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In vitro activity of macrolides and lincosamides against oral streptococci: a therapeutic alternative in prophylaxis for infective endocarditis

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Susceptibility to macrolides and lincosamides was tested in a total of 446 strains of oral streptococci belonging to eleven species, all isolated from dental plaque and/or saliva in 1991. Minimum inhibitory concentrations (MIC) were determined by the double serial dilutions method in agar. Clindamycin was the most effective antibiotic against all species tested. The results of nearly all parameters (range, mean, MIC₅₀, MIC₉₀) in all species showed erythromycin to be less effective than josamycin, making the latter antibiotic a possible alternative to erythromycin in prophylaxis for infective endocarditis caused by oral streptococci, not only in patients allergic to penicillin, but also in all patients with low-risk lesions, who will be treated with low-risk dental procedures. In these latter patients, antibiotic prophylaxis may be indicated if, after careful evaluation of the individual's situation, no other alternatives are available.

Key words: Oral streptococci; In vitro susceptibility; Endocarditis; Macrolides; Lincosamides

Introduction

Streptococci of the so-called *viridans* group are, together with staphylococci, the bacteria most frequently involved in the etiology of infective endocarditis. Approximately 40% of all subacute forms of the disease are caused by these microorganisms [1–4]. Oral streptococci are the largest subgroup within

the *viridans* group. Several studies have identified *Streptococcus sanguis*, *S. mitior* and *S. mutans* as the microorganisms most frequently isolated in subacute infective endocarditis caused by *viridans* group bacteria [5,6]. These organisms usually cause left endocarditis, while right endocarditis is less frequent [4]. Adhesion factors, especially glycoconjugate molecules [5,7–10], have been implicated in their ability to colonize the valves and the endothelium of great vessels; these factors would also afford the bacteria some protection against opsonization and phagocytosis. However, the existence of endocarditis-causing oral streptococci without glycoconjugate suggests

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that other pathogenic mechanisms are also involved [11–13].

Because of the seriousness of infective endocarditis, there is widespread agreement that treatment should consist of orally administered bactericidal antibiotics, such as a combination of beta lactams and aminoglycosides. In patients allergic to the former, the antibiotics of choice are vancomycin and teicoplanin. In subjects with high-risk lesions (e.g., valvular prosthesis or certain heart diseases), and in high-risk procedures, the prophylactic measures are similar, although the dosages are different [14–16].

There is disagreement as to the best antibiotic prophylaxis in patients with low-risk lesions to be treated with low-risk procedures. Most regimens are based on experimental models with bactericidal antibiotics alone or in combination [17–18]. Experimental studies with bacteriostatic antibiotics have provided conflicting results [19,20].

The objective of the present study was to analyze the susceptibility of oral streptococci to a group of antibiotics (macrolides and lincosamides) in search of a potential alternative for prophylaxis against infective endocarditis caused by these microorganisms in patients with low-risk lesions, who will be treated with low-risk dental procedures.

Materials and methods

Antimicrobial drugs

Standard powders were obtained from different manufacturers: erythromycin (Antibioticos, Madrid), lincomycin and clindamycin (Upjohn Farmoquímica, Madrid), spiramycin (Rhône-Poulenc, Alcorcon, Madrid), acetyl spiramycin (Hubber, Barcelona), josamycin (Ferrer International, Barcelona), roxitromycin (Russell, Madrid), oleandomycin and azitromycin (Pfizer, Madrid) and diacetylmidecamycin (Menarini, Barcelona).

Isolates

The microorganisms were isolated from dental plaque or saliva of different patients throughout the year 1991. All strains were fixed by lyophilization and identified according to Hardie's [21] or Loeche's [22] criteria. In all, 60 different strains of

Streptococcus mutans, 36 of *S. sobrinus*, 10 of *S. cricetus*, 10 of *S. macacae*, 10 of *S. rattus*, 56 of *S. mitior*, 40 of *S. mitis*, 68 of *S. salivarius*, 58 of *S. milleri*, 76 of *S. sanguis* and 22 of *S. oralis* were studied.

Susceptibility testing

The antibiotics were tested at concentrations ranging from 0.0015 to 64 µg/ml. The minimum inhibitory concentration (MIC) was determined using an agar dilution method, Steer's replicator and Wilkins-Chalgren medium (Difco Laboratories, Madrid). As the inoculum we used 10^5 to 10^6 cfu/ml, obtained from a 24-h culture grown in trypticase soy broth medium without dextrose (Scott Laboratories, Madrid). Plates were inoculated at 37°C in an anaerobic atmosphere containing 85% N₂, 10% H₂ and 5% CO₂, and readings were taken after 48 h [23]. The MIC was defined as the lowest concentration of antibiotics that visibly inhibited growth of the microorganisms. Linear extrapolation from values obtained from the next higher and next lower values were used to express the exact values of MIC₅₀ and MIC₉₀.

As a control for intraobserver variability, replicate cultures were done of eight strains of each species represented in the present study by 18 or more strains, and two strains of each species represented by four to six strains. The results of these cultures were read by independent observers.

Statistical procedure

Snedecor's F statistic was calculated for the statistical analysis of quantitative variables.

Results

The susceptibilities of oral streptococci to antibiotics are shown in Table 1. The highest mean value was recorded for *S. milleri* susceptibility to oleandomycin (14.24 µg/ml), followed by *S. sobrinus* susceptibility to acetylspiramycin (9.27 µg/ml). The lowest mean values were found for *S. sobrinus* (0.08 µg/ml) and *S. mutans* susceptibility (0.11 µg/ml) to clindamycin. The highest MIC₉₀ were those of ace-

TABLE 1
Susceptibility of 446 strains of oral streptococci to eight macrolides, lincomycin and clindamycin

Antimicrobial agents	<i>S. mutans</i> (60) ^a	<i>S. sobrinus</i> (36)	<i>S. sanguis</i> (76)	<i>S. mitior</i> (56)	<i>S. mitis</i> (40)	<i>S. salivarius</i> (68)	<i>S. milleri</i> (58)	Miscellaneous ^b (52)
Oleandomycin								
Range	0.25–4	1–8	0.12–32	0.06–32	0.25–32	0.5–32	0.5–32	0.25–8
\bar{x}	1.38	4.66	3.28	3.67	7.2	9.75	14.24	1.99
MIC ₅₀	0.67	3.01	0.45	0.7	0.9	1.79	1.5	1.2
MIC ₉₀	2	6.9	4.77	6.3	21.33	25.2	13.21	3.6
Erythromycin								
Range	0.03–16	0.06–4	0.03–4	0.03–16	0.06–16	0.05–16	0.06–16	0.06–8
\bar{x}	1.28	0.52	0.75	2.01	2.57	2.56	4.61	0.73
MIC ₅₀	0.08	0.12	0.09	0.06	0.12	0.1	0.15	0.09
MIC ₉₀	1	1.1	2.01	4.8	5.66	5.79	13.1	0.87
Spiramycin								
Range	0.25–1	0.5–16	0.25–16	0.25–16	0.25–16	0.25–16	0.25–16	0.25–16
\bar{x}	0.58	4.5	1.15	2.4	2.48	5.23	5.95	3.43
MIC ₅₀	0.37	0.75	0.36	0.43	0.58	0.68	0.48	1.23
MIC ₉₀	0.85	5.29	1.74	4.79	4	12.11	13.42	8.9
Acetylspiramycin								
Range	1–4	0.5–32	0.25–16	0.25–32	1–32	0.25–32	0.25–32	0.5–32
\bar{x}	1.6	9.27	1.84	3.72	5	6.76	4.41	8.14
MIC ₅₀	0.8	1	0.6	0.7	1.33	0.57	0.48	3.2
MIC ₉₀	3	24.8	3.2	6.42	4	22.9	24.26	17.6
Diacetylmidecamycin								
Range	0.03–1	0.25–2	0.03–16	0.03–16	0.12–16	0.03–16	0.03–16	0.03–8
\bar{x}	0.39	1.05	1.75	1.81	2.6	2.67	2.83	1.44
MIC ₅₀	0.18	0.63	0.10	0.08	0.4	0.5	0.31	0.7
MIC ₉₀	0.78	1.7	1.2	4.8	4	3.2	10.2	2.4
Josamycin								
Range	0.03–1	0.25–2	0.12–1	0.12–16	0.12–16	0.06–16	0.12–16	0.12–1
\bar{x}	0.3	0.8	0.36	1.65	1.55	1.6	2.62	0.52
MIC ₅₀	0.18	0.4	0.18	0.17	0.36	0.31	0.4	0.2
MIC ₉₀	0.45	1.55	0.64	2.4	2	2.6	6.8	0.8
Roxitromycin								
Range	0.03–0.5	0.25–2	0.06–16	0.06–16	0.06–16	0.03–16	0.06–16	0.06–4
\bar{x}	0.21	0.9	1.54	2.3	2.5	2.37	4.4	0.97
MIC ₅₀	0.12	0.44	0.1	0.14	0.18	0.12	0.37	0.42
MIC ₉₀	0.4	1.64	3.2	8.54	8	5.8	12.68	1.8
Azitromycin								
Range	0.12–1	0.5–16	0.12–16	0.12–16	0.12–16	0.12–16	0.25–16	0.12–16
\bar{x}	0.36	4.16	1.86	2.03	2.7	4.8	5.72	2.5
MIC ₅₀	0.18	1.5	0.27	0.20	0.25	0.5	0.63	0.3
MIC ₉₀	0.70	8.8	4.4	4.84	8	12.11	13.1	6.4
Lincomycin								
Range	0.06–0.05	0.25–1	0.12–4	0.06–4	0.06–4	0.06–4	0.06–4	0.06–8
\bar{x}	0.16	9.62	0.65	0.52	0.63	1.47	1.05	0.64
MIC ₅₀	0.09	0.40	0.16	0.23	0.29	0.41	0.53	0.18
MIC ₉₀	0.23	0.87	1.6	0.6	1	2.3	2.55	1.2
Clindamycin								
Range	0.015–1	0.015–0.12	0.03–2	0.03–4	0.03–4	0.03–4	0.03–4	0.03–4
\bar{x}	0.11	0.08	0.22	0.37	0.55	0.29	0.57	0.79
MIC ₅₀	0.035	0.14	0.07	0.05	0.072	0.08	0.08	0.04
MIC ₉₀	0.18	0.23	0.4	0.3	1	0.45	1.11	0.19

^aNumbers in parentheses are the number of strains studied.

^bThe miscellaneous group included 22 strains of *S. oralis*, and 10 each of *S. cricetus*, *S. macacae* and *S. rattus*.

tylspiramycin against *S. salivarius* (22.9 µg/ml) and *S. milleri* (24.26 µg/ml), while the lowest were those of clindamycin against *S. mutans* (0.18 µg/ml) and lincomycin and clindamycin against *S. mutans* and *S. sobrinus* respectively (0.23 µg/ml). The MIC₉₀ for all species were lowest for clindamycin, followed by lincomycin and josamycin. The exceptions were *S. sanguis*, in which the MIC₉₀ for josamycin (0.64 µg/ml) was lower than that for lincomycin (1.6 µg/ml), *S. sobrinus*, in which the MIC₉₀ for erythromycin (1.1 µg/ml) was lower than that for josamycin (1.55 µg/ml), and *S. mutans*, for which the MIC₉₀ for roxithromycin (0.4 µg/ml) was slightly lower than that for josamycin (0.45 µg/ml).

The results of statistical analysis showed significant differences between antibiotic activities in different species of oral streptococci, except for spiramycin, diacetylmidecamycin and clindamycin (Table 2). Significant differences in susceptibility were also found between the species, with the exception of *S. mitior* (Table 3).

Discussion

Several reviews have reported the frequencies of transient bacteremias caused by oral streptococci after dental procedures, or after simply chewing gum or paraffin, or brushing one's teeth. However, the risk of developing endocarditis is negligible [24,25] except in particularly susceptible patients, where it is nonetheless very low. Notwithstanding these slight probabilities, prophylactic antibiotic regimens have been developed for patients with low-risk lesions, due to undergo moderate-risk procedures (e.g., thor-

ough check-up and cleaning, scaling and extraction) [14–16]. Such regimens have involved a range of measures, from parenteral oral drug treatment [26] to oral amoxicillin [14–16]. These prophylactic measures have been effective in experimental models [17,27]. Erythromycin and clindamycin have been suggested as alternatives in patients allergic to penicillin [14–16]. As noted above, these measures are based on experimental studies in animal models, and the findings cannot be extrapolated to humans. Moreover, in addition to the almost complete absence of risk in dental patients, there is no clinical evidence of any beneficial effect of prophylactic antibiotic therapy [28]. Because the currently accepted norm seems excessively cautious [28], we sought equally reliable alternatives. If antibiotic prophylaxis for oral streptococcus-caused infective endocarditis is nonetheless being considered, the following facts should be taken into account:

- Some strains of oral streptococci are susceptible to penicillin (MIC ≤ 0.1 µg/ml). Against these microorganisms, prophylaxis with the same antibiotics as those to be used for treatment (or with similar antibiotics) seems unadvisable, as this would probably reduce the microorganism's susceptibility [29]. In fact, strains moderately resistant to penicillin have appeared in patients who received oral prophylaxis with this antibiotic [30].

- Penicillin-tolerant strains have been described for all the microorganisms tested in the present study [31] and in 20% of endocarditis patients [32]. These patients would probably require larger doses of antibiotics than are available via oral administration to attain bactericidal concentrations in serum. Such findings, although obtained in animal studies,

TABLE 2

Results of the comparisons of the activities of different antibiotics in oral streptococci

Antibiotic	Significance	Antibiotic	Significance
Oleandomycin	<i>P</i> < 0.01	Josamycin	<i>P</i> < 0.05
Erythromycin	<i>P</i> < 0.01	Roxithromycin	<i>P</i> < 0.05
Spiramycin	N.S.	Azithromycin	<i>P</i> < 0.01
Acetylspiramycin	<i>P</i> < 0.01	Lincomycin	<i>P</i> < 0.1
Diacetylmidecamycin	N.S.	Clindamycin	N.S.

TABLE 3

Results of the comparison of susceptibilities of oral streptococci to different antibiotics

Microorganism	Significance	Microorganism	Significance
<i>S. mutans</i>	<i>P</i> < 0.01	<i>S. mitis</i>	<i>P</i> < 0.05
<i>S. sobrinus</i>	<i>P</i> < 0.01	<i>S. salivarius</i>	<i>P</i> < 0.01
<i>S. sanguis</i>	<i>P</i> < 0.01	<i>S. milleri</i>	<i>P</i> < 0.01
<i>S. mitior</i>	N.S.	Miscellaneous	<i>P</i> < 0.01

should be taken into account, albeit with due caution [18,33–35].

- Some strains have been identified as nutritional variants; although apparently similar to other strains of a given species, they cause a greater tendency toward recurrence in patients with endocarditis [36–38].

- Nearly all species of oral streptococci produce glucanes and fructanes, which impede the penetration of beta lactams when bacterial vegetations lodge in the cardiac tissues [9,10].

This led us to test the hypothesis that macrolide and lincosamide antibiotics are effective in prophylaxis against infective endocarditis, not only in patients allergic to beta lactams, but in dental patients, after careful consideration of each individual case has ruled out present and future alternatives [28].

As shown by the results of the statistical analysis, not all species of oral streptococci were equally susceptible to macrolides and lincosamides, nor did all antibiotics show the same activity. Clindamycin was clearly the most effective antibiotic in vitro against all species we tested, and therefore represents the best alternative in prophylaxis against infective endocarditis caused by oral streptococci. This conclusion is supported by empirical studies of the prevention of endocarditis [15]. When clindamycin was compared with erythromycin, the result of the present study clearly favored the former. Although none of the species were unequivocally resistant to erythromycin, other macrolides were more active, e.g., roxitromycin against *S. mutans*, and josamycin against nearly all species. Erythromycin was less effective against *S. mutans* in the present study than in a previously published report [29]. This finding, together with the possibility of cross-resistance between erythromycin and other antibiotics (including tylosin, used as an additive in the meat packaging industry), and enhanced resistance caused by previous exposure [39], suggest that this antibiotic may be replaced, in prophylaxis against oral streptococci endocarditis, by macrolides such as josamycin, which involve fewer problems related to cross-resistance.

Oral streptococci are widely believed to be susceptible to antibiotics habitually used in clinical practice. Our findings partially confirm this assumption: inhibition of 90% of the strains of a given species

required relatively large amounts of some antibiotics, especially acetylspiramycin and spiramycin when tested against *S. sobrinus*, *S. salivarius* and *S. milleri*. The latter microorganism was the least susceptible to the antibiotics we tested, a logical finding in view of the heterogeneity of this group [40]. The other end of the spectrum was occupied by *S. mutans*, a highly susceptible species. As noted in a previous study [29], some antibiotics showed a notable decline in effectiveness between 1985 and 1989, e.g., erythromycin against *S. mutans*, acetylspiramycin and spiramycin against *S. sobrinus*. In contrast, other antibiotics have become more effective, e.g., erythromycin against *S. sobrinus*, and spiramycin against *S. mutans*. These observations exemplify the variations in susceptibility oral streptococci can display within a single environment.

In conclusion, clindamycin was the most effective of the antibiotics tested in vitro against oral streptococci. Although there is no experimental evidence for the usefulness of clindamycin as an antibiotic prophylactic against infective endocarditis caused by streptococci, in subjects with low-risk lesions subjected to low-risk dental procedures, this agent may be the drug of choice. Josamycin could be used as the antibiotic of second choice, given that our findings showed it to be more effective than erythromycin.

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References

- 1 Bayliss R, Clark C, Oakley CM et al. The microbiology and pathogenesis of infective endocarditis. *Br Heart J* 1983;50:513–519.
- 2 Kramer HN, Burgeois M, Liersch R et al. Current clinical aspects of bacterial endocarditis in infancy, childhood and adolescence. *Eur J Pediatr* 1983;140:253–259.

- 3 Van Hare FG, Ben-Schchar G, Liebman J et al. Infective endocarditis in infants and children during the past 10 years: a decade of change. *Am Heart J* 1984;107:1235–1240.
- 4 Grupo de Trabajo para el Estudio de Infecciones en Drogadictos. Estudio Multicentro de las Complicaciones Infecciosas en ADVP en España. Análisis de 6481 casos (1977–1986). *Enf Infecc Microbiol Clin* 1988;10:483–487.
- 5 Parker MT, Ball MC. Streptococci and aerococci associated with systemic infection in man. *J Med Microbiol* 1976;9:275–302.
- 6 Robert RB, Krieger AG, Schiller NL et al. *Viridans* streptococcal endocarditis: the role of various species, including pyrodoxal-dependent streptococci. *Rev Infect Dis* 1979;1:955–959.
- 7 Sande MM, Korzeniowski OM, Scheld WM. Factors influencing the pathogenesis and prevention of infective endocarditis. *Scand J Infect Dis* 1982;31(suppl):48–54.
- 8 Crawford I, Russell C. Streptococci isolated from bloodstream and gingival crevice of man. *J. Med Microbiol* 1983;10:274–276.
- 9 Pulliam L, Dall L, Inokuchi S et al. Effects of exopolysaccharide production by *viridans* streptococci on penicillin therapy of experimental endocarditis. *J Infect Dis* 1985;151:153–156.
- 10 Dall L, Barnes WG, Lane JW et al. Enzymatic modification of glycocalyx in the treatment of experimental endocarditis due to *viridans* streptococci. *J Infect Dis* 1987;156:736–740.
- 11 Watanakunakorn C. Infective endocarditis as a result of medical progress. *Am J Med* 1978;64:917–919.
- 12 Crawford I, Russell C. Comparative adhesion of seven species of streptococci isolated from the blood of patients with subacute bacterial endocarditis to fibrin-platelet clots in vitro. *J Appl Bacteriol* 1986;60:127–133.
- 13 Gossling J. Occurrence and pathogenicity of the *Streptococcus milleri* group. *Rev Infect Dis* 1988;10:257–285.
- 14 Malinverni R, Francioli P, Gerber A et al. Prophylaxe der bakteriellen Endokarditis. Empfehlungen der Schweizerischen Arbeits-gruppe für Endokarditisprophylaxe. *Schweiz Med Wochenschr* 1984;114:1246–1252.
- 15 Working Party of the British Society for Antimicrobial Chemotherapy. The antibiotic prophylaxis of infective endocarditis. Report of a working party of the British Society for Antimicrobial Chemotherapy. *Lancet* 1990;1:88–89.
- 16 Dajani AS, Bisno AL, Chung KJ et al. Prevention of bacterial endocarditis. Recommendations by the American Heart Association. *JAMA* 1990;264:2919–2922.
- 17 Scheld WM, Zak O, Vosbeck K et al. Bacterial adhesion in the pathogenesis of infective endocarditis. Effect of subinhibitory antibiotic concentrations on streptococcal adhesion in vitro and the development of endocarditis in rabbits. *J Clin Invest* 1981;68:1381–1384.
- 18 Hess J, Dankerts J, Durack D. Significance of penicillin tolerance *in vivo*: prevention of experimental *Streptococcus sanguis* endocarditis. *J Antimicrob Chemother* 1983;11:555–564.
- 19 Southwick FS, Durack DT. Chemotherapy of experimental streptococcal endocarditis. III. Failure of a bacteriostatic agent (tetracycline) in prophylaxis. *J Clin Chem* 1974;27:261–264.
- 20 Glauser MP, Francioli P. Successful prophylaxis against experimental streptococcal endocarditis with bacteriostatic antibiotics. *J Infect Dis* 1982;146:806–810.
- 21 Hardie JM. Genus *Streptococcus*. In: Sneath PHA, Mair NS, Sharpe ME, Holt JG, eds. *Bergey's Manual of Systematic Bacteriology*, vol. 2. Baltimore: Williams & Wilkins, 1986;1043–1071.
- 22 Loesche WJ. Role of *Streptococcus mutans* in human dental decay. *Microbiol Rev* 1986;50:356–380.
- 23 National Committee for Clinical Laboratory Standards. Tentative standard reference agar dilution procedure for antimicrobial susceptibility testing of anaerobic bacteria. *Villanova NCCLS* 1982;2:70–101.
- 24 Scheld WM, Sande ME. Endocarditis e infecciones intravasculares. In Mandel GL, Douglas RG, Bennett JE, eds. *Enfermedades Infecciosas. Principio y Práctica*. Buenos Aires: Panamericana, 1991;705–742.
- 25 Durack DT. Profilaxis de la endocarditis infecciosa. In Mandel GL, Douglas RG, Bennett JE, eds. *Enfermedades Infecciosas. Principio y Práctica*. Buenos Aires: Panamericana, 1991;752–758.
- 26 Committee on Rheumatic Fever and Bacterial Endocarditis of the Council on Cardiovascular Disease in the Young of the American Heart Association. Prevention of bacterial endocarditis. *Circulation* 1977;56(suppl):139A–143A.
- 27 Shanson DC, Cannon P, Wilks M. Amoxycillin compared with penicillin V for the prophylaxis of dental bacteriemia. *J Antimicrob Chemother* 1978;4:431–436.
- 28 Barco CT. Prevention of infective endocarditis: a review of the medical and dental literature. *J Periodontol* 1991;62:510–523.
- 29 Liébana J, Castillo A, Peis J et al. Antimicrobial susceptibility of 1042 strains of *Streptococcus mutans* and *Streptococcus sobrinus*: comparison from 1985 to 1989. *Oral Microb Immunol* 1991;6:146–150.
- 30 Parrillo JE, Borst GC, Mazur MH et al. Endocarditis due to resistant *viridans* streptococci during oral penicillin chemoprophylaxis. *N Engl J Med* 1979;300:296–298.
- 31 Horaud T, Delbos F. *Viridans* streptococci in infective endocarditis: species distribution and susceptibility to antibiotics. *Eur Heart J* 1984;5(suppl C):39–44.
- 32 Holloway Y, Dankert J, Hess J. Penicillin tolerance and bacterial endocarditis. *Lancet* 1981;i:589.
- 33 Pujadas R, Escrivá E, Argimon J et al. Amoxycillin prophylaxis and infective endocarditis. *Lancet* 1986;ii:746.
- 34 Brennan RO, Durack DT. Therapeutic significance of penicillin tolerance in experimental streptococcal endocarditis. *Antimicrob Agents Chemother* 1983;23:276–281.
- 35 Pulliam L, Inokuchi S, Hadley WK et al. Penicillin tolerance of *viridans* streptococci delays sterilization of vegetation in experimental endocarditis. *Clin Res* 1980;28:45A.
- 36 Bouvet A, Acar JF. Isolement et étude morphologique des streptococques déficients, cultivant en satellitisme. I. Mise en

- évidence au cours des endocarditis bactériennes. INSERM 1976;65:327–338.
- 37 Gephart JF, Washington JA. Antimicrobial susceptibility of nutritionally variant *viridans* streptococci. J Infect Dis 1982;146:536–539.
- 38 Stein DS, Nelson KE. Endocarditis due to nutritionally deficient streptococci. Therapeutic dilemma. Rev Infect Dis 1987;9:908–916.
- 39 Little WA, Thompson LA, Bowen WH. Antibiotic susceptibility of *Streptococcus mutans*: comparison of serotype profiles. Antimicrob Agents Chemother 1979;15:440–443.
- 40 Whiley RA, Fraser H, Hardie JM, Beighton D. Phenotypic differentiation of *Streptococcus intermedius*, *Streptococcus constellatus* and *Streptococcus anginosus* strains within the ‘*Streptococcus milleri* group’. J Clin Microbiol 1990;7:1497–1502.