.15549)
- 99. Mohammadzadeh Honarvar, N.; Samadi, M.; Seyedi Chimeh, M.; Gholami, F.; Bahrampour, N.; Jalali, M.; Effatpanah, M.; Yekaninejad, M.S.; Abdolahi, M.; Chamari, M. Effect of Vitamin D on Paraxonase-1, Total Antioxidant Capacity, and 8-Isoprostan in Children with Attention Deficit Hyperactivity Disorder. *Int. J. Clin. Pract.* **2022**, *2022*, 4836731. [\[CrossRef\]](https://doi.org/10.1155/2022/4836731) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35685610)
- 100. Motamed, S.; Nikooyeh, B.; Anari, R.; Motamed, S.; Mokhtari, Z.; Neyestani, T. The effect of vitamin D supplementation on oxidative stress and inflammatory biomarkers in pregnant women: A systematic review and meta-analysis of clinical trials. *BMC Pregnancy Childbirth* **2022**, *22*, 816. [\[CrossRef\]](https://doi.org/10.1186/s12884-022-05132-w) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36335311)
- 101. Perichart-Perera, O.; Avila-Sosa, V.; Solis-Paredes, J.M.; Montoya-Estrada, A.; Reyes-Munoz, E.; Rodriguez-Cano, A.M.; Gonzalez-Leyva, C.P.; Sanchez-Martinez, M.; Estrada-Gutierrez, G.; Irles, C. Vitamin D Deficiency, Excessive Gestational Weight Gain, and Oxidative Stress Predict Small for Gestational Age Newborns Using an Artificial Neural Network Model. *Antioxidants* **2022**, *11*, 574. [\[CrossRef\]](https://doi.org/10.3390/antiox11030574)
- 102. Khanolkar, S.; Hirani, S.; Mishra, A.; Vardhan, S.; Hirani, S.; Prasad, R.; Wanjari, M. Exploring the Role of Vitamin D in Atherosclerosis and Its Impact on Cardiovascular Events: A Comprehensive Review. *Cureus* **2023**, *15*, e42470. [\[CrossRef\]](https://doi.org/10.7759/cureus.42470)
- 103. Marefati, N.; Beheshti, F.; Anaeigoudari, A.; Alipour, F.; Shafieian, R.; Akbari, F.; Pirasteh, M.; Mahmoudabady, M.; Salmani, H.; Mawdodi, S.; et al. The effects of vitamin D on cardiovascular damage induced by lipopolysaccharides in rats. *J. Cardiovasc. Thorac. Res.* **2023**, *15*, 106–115. [\[CrossRef\]](https://doi.org/10.34172/jcvtr.2023.31719)
- 104. Forstermann, U.; Munzel, T. Endothelial nitric oxide synthase in vascular disease: From marvel to menace. *Circulation* **2006**, *113*, 1708–1714. [\[CrossRef\]](https://doi.org/10.1161/CIRCULATIONAHA.105.602532)
- 105. Cojic, M.; Kocic, R.; Klisic, A.; Kocic, G. The Effects of Vitamin D Supplementation on Metabolic and Oxidative Stress Markers in Patients With Type 2 Diabetes: A 6-Month Follow Up Randomized Controlled Study. *Front. Endocrinol.* **2021**, *12*, 610893. [\[CrossRef\]](https://doi.org/10.3389/fendo.2021.610893) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34489860)
- 106. Alfaqih, M.A.; Ababneh, E.; Mhedat, K.; Allouh, M.Z. Vitamin D Reduces the Activity of Adenosine Deaminase and Oxidative Stress in Patients with Type Two Diabetes Mellitus. *Mol. Nutr. Food Res.* **2024**, *68*, e2300870. [\[CrossRef\]](https://doi.org/10.1002/mnfr.202300870) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38816753)
- 107. Yu, L.; Zhu, Y.; Geng, L.; Xu, Y.; Zhao, M. Effect of different nutrients on blood glucose, inflammatory response and oxidative stress in gestational diabetes mellitus: A network meta-analysis. *Br. J. Nutr.* **2024**, *131*, 1513–1527. [\[CrossRef\]](https://doi.org/10.1017/S0007114523003069)
- 108. Anandabaskar, N.; Selvarajan, S.; Dkhar, S.A.; Kamalanathan, S.K.; Tamilarasu, K.; Bobby, Z. Effect of Vitamin D Supplementation on Vascular Functions and Oxidative Stress in Type 2 Diabetic Patients with Vitamin D Deficiency. *Indian. J. Endocrinol. Metab.* **2017**, *21*, 555–563. [\[CrossRef\]](https://doi.org/10.4103/ijem.IJEM_140_17) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28670540)
- 109. Zhang, Q.; Cheng, Y.; He, M.; Li, T.; Ma, Z.; Cheng, H. Effect of various doses of vitamin D supplementation on pregnant women with gestational diabetes mellitus: A randomized controlled trial. *Exp. Ther. Med.* **2016**, *12*, 1889–1895. [\[CrossRef\]](https://doi.org/10.3892/etm.2016.3515)
- 110. Iqbal, S.; Khan, S.; Naseem, I. Antioxidant Role of Vitamin D in Mice With Alloxan-Induced Diabetes. *Can. J. Diabetes* **2018**, *42*, 412–418. [\[CrossRef\]](https://doi.org/10.1016/j.jcjd.2017.10.032) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29217217)
- 111. Tiwari, B.K.; Pandey, K.B.; Abidi, A.B.; Rizvi, S.I. Markers of Oxidative Stress during Diabetes Mellitus. *J. Biomark.* **2013**, *2013*, 378790. [\[CrossRef\]](https://doi.org/10.1155/2013/378790)
- 112. Nikooyeh, B.; Neyestani, T.R. Oxidative stress, type 2 diabetes and vitamin D: Past, present and future. *Diabetes Metab. Res. Rev.* **2016**, *32*, 260–267. [\[CrossRef\]](https://doi.org/10.1002/dmrr.2718)
- 113. Rastegar-Moghaddam, S.H.; Hosseini, M.; Alipour, F.; Rajabian, A.; Ebrahimzadeh Bideskan, A. The effects of vitamin D on learning and memory of hypothyroid juvenile rats and brain tissue acetylcholinesterase activity and oxidative stress indicators. *Naunyn Schmiedebergs Arch. Pharmacol.* **2022**, *395*, 337–351. [\[CrossRef\]](https://doi.org/10.1007/s00210-021-02195-y)
- 114. Maes, M.; Galecki, P.; Chang, Y.S.; Berk, M. A review on the oxidative and nitrosative stress (O&NS) pathways in major depression and their possible contribution to the (neuro)degenerative processes in that illness. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **2011**, *35*, 676–692. [\[CrossRef\]](https://doi.org/10.1016/j.pnpbp.2010.05.004) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20471444)
- 115. Aslan, C.; Aslankoc, R.; Ozmen, O.; Suluk, B.N.; Kavrik, O.; Gumral, N. Protective effect of vitamin D on learning and memory impairment in rats induced by high fructose corn syrup. *Behav. Brain Res.* **2024**, *459*, 114763. [\[CrossRef\]](https://doi.org/10.1016/j.bbr.2023.114763)
- 116. Janjusevic, M.; Gagno, G.; Fluca, A.L.; Padoan, L.; Beltrami, A.P.; Sinagra, G.; Moretti, R.; Aleksova, A. The peculiar role of vitamin D in the pathophysiology of cardiovascular and neurodegenerative diseases. *Life Sci.* **2022**, *289*, 120193. [\[CrossRef\]](https://doi.org/10.1016/j.lfs.2021.120193) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34864062)
- 117. Brewer, L.D.; Thibault, V.; Chen, K.-C.; Langub, M.C.; Landfield, P.W.; Porter, N.M. Vitamin D hormone confers neuroprotection in parallel with downregulation of L-type calcium channel expression in hippocampal neurons. *J. Neurosci.* **2001**, *21*, 98–108. [\[CrossRef\]](https://doi.org/10.1523/JNEUROSCI.21-01-00098.2001)
- 118. Thibault, O.; Landfield, P.W. Increase in single L-type calcium channels in hippocampal neurons during aging. *Science* **1996**, *272*, 1017–1020. [\[CrossRef\]](https://doi.org/10.1126/science.272.5264.1017)
- 119. Porter, N.M.; Thibault, O.; Thibault, V.; Chen, K.-C.; Landfield, P.W. Calcium channel density and hippocampal cell death with age in long-term culture. *J. Neurosci.* **1997**, *17*, 5629–5639. [\[CrossRef\]](https://doi.org/10.1523/JNEUROSCI.17-14-05629.1997)
- 120. Gezen-Ak, D.; Dursun, E.; Yilmazer, S. Vitamin D inquiry in hippocampal neurons: Consequences of vitamin D-VDR pathway disruption on calcium channel and the vitamin D requirement. *Neurol. Sci.* **2013**, *34*, 1453–1458. [\[CrossRef\]](https://doi.org/10.1007/s10072-012-1268-6) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23250517)
- 121. Sutherland, M.K.; Somerville, M.J.; Yoong, L.K.; Bergeron, C.; Haussler, M.R.; McLachlan, D.R.C. Reduction of vitamin D hormone receptor mRNA levels in Alzheimer as compared to Huntington hippocampus: Correlation with calbindin-28k mRNA levels. *Mol. Brain Res.* **1992**, *13*, 239–250. [\[CrossRef\]](https://doi.org/10.1016/0169-328X(92)90032-7)
- 122. Almeras, L.; Eyles, D.; Benech, P.; Laffite, D.; Villard, C.; Patatian, A.; Boucraut, J.; Mackay-Sim, A.; McGrath, J.; Féron, F. Developmental vitamin D deficiency alters brain protein expression in the adult rat: Implications for neuropsychiatric disorders. *Proteomics* **2007**, *7*, 769–780. [\[CrossRef\]](https://doi.org/10.1002/pmic.200600392) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17295352)
- 123. Cui, C.; Wang, C.; Jin, F.; Yang, M.; Kong, L.; Han, W.; Jiang, P. Calcitriol confers neuroprotective effects in traumatic brain injury by activating Nrf2 signaling through an autophagy-mediated mechanism. *Mol. Med.* **2021**, *27*, 118. [\[CrossRef\]](https://doi.org/10.1186/s10020-021-00377-1) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34556021)
- 124. Janosa, G.; Pandur, E.; Pap, R.; Horvath, A.; Sipos, K. Interplay of Vitamin D, Unfolded Protein Response, and Iron Metabolism in Neuroblastoma Cells: A Therapeutic Approach in Neurodegenerative Conditions. *Int. J. Mol. Sci.* **2023**, *24*, 6883. [\[CrossRef\]](https://doi.org/10.3390/ijms242316883) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38069206)
- 125. Sepidarkish, M.; Farsi, F.; Akbari-Fakhrabadi, M.; Namazi, N.; Almasi-Hashiani, A.; Maleki Hagiagha, A.; Heshmati, J. The effect of vitamin D supplementation on oxidative stress parameters: A systematic review and meta-analysis of clinical trials. *Pharmacol. Res.* **2019**, *139*, 141–152. [\[CrossRef\]](https://doi.org/10.1016/j.phrs.2018.11.011) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30447293)
- 126. Cui, C.; Xu, P.; Li, G.; Qiao, Y.; Han, W.; Geng, C.; Liao, D.; Yang, M.; Chen, D.; Jiang, P. Vitamin D receptor activation regulates microglia polarization and oxidative stress in spontaneously hypertensive rats and angiotensin II-exposed microglial cells: Role of renin-angiotensin system. *Redox Biol.* **2019**, *26*, 101295. [\[CrossRef\]](https://doi.org/10.1016/j.redox.2019.101295) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31421410)
- 127. Mansouri, F.; Ghanbari, H.; Marefati, N.; Arab, Z.; Salmani, H.; Beheshti, F.; Hosseini, M. Protective effects of vitamin D on learning and memory deficit induced by scopolamine in male rats: The roles of brain-derived neurotrophic factor and oxidative stress. *Naunyn Schmiedebergs Arch. Pharmacol.* **2021**, *394*, 1451–1466. [\[CrossRef\]](https://doi.org/10.1007/s00210-021-02062-w) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33649977)
- 128. Mohanad, M.; Mohamed, S.K.; Aboulhoda, B.E.; Ahmed, M.A.E. Neuroprotective effects of vitamin D in an Alzheimer's disease rat model: Improvement of mitochondrial dysfunction via calcium/calmodulin-dependent protein kinase kinase 2 activation of Sirtuin1 phosphorylation. *Biofactors* **2024**, *50*, 371–391. [\[CrossRef\]](https://doi.org/10.1002/biof.2013) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37801071)
- 129. Kouba, B.R.; Camargo, A.; Rodrigues, A.L.S. Neuroinflammation in Alzheimer's disease: Potential beneficial effects of vitamin D. *Metab. Brain Dis.* **2023**, *38*, 819–829. [\[CrossRef\]](https://doi.org/10.1007/s11011-023-01188-5) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36862275)
- 130. Mehri, N.; Haddadi, R.; Ganji, M.; Shahidi, S.; Soleimani Asl, S.; Taheri Azandariani, M.; Ranjbar, A. Effects of vitamin D in an animal model of Alzheimer's disease: Behavioral assessment with biochemical investigation of Hippocampus and serum. *Metab. Brain Dis.* **2020**, *35*, 263–274. [\[CrossRef\]](https://doi.org/10.1007/s11011-019-00529-7)
- 131. Lin, J.; Niu, Z.; Xue, Y.; Gao, J.; Zhang, M.; Li, M.; Peng, Y.; Zhang, S.; Li, W.; Zhang, Q.; et al. Chronic vitamin D(3) supplementation alleviates cognition impairment via inhibition of oxidative stress regulated by PI3K/AKT/Nrf2 in APP/PS1 transgenic mice. *Neurosci. Lett.* **2022**, *783*, 136725. [\[CrossRef\]](https://doi.org/10.1016/j.neulet.2022.136725)
- 132. Verma, S.; Singh, P.; Khurana, S.; Ganguly, N.K.; Kukreti, R.; Saso, L.; Rana, D.S.; Taneja, V.; Bhargava, V. Implications of oxidative stress in chronic kidney disease: A review on current concepts and therapies. *Kidney Res. Clin. Pract.* **2021**, *40*, 183–193. [\[CrossRef\]](https://doi.org/10.23876/j.krcp.20.163) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34078023)
- 133. Tonetti, M.S.; Van Dyke, T.E.; Working Group 1 of the Joint EFP/AAP Workshop. Periodontitis and atherosclerotic cardiovascular disease: Consensus report of the Joint EFP/AAPWorkshop on Periodontitis and Systemic Diseases. *J. Periodontol.* **2013**, *84*, S24–S29. [\[CrossRef\]](https://doi.org/10.1902/jop.2013.1340019) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23631582)
- 134. Ahmed Sharif, D. The Effectiveness of Vitamin D Supplementation on Oxidative and Inflammatory Markers in Patients Suffering from End-stage Renal Disease, a Randomized Controlled Trial. *Cell Mol. Biol.* **2022**, *68*, 7–15. [\[CrossRef\]](https://doi.org/10.14715/cmb/2022.68.5.2) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36029489)
- 135. Ganimusa, I.; Chew, E.; Lu, E.M. Vitamin D Deficiency, Chronic Kidney Disease and Periodontitis. *Medicina* **2024**, *60*, 420. [\[CrossRef\]](https://doi.org/10.3390/medicina60030420) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38541146)
- 136. Matsumoto, A.K.; Maes, M.; Michelin, A.P.; Soares, A.E.; Semeao, L.O.; Godeny, P.; Venturini, D.; Barbosa, D.S.; Delfino, V.D.A. Vitamin D deficiency is not associated with increased oxidative stress in chronic kidney disease pre-dialysis patients. *J. Bras. Nefrol.* **2020**, *42*, 420–428. [\[CrossRef\]](https://doi.org/10.1590/2175-8239-jbn-2019-0156) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32406474)
- 137. Park, J.S.; Na, H.J.; Kim, Y.J. The anti-aging effect of vitamin D and vitamin D receptor in Drosophila midgut. *Aging* **2024**, *16*, 2005–2025. [\[CrossRef\]](https://doi.org/10.18632/aging.205518) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38329439)
- 138. Kazemian, E.; Akbari, M.E.; Moradi, N.; Gharibzadeh, S.; Amouzegar, A.; Jamshidi-Naeini, Y.; Mondul, A.M.; Khademolmele, M.; Ghodoosi, N.; Zarins, K.R.; et al. Effect of vitamin D receptor polymorphisms on plasma oxidative stress and apoptotic biomarkers among breast cancer survivors supplemented vitamin D3. *Eur. J. Cancer Prev.* **2020**, *29*, 433–444. [\[CrossRef\]](https://doi.org/10.1097/CEJ.0000000000000576) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32740169)
- 139. Shang, Q.X.; Yang, Y.S.; Zhang, H.L.; Cheng, Y.P.; Lu, H.; Yuan, Y.; Chen, L.Q.; Ji, A.F. Vitamin D receptor induces oxidative stress to promote esophageal squamous cell carcinoma proliferation via the p53 signaling pathway. *Heliyon* **2024**, *10*, e23832. [\[CrossRef\]](https://doi.org/10.1016/j.heliyon.2023.e23832) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38234882)
- 140. Seraphin, G.; Rieger, S.; Hewison, M.; Capobianco, E.; Lisse, T.S. The impact of vitamin D on cancer: A mini review. *J. Steroid Biochem. Mol. Biol.* **2023**, *231*, 106308. [\[CrossRef\]](https://doi.org/10.1016/j.jsbmb.2023.106308)
- 141. Wimalawansa, S.J. Vitamin D Deficiency: Effects on Oxidative Stress, Epigenetics, Gene Regulation, and Aging. *Biology* **2019**, *8*, 30. [\[CrossRef\]](https://doi.org/10.3390/biology8020030)
- 142. Slominski, A.T.; Kim, T.K.; Li, W.; Postlethwaite, A.; Tieu, E.W.; Tang, E.K.Y.; Tuckey, R.C. Detection of novel CYP11A1-derived secosteroids in the human epidermis and serum and pig adrenal gland. *Sci. Rep.* **2015**, *5*, 14875. [\[CrossRef\]](https://doi.org/10.1038/srep14875) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26445902)
- 143. Karampinis, E.; Aloizou, A.M.; Zafiriou, E.; Bargiota, A.; Skaperda, Z.; Kouretas, D.; Roussaki-Schulze, A.V. Non-Melanoma Skin Cancer and Vitamin D: The "Lost Sunlight" Paradox and the Oxidative Stress Explanation. *Antioxidants* **2023**, *12*, 1107. [\[CrossRef\]](https://doi.org/10.3390/antiox12051107) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37237973)
- 144. Christakos, S.; Ajibade, D.V.; Dhawan, P.; Fechner, A.J.; Mady, L.J. Vitamin D: Metabolism. *Endocrinol. Metab. Clin. N. Am.* **2010**, *39*, 243–253. [\[CrossRef\]](https://doi.org/10.1016/j.ecl.2010.02.002) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20511049)
- 145. Deinum, J.; Groenewoud, H.; van der Wilt, G.J.; Lenzini, L.; Rossi, G.P. Adrenal venous sampling: Cosyntropin stimulation or not? *Eur. J. Endocrinol.* **2019**, *181*, D15–D26. [\[CrossRef\]](https://doi.org/10.1530/EJE-18-0844) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31176302)
- 146. Reichrath, J.; Lindqvist, P.G.; FR, D.E.G.; Pilz, S.; Kimball, S.M.; Grant, W.B.; Holick, M.F. A Critical Appraisal of the Recent Reports on Sunbeds from the European Commission's Scientific Committee on Health, Environmental and Emerging Risks and from the World Health Organization. *Anticancer Res.* **2018**, *38*, 1111–1120. [\[CrossRef\]](https://doi.org/10.21873/anticanres.12330) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29374748)
- 147. Verstuyf, A.; Carmeliet, G.; Bouillon, R.; Mathieu, C. Vitamin D: A pleiotropic hormone. *Kidney Int.* **2010**, *78*, 140–145. [\[CrossRef\]](https://doi.org/10.1038/ki.2010.17) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20182414)
- 148. da Silva, D.C.; Andrade, P.B.; Valentao, P.; Pereira, D.M. Neurotoxicity of the steroidal alkaloids tomatine and tomatidine is RIP1 kinase- and caspase-independent and involves the eIF2alpha branch of the endoplasmic reticulum. *J. Steroid Biochem. Mol. Biol.* **2017**, *171*, 178–186. [\[CrossRef\]](https://doi.org/10.1016/j.jsbmb.2017.03.009) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28300624)
- 149. Tuckey, R.C.; Cheng, C.Y.S.; Slominski, A.T. The serum vitamin D metabolome: What we know and what is still to discover. *J. Steroid Biochem. Mol. Biol.* **2019**, *186*, 4–21. [\[CrossRef\]](https://doi.org/10.1016/j.jsbmb.2018.09.003)
- 150. Bikle, D.; Bouillon, R.; Thadhani, R.; Schoenmakers, I. Vitamin D metabolites in captivity? Should we measure free or total 25(OH)D to assess vitamin D status? *J. Steroid Biochem. Mol. Biol.* **2017**, *173*, 105–116. [\[CrossRef\]](https://doi.org/10.1016/j.jsbmb.2017.01.007)
- 151. Aranow, C. Vitamin D and the immune system. *J. Investig. Med.* **2011**, *59*, 881–886. [\[CrossRef\]](https://doi.org/10.2310/JIM.0b013e31821b8755)
- 152. Taskinen, M.R.; Soderlund, S.; Bogl, L.H.; Hakkarainen, A.; Matikainen, N.; Pietilainen, K.H.; Rasanen, S.; Lundbom, N.; Bjornson, E.; Eliasson, B.; et al. Adverse effects of fructose on cardiometabolic risk factors and hepatic lipid metabolism in subjects with abdominal obesity. *J. Intern. Med.* **2017**, *282*, 187–201. [\[CrossRef\]](https://doi.org/10.1111/joim.12632) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28548281)

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.