#### Supplementary Data to:

# Investigating vulnerability of the conserved SARS-CoV-2 spike's heptad repeat 2 as target for fusion inhibitors using chimeric miniproteins

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# Table S1-A. Mutations at the HR1 and HR2 regions for the major SARS-CoV-2 variants of concern (VOCs). Mutations are highlighted in red.

## HR1

Variant (Seq. reference)	
	913 95
Wuhan-Hu-1 (NC_045512.2)	TQNVLYENQKLIANQFNSAIGKIQDSLSSTASALGKLQD
Alpha B.1.1.7(MZ344997.1)	TQNVLYENQKLIANQFNSAIGKIQDSLSSTASALGKLQD
Beta B.1.351(MW598419.1)	TQNVLYENQKLIANQFNSAIGKIQDSLSSTASALGKLQD
Gamma P1(MW642250.1)	TQNVLYENQKLIANQFNSAIGKIQDSLSSTASALGKLQD
Delta B.1.617.2(MZ009823.1)	TQNVLYENQKLIANQFNSAIGKIQDSLSSTASALGKLQ <mark>N</mark>
Omicron BA.1(OL672836.1)	TQNVLYENQKLIANQFNSAIGKIQDSLSSTASALGKLQD
Omicron BA.2(OM371884.1)	TQNVLYENQKLIANQFNSAIGKIQDSLSSTASALGKLQD
Omicron BA.4(ON373214.1)	TQNVLYENQKLIANQFNSAIGKIQDSLSSTASALGKLQD
Omicron E.G.5.1(OP790748.1)	TQNVLYENQKLIANQFNSAIGKIQDSLSSTASALGKLQD
	951 988
Wuhan-Hu-1 (NC 045512.2)	VVNONAOALNTLVKOLSSNFGAISSVLNDILSRLDKVE
Alpha B.1.1.7 (MZ344997.1)	VVNONAOALNTLVKOLSSNFGAISSVLNDIL <b>A</b> RLDKVE
Beta B.1.351 (MW598419.1)	VVNONAOALNTLVKOLSSNFGAISSVLNDILSRLDKVE
Gamma P1(MW642250.1)	VVNONAOALNTLVKOLSSNFGAISSVLNDILSRLDKVE
Delta B.1.617.2(MZ009823.1)	VVNONAOALNTLVKOLSSNFGAISSVLNDILSRLDKVE
Omicron BA.1(OL672836.1)	VVNHNAOALNTLVKOLSSKFGAISSVLNDIFSRLDKVE
Omicron BA.2(OM371884.1)	VVNHNAOALNTLVKOLSSKFGAISSVLNDILSRLDKVE
Omicron BA.4 (ON373214.1)	VVNHNAOALNTLVKOLSSKFGAISSVLNDILSRLDKVE
Omicron E.G.5.1(OP790748.1)	VVN <mark>H</mark> NAQALNTLVKQLSS <mark>K</mark> FGAISSVLNDILSRLDKVE
	1102
1104	<b>HR2</b>
Wunan-Hu-I (NC_045512.2)	VDLGDISGINASVVNIQKEIDRLNEVAKNLNESLIDLQE
Alpha B.I.I./(MZ34499/.I)	VULGUISGINASVVNIQKEIDRLNEVAKNLNESLIDLQE
Beta B.I.351 (MW598419.1)	
Gamma PI (MW642250.1)	VDLGDISGINAS <mark>F</mark> VNIQKEIDRLNEVAKNLNESLIDLQE
Delta B.1.617.2(MZ009823.1)	VDLGDISGINASVVNIOKEIDRLNEVAKNLNESLIDLOE

Wuhan-Hu-1 (NC 045512.2)	VDLGDISGINASVVNIOKEIDRLNEVAKNLNESLIDLOE
Alpha B.1.1.7 (MZ344997.1)	VDLGDISGINASVVNIQKEIDRLNEVAKNLNESLIDLQE
Beta B.1.351 (MW598419.1)	VDLGDISGINASVVNIQKEIDRLNEVAKNLNESLIDLQE
Gamma P1(MW642250.1)	VDLGDISGINAS <b>F</b> VNIQKEIDRLNEVAKNLNESLIDLQE
Delta B.1.617.2(MZ009823.1)	VDLGDISGINASVVNIQKEIDRLNEVAKNLNESLIDLQE
Omicron BA.1(OL672836.1)	VDLGDISGINASVVNIQKEIDRLNEVAKNLNESLIDLQE
Omicron BA.2(OM371884.1)	VDLGDISGINASVVNIQKEIDRLNEVAKNLNESLIDLQE
Omicron BA.4(ON373214.1)	VDLGDISGINASVVNIQKEIDRLNEVAKNLNESLIDLQE
Omicron E.G.5.1(OP790748.1)	VDLGDISGINASVVNIQKEIDRLNEVAKNLNESLIDLQE

950

Table S1-B. Sequence alignment of the Spikes's HR1 and HR2 regions for human coronaviruses. Alignment has been carried out with Clustal Omega at the EMBL's European Bioinformatics Institute (https://www.ebi.ac.uk/jdispatcher/msa/clustalo) [1]

NCBI Ref.: GenBank: NCBI Ref.: GenBank: GenBank: GenBank:	NC_045512.2 AAP13441.1 YP_009047204.1 AMK59677.1 AXT92561.1 AAS58177.1 QNT54842.1	SARS-COV-2 ( SARS-COV MERS-COV HCOV-OC43 HCOV-HKU1 HCOV-NL63 HCOV-229E	WUHAM)
HR1			
SARS-CoV-2	TQNVLYENQKLIANQFNSAI	IGKIQDSLSSTA-	SALGKLQD
SARS-CoV	TQNVLYENQKQIANQFNKAI	ISQIQESLTTTS-	TALGKLQD
MERS-COV	TQQVLSENQKLIANKFNQAI	LGAMQTGFTTTN-	EAFQKVQD
HCOV-OC43	TMDVLSQNQKLIANAFNNAI	LDAIQEGFDATN-	SALVKIQA
HCOV-HKU1	TMDVLNKNQKLIANAFNKAI	LLSIQNGFTATN-	SALAKIQS
HCOV-NL63	QTDVLQENQKILAASFNKA	INNIVASFSSVND	AITQTAEAIHTVTIALNKIQD
HCOV-229E	QTDVLQENQRILAASFNKAN :** :**: :* **.*:	ATNIVDAFTGVND	AITQTSQALQTVATALNKIQD *: *:*
SARS-CoV-2	VVNQNAQALNTLVKQLSSN	FGAISSVLNDILS	RLDKVE
SARS-CoV	VVNQNAQALNTLVKQLSSNE	GAISSVLNDILS	RLDKVE
MERS-CoV	AVNNNAQALSKLASELSNTH	FGAISASIGDIIQ	RLDVLE
HCOV-OC43	VVNANAEALNNLLQQLSNR	FGAISSSLQEILS	RLDALE
HCOV-HKU1	VVNVNAQALNSLLQQLFNKI	FGAISSSLQEILS	RLDNLE
HCOV-NL63	VVNQQGSALNHLTSQLRHNH	FQAISNSIQAIYD	RLDSIQ
HCOV-229E	VVNQQGNSLNHLTSQLRQN	FQAISSSIQAIYD	RLDIIQ
	.** ::*. * .:* *	* * * * • * •	*** ::
HR2			
SARS-CoV-2	-VDLGDISGINASVVNIQKE	EID	RLNEVAKNLNESLIDLQE
SARS-CoV	-VDLGDISGINASVVNIQKE	EID	RLNEVAKNLNESLIDLQE
MERS-CoV	-PNFGSLTQINTTLLDLTYE	EML	SLQQVVKALNESYIDLKE
HCOV-OC43	-PDLS-LDYINVTLLDLQVE	EMN	RLQEAIKVLNQSYINLKD
HCOV-HKU1	-PNLTFNSHINATFLDLYYE	EMN	VIQESIKSLNSSFINLKE
HCOV-NL63	KPNFD-LTPFNLTYLNLSSE	ELKQLEAKTASLF	QTTVELQGLIDQINSTYVDLKL
HCOV-229E	VPDLV-VEQYNQTILNLTSE	EISTLENKSAELN	YTVQKLQTLIDNINSTLVDLKW
	* * * * * * *	*:	•• • • • • • • • • •

Protein	-		Sequence		
variant			_		
L3B	DVLYENQKLI	AN <b>K</b> FNSAIGK	IQDSLSSTAS	<b>E</b> LGKLQD <b>E</b> VN	QNAQ <b>D</b> LNTLV
	KQLSSNFG <b>R</b> I	SS <b>E</b> LNDILSR	ldk <mark>gepa</mark> kdl	RS <mark>D</mark> IDNL <b>E</b> SK	IAGFNSSLQK
	VLTNLAQ <b>K</b> NQ	NVEDKLKGLE	S <b>r</b> tssl <b>ek</b> qi	KGIASNFQN <b>E</b>	ILKQ <b>r</b> eylvn
	<mark>KGSG</mark> NVLYEN	QKLIENQFNS	AIGKIQDSLS	ST <b>K</b> SALGKL <mark>K</mark>	DVVNQN <mark>K</mark> QAL
	NTLVKQLSSN	FGAISSVLND	I <b>K</b> SRLDKVE		
N2A	<b>D</b> VLYENQKLI	AN <mark>K</mark> FNSAIGK	IQDSLSSTAS	<b>E</b> LGKLQD <b>E</b> VN	<mark>k</mark> gesk <mark>k</mark> nv <b>e</b> d
	<b>k</b> lkgl <b>e</b> s <b>r</b> ts	SL <mark>ek</mark> qikgia	SNFQN <b>E</b> ILKQ	<b>r</b> eylvn <mark>kgsg</mark>	NVLYENQKLI
	<b>E</b> NQFNSAIGK	IQDSLSST <mark>k</mark> S	ALGKL <mark>K</mark> DVVN	QN	
N2B	DVLYENQKLI	AN <b>K</b> FNSAIGK	IQDSLSSTAS	<b>E</b> LGKLQD <b>E</b> VN	QNAQ <b>D</b> LNT <mark>GK</mark>
	<mark>SG</mark> ENLAQ <mark>K</mark> NQ	NVEDKLKGLE	S <b>r</b> tssl <b>ek</b> qi	KGIASNFQN <b>E</b>	ILKQ <mark>r</mark> eylvn
	<mark>KGSG</mark> NVLYEN	QKLIENQFNS	AIGKIQDSLS	ST <b>K</b> SALGKL <mark>K</mark>	DVVNQN <mark>K</mark> QAL
	NT				
C2A	S <b>e</b> lgklqd <b>e</b> v	NQNAQ <b>D</b> LNTL	VKQLSSNFG <mark>R</mark>	ISSELNDILS	rldk <mark>gepa</mark> kd
	LRS <b>D</b> IDNL <b>E</b> S	<b>K</b> IAGFNSSLQ	KVLTNLAQ <mark>K</mark> N	QNV <b>E</b> DKLK <mark>K</mark> A	<mark>gsd</mark> v <mark>k</mark> kl <b>k</b> dv
	VNQN <mark>K</mark> QALNT	LVKQLSSNFG	AISSVLNDI <mark>k</mark>	SRLDKVE <mark>W</mark>	
L3C	DVLYENQKLI	AN <mark>K</mark> FNSAI <mark>K</mark> K	IQDSLSSTAS	EL <mark>K</mark> KLQD <b>E</b> VN	QNAQ <b>D</b> LNTLV
	KQLSSNF <mark>K</mark> RI	SSELNDILSR	LDK <mark>GEPA</mark> KDL	RS <b>D</b> IDNL <b>E</b> SK	IA <mark>K</mark> FNSSLQK
	VLTNLAQ <b>K</b> NQ	NVEDKLK <mark>T</mark> LE	S <b>r</b> tssl <b>ek</b> QI	K <mark>K</mark> IASNFQN <b>E</b>	ILKQ <b>r</b> eylvn
	<mark>KGSG</mark> NVLYEN	QKLIENQFNS	AI <mark>K</mark> KIQDSLS	ST <b>K</b> SAL <mark>K</mark> KL <b>K</b>	DVVNQN <mark>K</mark> QAL
	NTLVKQLSSN	F <mark>S</mark> AISSVLND	I <b>K</b> SRLDKVE		
N2C	DVLYENQKLI	AN <mark>K</mark> FNSAI <mark>K</mark> K	IQDSLSSTAS	EL <mark>K</mark> KLQD <b>E</b> VN	<mark>k</mark> gesk <mark>k</mark> nv <b>e</b> d
	KLK <mark>T</mark> LESRTS	SL <b>ek</b> qik <mark>k</mark> ia	SNFQNEILKQ	<b>r</b> eylvn <mark>kgsg</mark>	NVLYENQKLI
	<b>E</b> NQFNSAI <mark>K</mark> K	IQDSLSSTKS	AL <mark>K</mark> KL <b>K</b> DVVN	QN	
N2D	DVLYENQKLI	AN <mark>K</mark> FNSAI <mark>K</mark> K	IQDSLSSTAS	EL <mark>K</mark> KLQD <b>E</b> VN	QNAQ <b>D</b> LNT <mark>GK</mark>
	<mark>SG</mark> ENLAQ <b>K</b> NQ	NVEDKLK <mark>T</mark> LE	S <b>r</b> tssl <b>ek</b> qi	K <mark>K</mark> IASNFQN <b>E</b>	ILKQ <mark>r</mark> eylvn
	<mark>KGSG</mark> NVLYEN	QKLIENQFNS	AI <mark>K</mark> KIQDSLS	ST <b>k</b> SAL <mark>k</mark> KL <b>k</b>	DVVNQN <mark>K</mark> QAL
	NT				
C2C	S <b>E</b> L <mark>K</mark> KLQD <b>E</b> V	NQNAQDLNTL	VKQLSSNF <mark>K</mark> R	ISSELNDILS	rldk <mark>gepa</mark> kd
	LRS <b>D</b> IDNL <b>E</b> S	<b>k</b> ia <mark>k</mark> fnsslQ	KVLTNLAQ <b>K</b> N	QNV <b>E</b> DKLK <mark>K</mark> A	<mark>gsd</mark> v <mark>k</mark> kl <b>k</b> dv
	VNQN <mark>K</mark> QALNT	LVKQLSSNF <mark>S</mark>	AISSVLNDI <mark>k</mark>	SRLDKVE <mark>w</mark>	

Table S2. Amino acid sequences of the shortened CoVS-HR1 miniproteins.

All proteins contain an N-terminal methionine and a C-terminal tail of sequence GGGGSHHHHHH. A C-terminal tryptophan added to confer UV absorption is highlighted in green for C2A and C2C proteins. Loop residues connecting the helices are highlighted in yellow. Mutated amino acids to substitute glycine residues are highlighted in magenta. Other amino acid substitutions made to engineer stabilizing salt bridges in the trimeric coiled-coil structure are colored in red (negative charge) or blue (positive charge).



**Figure S1**: Sequence and topological organization of the CoVS-HR1 miniproteins. The newly built loops connecting each consecutive helix have been highlighted in yellow. Engineered residues to create stabilizing salt bridges at (e) and (g) positions of the heptad repeats are colored in blue and red. Substituted glycine residues in the L3C, N2C, N2D and C2C variants are colored in magenta.



**Figure S2:** Far-UV CD spectra of the CoVS-HR1 miniproteins at 25°C and different pH. Spectra were recorded at a protein concentration of 15  $\mu$ M in different buffers (20 mM glycine/HCl pH 2.5; 50 mM sodium acetate pH 4; 50 mM sodium cacodilate pH 6; 50 mM sodium phosphate pH 7.4; 50 mM sodium carbonate pH 9.4). The data are normalized as mean-residue molar ellipticity.

			Experimental R <sub>h</sub> (nm)				Calculated Rh (monomer) <sup>(a)</sup>
Protein↓	$pH \rightarrow$	2.5	4.0	6.0	7.4	9.4	
N2A		3.3	2.3	2.2	2.5	2.2	2.3
N2B		3.0	2.6	2.5	2.6	2.6	2.4
N2C		2.5	2.3	2.3	2.3	2.2	2.3
N2D		2.8	2.6	2.5	2.4	2.6	2.4
C2A		3.1	3.0	3.4	3.3	3.3	2.3
C2C		2.7	2.5	2.7	3.3	3.1	2.3
$L3B^{(b)}$		3.0	3.6	2.8	3.1	3.4	3.3
L3C <sup>(b)</sup>		2.9	3.0	3.2	3.0	3.0	3.3

Table S3. Hydrodynamic radii (R<sub>h</sub>) of the CoVS-HR1 proteins measured by dynamic light scattering (DLS) at 25°C

<sup>(a)</sup> Calculated with HYDROPRO software [2] using the models of the miniproteins. <sup>(b)</sup> Data taken from [3].

Protein	pН	T <sub>m</sub> (°C) <sup>(a)</sup>	$\Delta H_{U,m}$ (kJ mol <sup>-1</sup> ) <sup>(a)</sup>	$\Delta C_{pU}$ (kJ mol <sup>-1</sup> ) <sup>(b)</sup>
	2.5	4.5	0.06	
	4	44.6	204	
N2A	6	53.1	247	5.1
	7.4	53.4	252	
	9.4	48.5	261	
	2.5	4.9	-1.4	
	4	42.3	234	
N2B	6	53.2	298	6.2
	7.4	51.9	300	
	9.4	47.7	295	
	2.5	35.5	166	
	4	65.4	301	
N2C	6	76.2	340	4.3
	7.4	81.1	359	
	9.4	72.5	400	
	2.5	34.2	185	
	4	63.1	314	
N2D	6	74.2	351	4.2
	7.4	77.7	368	
	9.4	70.7	391	
	2.5	34.2	185	
	4	63.1	314	
C2A	6	74.2	351	n.d.
	7.4	77.7	368	
	9.4	70.7	391	
	2.5	34.2	185	
	4	63.1	314	
C2C	6	74.2	351	n.d.
	7.4	77.7	368	
	9.4	70.7	391	

Table S4. Thermodynamic parameters of unfolding of the CoVS-HR1 miniproteins measured by DSC.

(a) Determined from non-linear least-squares fittings using the two-state unfolding model for the N miniproteins and from direct peak integration for the C miniproteins.

(b) Determined from linear fittings of  $\Delta H_{U,m}$  vs  $T_m$  values for each miniprotein. Data at pH 9.4 were excluded from the linear fittings. n.d.: Not determined



**Figure S3.** Plots of unfolding enthalpy  $(\Delta H_{U,m})$  versus melting temperature  $(T_m)$  for the CoVS-HR1 miniproteins. The unfolding of the N-miniproteins follows the two-state model. The lines represent linear fits of the data between pH 2.5 and pH 7.4. The values for the C-miniproteins have been determined by direct integration of the unfolding peaks.



**Figure S4.** Effect of protein concentration upon the DSC thermograms of the C miniproteins. The experiments were carried out ay pH 7.4, in 50 mM sodium phosphate buffer, at a scan rate of 90 °C/h at the indicated protein concentrations. The curves of C2A have been displaced vertically by 40 kJ K<sup>-1</sup> mol<sup>-1</sup> for the sake of clarity.



**Figure S5:** Model structures of the hypothetical complexes between the CoVS-HR1 miniproteins with HR2-derived peptides. The crystallographic structure of the complex between L3B and V39E (HR2 residues 1164-1202) is also shown for reference (PDB id: 7ZR2 [3]). The model of the complexes between the N2A, N2B and C2A miniproteins and the complementary HR2 peptides (V27E: HR2 residues 1176-1202; V19E: HR2 residues 1164-1182) have been created by structural alignment of each miniprotein with the L3B-V39E complex. The sequences of the HR2 peptides are shown at the bottom. The three peptides contain a C-terminal SGGY tag and are N-acetylated and C-amidated.

Protein	HR2 peptide	K <sub>b</sub> (·10 <sup>3</sup> M <sup>-1</sup> )	K <sub>d</sub> (µM)	ΔG <sub>b</sub> (kJ mol <sup>-1</sup> )	ΔH <sub>b</sub> (kJ mol <sup>-1</sup> )	T·ΔS₅ (kJ mol <sup>-1</sup> )	n
N2A		$270\pm100$	$3.7 \pm 1.3$	$-31.0\pm0.9$	$-9.2\pm0.9$	$+21.8\pm1.8$	$1.10\pm0.05$
$N2B^{(b)}$	VO7E	$107 \pm 23$	9.3 ± 2.0	$-29.9\pm0.6$	$-68 \pm 5$	$-33 \pm 5$	$0.91\pm0.04$
N2C	V2/E	$340\pm130$	$3.0 \pm 1.1$	$-31.6\pm0.9$	$+6.1\pm0.5$	$+39.3\pm1.6$	$1.21\pm0.05$
N2D		$360\pm80$	$2.8\pm0.6$	$-31.7\pm0.6$	$+4.6\pm0.2$	$+35.7\pm0.8$	$1.09\pm0.03$
C2C	V39E	$121 \pm 11$	$8.3\pm0.8$	$-29.0\pm0.2$	$-6.2\pm0.2$	$+22.8\pm0.4$	$1.02\pm0.02$

Table S5: Thermodynamic parameters of binding of HR2 peptides to the CovS-HR1 proteins measured by ITC at 25 °C.

<sup>(b)</sup> Parameters measured at 37 °C

Table S6: Thermodynamic parameters of binding of HR2 peptides to the CoVS-HR1 proteins measured by DSC using a model of two-state unfolding coupled to ligand binding (NL  $\Leftrightarrow$  N + L $\Leftrightarrow$  U + L).

Protein	HR2 peptide	K <sub>b</sub> <sup>(a)</sup> (·10 <sup>3</sup> M <sup>-1</sup> )	ΔH <sub>b</sub> <sup>(a)</sup> (kJ mol <sup>-1</sup> )	ΔC <sub>p,b</sub> <sup>(a)</sup> (kJ K <sup>-1</sup> mol <sup>-1</sup> )	K <sub>d</sub> (µM)
N2A	V27E	34.1 ± 0.6	$-47.7\pm0.5$	$-3.38\pm0.04$	29.3
N2B		$27.6\pm0.7$	$-23.4 \pm 1.1$	$-3.45\pm0.05$	36.2
N2C		$100 \pm 10$	$-54 \pm 5$	$-2.65\pm0.25$	10
N2D		$110.5\pm4.9$	$-44.4 \pm 1.3$	$-3.66\pm0.04$	9.1
N2A	V39E	$13.6\pm0.4$	$-35.9\pm0.9$	$-3.64\pm0.04$	73.3
N2B		$19.6\pm0.4$	$-41.1\pm0.8$	$-3.14\pm0.04$	51.0
N2C		$27.7\pm1.1$	$-35.6\pm0.8$	$-4.06\pm0.06$	36.0
N2D		$52.2 \pm 1.8$	$-36.2\pm0.8$	$-4.90\pm0.05$	19.2

<sup>(a)</sup> Parameters at 37 °C determined from the global fitting of the DSC thermograms in presence of peptides at different peptide:protein ratio. Uncertainties correspond to 95% confidence intervals of the fittings.



**Figure S6:** DSC thermograms of the N miniproteins in absence and in presence of three different concentrations of the V39E peptide. Experiments were carried out at pH 7.4 in 50 mM sodium phosphate buffer with a protein concentration of 30  $\mu$ M. The black curves represent the global fits using a model of binding coupled to unfolding (NL  $\Leftrightarrow$  N + L $\Leftrightarrow$  U + L).



**Figure S7:** DSC thermograms of C2A and C2C in absence and in presence of two different concentrations of the V19E peptide. Experiments were carried out at pH 7.4 in 50 mM sodium phosphate buffer with a protein concentration of  $30 \mu$ M.



**Figure S8**: Far-UV CD spectra of the CoVS-HR1 miniproteins in presence of a four-fold molar excess of V39E peptide. The spectra have been recorded at 25 °C in 50 mM sodium phosphate buffer at pH 7.4. The spectra have been normalized per mole of molecules for a proper comparison. The dashed lines represent the spectra that would be obtained for mixtures of non-interacting molecules.

## References

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