

1 **Beyond the Reproductive Tract: Gut Microbiome and Its Influence on**  
2 **Gynecological Health**

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20 **KEY POINTS**

- 21 • Growing evidence defines the gut microbiome as an endocrine organ and suggests  
22 its potential role in gynecological physiology and pathophysiology due to its  
23 bidirectional relationship with female hormone levels.
- 24 • The estrobolome is defined as the collection of the gut microbial genes that encode  
25 enzymes implicated in estrogen activation, therefore potentially affecting  
26 different hormone-dependent gynecological functions.
- 27 • Gut microbial dysbiosis may lead to the activation of the immune responses and  
28 inflammation, and hormone dysregulation, thereby contributing to the onset and  
29 progression of multiple estrogen-driven inflammatory pathologies.
- 30 • The gut microbiome has been associated with endometriosis, polycystic ovary  
31 syndrome (PCOS), gynecological cancer, abortion and infertility.
- 32 • Current knowledge about the association of the gut microbiome with  
33 gynecological health is under-examined and requires well-designed studies based  
34 on standardized protocols for consistent findings.

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43 **ABSTRACT**

44 **Purpose of review:** The analysis of microbiome in association with female health is today  
45 a “hot topic” with the main focus on microbes in the female reproductive tract.  
46 Nevertheless, recent studies are providing novel information of the possible influence of  
47 the gut microbiome on gynecological health outcomes, especially as we start to  
48 understand that the gut microbiome is an extended endocrine organ influencing female  
49 hormonal levels. This review summarizes the current knowledge of the gut microbes in  
50 association with gynecological health.

51 **Recent findings:** The gut microbiome has been associated with endometriosis,  
52 polycystic ovary syndrome, gynecological cancers, and infertility, although there is a lack  
53 of consistency and consensus among studies due to different study designs and protocols  
54 used, and the studies in general are underpowered.

55 **Summary:** The interconnection between the gut microbiome and reproductive health is  
56 complex and further research is warranted. The current knowledge in the field emphasizes  
57 the link between the microbiome and gynecological health outcomes, with high potential  
58 for novel diagnostic and treatment tools via modulation of the microenvironment.

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60 **Keywords:** microbiota, endometriosis, polycystic ovary syndrome, cancer, infertility

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## 65 1. INTRODUCTION

66 The human gastrointestinal tract harbors trillions of microorganisms including bacteria,  
67 archaea, viruses, protozoa, and fungi, being the largest and most diverse microbial  
68 ecosystem within the human body. The collection of the intestinal microbial genomes,  
69 the gut microbiome, is a research field of increasing interest since it represents a genetic  
70 pool more than one order of magnitude higher in genes than the human genome [1]. The  
71 vast majority of these microbial communities co-evolved symbiotically with the host and  
72 contribute to important metabolic, immune and epithelial functions, being crucial for the  
73 host physiology and pathophysiology [2–4].

74 Recent evidence refers to the gut microbiome as an extended endocrine organ due to its  
75 profound interaction with hormone levels [5]. Furthermore, a sex bias has been identified  
76 in microbiome-related diseases which are associated with sex hormones [6]. In this  
77 context, the term “microgenderome” has emerged to define the interactions between the  
78 microbiome, sex hormones and the immune system [7]. This interaction could unravel  
79 the molecular mechanisms underlying the gut microbiome influence on female  
80 reproductive health and how its dysbiosis could lead to different pathologies [8].

81 The gut microbiome-estrogen axis has been proposed as a cornerstone implicated in the  
82 pathogenesis of different gynecological conditions, such as polycystic ovary syndrome  
83 (PCOS), endometriosis, gynecological cancer, infertility and adverse pregnancy  
84 conditions [9] (Figure 1). This crosstalk between the gut microbiome and estrogens is  
85 regulated by the estrobolome, the aggregate of gut bacterial genes whose products are  
86 capable of metabolizing estrogens [10]. Certain enteric bacteria secrete  $\beta$ -glucuronidase,  
87 the main estrogen-regulator of the estrobolome, that converts the conjugated estrogen  
88 (glucuronic acid) into its deconjugated form that exerts its biological activity [11]. Thus,

89 an optimal  $\beta$ -glucuronidase activity reduces the inactivation of estrogen, leading to a  
90 balanced hormone circulating levels. However, a reduction in the gut microbial diversity  
91 as a result of dysbiosis and inflammation could reduce the  $\beta$ -glucuronidase activity and  
92 this reduction has been linked to hypoestrogenic pathologies such as obesity, metabolic  
93 syndrome, cardiovascular disease and cognitive decline [9]. Otherwise, an increased  $\beta$ -  
94 glucuronidase activity is associated with hyperestrogenic conditions and can lead to the  
95 progression of gynecological estrogen-driven diseases [9] (Figure 1). Since the  
96 estrobolome may have a profound influence on these pathologies, future interventions  
97 targeting the estrobolome-microbiome axis emerge as promising diagnostic and treatment  
98 tools for women's health [5]. However, the molecular mechanisms underlying the  
99 relationships between the gut microbiome, estrogen metabolism and gynecological  
100 outcomes are still in its infancy. In this narrative review, we summarize the whole body  
101 of knowledge of the gut microbiome and its interactions with different gynecological  
102 health conditions.

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## 104 **2. ENDOMETRIOSIS**

105 Endometriosis is defined as an estrogen-dependent chronic inflammatory gynecological  
106 disease characterized by endometrial-like tissue present outside of the uterus. It represents  
107 a major health concern since it affects 6-10% of women in reproductive age [12].  
108 Regardless of the continuous research, the exact mechanisms of endometriosis are still  
109 undetermined. The most widely accepted hypothesis for the origin of endometriosis is  
110 Sampson's retrograde menstruation, which explains that women commonly have  
111 retrograde menstrual flow [13]. Nevertheless, only 10% of women are diagnosed with  
112 endometriosis [14]. Growing evidence proposes a multifactorial origin for endometriosis

113 development driven by genetic predisposition, environmental factors, inflammation,  
114 immune activation, hormone dysregulation and microbial dysbiosis [15, 16].

115 Much is known of the role of the gut microbiome in maintaining the integrity of the  
116 gastrointestinal epithelial lining and immune balance to prevent bacterial translocation,  
117 which can cause low-grade systemic inflammation [17]. While it is well-established that  
118 the gut microbes influence immunomodulation and the development of various  
119 inflammatory diseases [18], current studies have highlighted the potential implication of  
120 enteric microbes in the pathogenesis of endometriosis [11, 19]. The “bacterial  
121 contamination hypothesis” proposes that besides estrogen regulation, the gut microbiome  
122 could contribute to the onset of endometriosis through lipopolysaccharide (LPS)  
123 endotoxin as the initial trigger and bacterial contamination as its source in the intrauterine  
124 environment [20]. LPS is found in the cell wall of Gram-negative bacteria, and is a marker  
125 of inflammation which has been linked to endometriosis lesions activating the immune  
126 response by binding with Toll-like receptor 4 [20]. A systematic review concluded that  
127 increased abundance of *Proteobacteria*, *Enterobacteriaceae*, *Streptococcus* and  
128 *Escherichia* in the gut associated with the presence of endometriosis [21] (Figure 1). In  
129 line, a *Shigella/Escherichia* dominant gut microbiome in women with advanced stages  
130 3/4 endometriosis have been described [22], and an increase in *Streptococcus* in the gut  
131 of patients with the 3/4 endometriosis stages has been reported [23]. Further, a recent  
132 study detected a higher abundance of *Shigella flexneri* (*Proteobacteria* phylum) in  
133 patients with external genital endometriosis compared to controls [24]. Additionally, a  
134 higher proportion of Gram-negative bacteria belonging to *Desulfobacterota* phylum have  
135 been detected in women with endometriosis when compared to healthy controls [25].  
136 *Proteobacteria* and *Desulfobacterota* phyla are both characterized by Gram-negative  
137 staining, and, therefore, presenting LPS in the outer membrane [26]. Interestingly, a

138 recent translational study demonstrated a pathogenic mechanism via *Fusobacterium* (a  
139 Gram-negative bacterial genus) infection in endometrial cells through activation of  
140 transforming growth factor- $\beta$  (TGF- $\beta$ ) signaling [27]. This activation leads to the  
141 transition from quiescent fibroblasts to transgelin (TAGLN)-positive myofibroblasts,  
142 which are able to proliferate, adhere, and migrate *in vitro* [27]. It was also observed that  
143 inoculation of *Fusobacterium nucleatum* in a murine model of endometriosis resulted in  
144 increased numbers and weights of endometriotic lesions [27].

145 Short-chain fatty acids (SCFAs) are microbial metabolites with pleiotropic beneficial  
146 effects for the host metabolism and immune regulation through their action on T-  
147 regulatory cells [28, 29]. *Lachnospiraceae*, *Eubacteriaceae* and *Ruminocacceae* family  
148 members are the main producers of SCFAs in the intestine, particularly producing acetate  
149 and butyrate [30, 31] (Figure 1). Several studies have found lower abundances of  
150 butyrate-producing microbes such as *Lachnospira*, *Ruminococcus*, *Eubacterium eligens*  
151 and *Coprococcus catus* in women with endometriosis [23, 24, 32, 33]. In particular,  
152 butyrate has been described as an anti-inflammatory mediator that could indirectly  
153 regulate endometriosis-related symptoms such as visceral inflammatory pain [34].  
154 Considering the connection between the dysbiosis (through an increase of Gram-negative  
155 bacteria and/or depletion of SCFAs producers) and immune dysfunction, future studies  
156 are needed to study whether imbalances within these gut microbes are the cause,  
157 consequence or enhancer of endometriosis.

158 Given the hyperestrogenic conditions associated with endometriosis, there is growing  
159 interest focused on the estrobolome as a key factor contributing to the progression of the  
160 disease. Alterations in the gut microbiome that result in overexpression of estrobolome  
161 associated genes could both trigger formation and maintenance of the lesions.  
162 Interestingly,  $\beta$ -glucuronidase activity has been found in Gram-negative bacteria [25].

163 Moreover, an analysis of microbial genomes associated enteric *Bacteroides*,  
164 *Bifidobacterium*, *Escherichia* and *Lactobacillus* with  $\beta$ -glucuronidase production [35]  
165 (Figure 1). In endometriosis population, several studies have reported higher abundances  
166 in these bacteria [23, 32]. Nevertheless, an enzymatic activity study of fecal samples did  
167 not reveal significant differences in  $\beta$ -glucuronidase activity in women with and without  
168 endometriosis [36]. We have recently performed the first whole metagenome study on a  
169 cohort of 1,000 women with and without endometriosis, and did not find any microbial  
170 features (species or pathways) associated with the disease [37, 38]. Furthermore, we  
171 analyzed the estrobolome-associated genes and did not find any significant differences  
172 between the two study groups [38].

173 Altogether, the previous results pave the way for future strategies targeted to the gut  
174 microbiome-estrobolome axis, nevertheless the studies in the field lack consensus and the  
175 identification of an endometriosis-associated microbiome profile still constitutes a debate  
176 without a definitive answer. It is clear that endometriosis is a complex heterogenic disease  
177 and further studies applying well defined, adequately powered study groups are warranted  
178 to determine the core microbial composition in endometriosis.

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### 180 **3. POLYCYSTIC OVARY SYNDROME**

181 The polycystic ovary syndrome (PCOS) is one of the most prevalent endocrine disorders  
182 in women of reproductive age, affecting up to 20% of women [39]. Despite its high  
183 prevalence, its multifactorial complexity has made it challenging to understand the  
184 underlying etiology. Possible triggers include genetic factors, intrauterine environment,  
185 lifestyle, and, in an increasingly explored approach, alterations in the gut microbiome [40,  
186 41].



187 Considering the role of the gut microbes in metabolic disorders, the search of the link  
188 between the gut microbes and PCOS is plausible. Numerous studies have detected  
189 alterations in the microbial richness, diversity, and microbial composition in PCOS [41].  
190 Specifically, PCOS patients have been observed to exhibit a decrease in  $\alpha$ -diversity  
191 indices (diversity within a sample) compared to controls [42–45]. Regarding  $\beta$ -diversity  
192 (dissimilarity of the microbial community between samples), previous studies have  
193 identified differences in the microbial composition among samples from PCOS patients  
194 compared to healthy controls [42, 44–47]. While other studies have not observed any  
195 significant differences in microbial composition in PCOS [48–52]. Our recent systematic  
196 review summarizes that the women PCOS have decreased microbial diversity in the gut  
197 and that the prevalent taxa are *Bacteroides spp.*, *Parabacteroides spp.*, *Prevotella*,  
198 *Megamonas spp.*, *Megasphaera massiliensis*, *Escherichia/Shigella*, while  
199 *Bifidobacterium spp.*, *Lactobacillus spp.*, *Faecalibacterium*, and *Blautia* are reduced  
200 [41].

201 The presence of obesity and insulin resistance also emerges as crucial factors in the study  
202 of the gut microbes in PCOS. Obesity, a common characteristics in PCOS patients, has  
203 been analyzed in relation to the intestinal microbiome, revealing significant differences  
204 in the gut microbiome  $\beta$ -diversity in PCOS patients with obesity compared to non-obese  
205 patients [53, 54]. In PCOS groups with high BMI, an increased abundance of  
206 *Erysipelotrichaceae*\_UCG-003 [45], *Prevotellaceae* [54], *Streptococcus*, *Fusobacterium*,  
207 *Rhizobacter*, and *Achromobacter* [49] was observed. Nevertheless, other previous studies  
208 have not observed any microbiome differences between the groups [48, 49, 51].  
209 Regarding insulin resistance, studies have shown a decrease in the  $\alpha$ -diversity when  
210 compared to controls [48, 54, 55], and the most abundant bacteria such as *Prevotella*,

211 *Megamonas*, *Dialister* [54], *Prevotella stercorea* [48] and *Faecalibacterium* [55] have  
212 been identified in PCOS patients with insulin resistance.

213 Beyond the bacteria, a number of studies have explored the mycobiome and virome in  
214 PCOS patients, where the increased abundance of fungi: *Saccharomyces*, *Lentinula*, and  
215 *Aspergillus* [46], *Candida*, *Malassezia*, *Kazachstania*, *Microascus*, *Coniochaeta*,  
216 *Xepicula*, *Paraphoma*, *Pyrenochaetopsis*, *Cephaliphora*, *Epicoccum*, and *Sclerophora*  
217 has been observed in PCOS patients [45]. The analysis of the gut virome detected lower  
218 viral diversity and significant alterations in virome composition in women with PCOS,  
219 where the most enriched taxon was *Quimbyviridae* when compared to healthy controls  
220 [56].

221 Altogether, there seems to be a common trend of reduced microbial diversity in PCOS,  
222 nevertheless, the studies are performed on limited sample size and lack consensus.

223

#### 224 **4. GYNECOLOGICAL CANCER**

225 Within the recent years, there has been a growing interest in understanding the connection  
226 between the human microbiome and various types of cancers, including gynecological  
227 cancers. Among these, the most common are endometrial, cervical, and ovarian cancers,  
228 where the endometrial cancer is the most prevalent [57]. However, the most lethal is  
229 ovarian cancer, accounting for 5% of total cancer-related deaths [8, 57]. These cancers  
230 are characterized by being estrogen-mediated tumors [58]. Estrogens have the capacity to  
231 modulate the inflammatory response and increase the production of pro-inflammatory  
232 mediators (IL-6 and TNF- $\alpha$ ) [59]. This can establish a feedback loop that influences the  
233 expression of enzymes associated with ovarian steroidogenesis. The gut microbiome is  
234 able to metabolize these estrogens, increasing their concentration and thereby enhancing

235 the development of endometrial cancer [59]. Simultaneously, this increase in the estrogen  
236 levels also has the potential to induce changes in the gut microbiome, indirectly  
237 contributing to the cancer progression [8].

238 Although there is still limited literature of the relationship between the gut microbiome  
239 and endometrial cancer, existing studies provide contradicting results. One study found a  
240 significant reduction in the gut microbial  $\alpha$ -diversity and differences in  $\beta$ -diversity among  
241 endometrial cancer patients [60], while other did not detect any significant differences  
242 [61]. These discrepancies are also reflected in the phylogenetic composition, with  
243 variations in the abundances of *Firmicutes*, *Proteobacteria*, *Actinobacteria*, and  
244 *Bacteroidetes* in the gut microbiome among endometrial cancer patients when compared  
245 to controls [60, 61].

246 In ovarian cancer studies, changes in  $\beta$ -diversity were consistently observed between  
247 patients and controls, however, in richness, no significant differences were detected  
248 [62,63]. Moreover, increase in specific bacterial abundances such as *Firmicutes*,  
249 *Proteobacteria*, and *Bacteroidetes* phyla have been reported in ovarian cancer patients  
250 [62].

251 The research of the gut microbiome in gynecological cancers is very preliminary and  
252 future research is needed to clarify the potential cancer-associated microbial profile and  
253 unravel the complexity of the relationship between gut microbes and gynecological  
254 cancers.

255

## 256 **5. INFERTILITY**

257 The vaginal microbiome has been the subject of extensive research in relation to female  
258 fertility; however, the influence of the gut microbiome is still relatively unstudied. Recent

259 investigations suggest that the gut microbiome may play a crucial role in the modulation  
260 of the reproductive system through the gut-uterus axis. It has been observed that even a  
261 small alteration in the commensal and symbiotic gut microbes can trigger dysbiosis,  
262 disrupting intestinal homeostasis and increase the risk of inflammatory processes  
263 associated with adverse reproductive pathologies [64] (Figure 1).

264 A previous study highlighted that the diversity and composition of the intestinal  
265 microbiome, along with its metabolite profiles, show significant alterations in patients  
266 who had experienced spontaneous abortions [64]. When analyzing the fecal microbiome  
267 in association with spontaneous abortions, an overrepresentation of various opportunistic  
268 pathogens (*Prevotellaceae\_NK3B31\_group*, *Bacteroidales\_S24\_7\_group*, and  
269 *Eubacterium ruminantium*) was identified in the affected group, while other  
270 microorganisms (*Prevotellaceae*, *Prevotella\_1*, and *Gammaproteobacteria*) were more  
271 abundant in the control group [64]. Additionally, a significant correlation was found  
272 between the metabolites associated with these microorganisms and an increase in  
273 cytokines linked to Th1 and Th17 [64]. The reduction in the richness and diversity of the  
274 microbiome in patients who had suffered abortions supported these findings,  
275 corroborating previous results linking microbiome composition to infertility [65-67].

276 In a similar context, notable differences in the composition of the gut microbiome  
277 between patients with infertility (recurrent implantation failure –RIF- and unexplained  
278 infertility) and controls have been detected [65]. *Bacteroides* and *Hungatella* stood out as  
279 the most abundant genera in the gut in infertile women, especially in cases of  
280 unexplained infertility. A decrease in the genera *Prevotella* 9, *Ruminococcaceae* UCG-  
281 004, *Ruminococcaceae* UCG-010, and an increase in *Bacteroides*, *Dorea* oral clone  
282 FR58, and *Peptoniphilus* were detected in the gut microbiome when compared to controls  
283 [65]. Further studies have detected higher abundance of *Verrucomicrobia*, and members

284 of *Barnesiellaceae* and *Phascolarctobacterium* in the gut [67], while the genera  
285 *Stenotrophomonas*, *Streptococcus*, and *Roseburia* showed a decrease in patients with  
286 infertility [67]. Altogether, regardless of these preliminary studies, there seems to be a  
287 consensus in the increase of the gut *Firmicutes/Bacteroidetes* ratio in infertile women  
288 when compared to controls [64, 66].

289

## 290 6. CONCLUSIONS

291 There is growing body of evidence demonstrating that the gut microbes play important  
292 role in female physiology and pathophysiology and that via its endocrine and hormonal  
293 regulation, specifically estrobolome regulation can influence female reproductive health.  
294 In this review we gather the knowledge of the gut microbiome involvement in  
295 endometriosis, PCOS, cancer and infertility, and with the time, the list of different  
296 gynecological disorders in association with the gut microbiome will definitely grow.

297 The current knowledge of the microbe-disease associations encompasses the microbial  
298 diversity analyses and identification of specific bacterial genera. The majority of the  
299 studies of the gut microbiome in female gynecological health have applied the 16S rRNA  
300 gene sequencing technique that does not have sufficient specificity to identify bacteria on  
301 species level, which makes the generalization of the findings of the current studies  
302 imprecise. The whole metagenome sequencing method, although more expensive and  
303 requiring advanced bioinformatics skills, would provide more detailed information of the  
304 exact bacterial species and detects also other microorganisms within the sample, such as  
305 viruses, fungi, archaea and other microeukaryotes.

306 Another important aspect that calls for caution when interpreting the previous findings is  
307 that most of the studies have been performed on limited sample size, lacking detection

308 power and negative/positive controls. It is known that inherent elements of study design,  
309 such as sample size, sample collection method, DNA extraction process, type of  
310 sequencing employed, and data analysis, represent limitations that can influence the  
311 accurate detection of microorganisms [68, 69].

312 Furthermore, the currently applied next-generation sequencing-based microbiome  
313 analysis techniques assess DNA sequences, which do not necessarily equate with the  
314 presence of live bacteria [70]. Thus, DNA-based techniques characterize a microbiome  
315 but do not mean that the detected sequences are functionally active microbes. RNA  
316 analysis-based technique (i.e. meta-transcriptomics [71] and culturomics [72], together  
317 with integration with other omics analysis platforms would provide further knowledge of  
318 the functionality of the microbes in the gut in reproductive health and disease.

319 In conclusion, the gut microbiome studies in the female gynecological health are in its  
320 infancy, and future research on bigger, well-designed studies together with novel methods  
321 are warranted to unravel the core microbial compositions in gynecological health and to  
322 understand the function of specific microbes in the disease development.

323

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333

334 **CONFLICTS OF INTEREST**

335 The authors have no conflicts of interest.

336

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553 **FIGURE LEGEND**

554 **Figure 1. Potential gut microbiome-driven mechanisms underlying gynecological**  
555 **physiology and pathophysiology.** In healthy (eubiotic) conditions, the estrobolome  
556 contributes to estrogen activation through the secretion of beta-glucuronidase.  
557 Homeostatic circulating estrogen levels regulate menstrual cycle and contribute to uterine  
558 health. Moreover, several gut microbes release short-chain fatty acids (SCFA; e.g.,  
559 butyrate, acetate and propionate), which participate as anti-inflammatory mediators  
560 maintaining gut barrier function and physiological inflammation. When the gut  
561 microbiome is disrupted (dysbiosis), the overgrowth of  $\beta$ -glucuronidase-producing  
562 bacteria may lead to hyperestrogenic levels commonly reported in different gynecological  
563 pathologies. On the other hand, gut dysbiosis can be linked to a reduction of SCFA-  
564 producing bacteria, resulting in increased pro-inflammatory mediators and systemic  
565 inflammation. \*Reported  $\beta$ -glucuronidase-producing genera associated with any  
566 gynecological disease in case-control studies. (Created with BioRender.com).