ORIGINAL ARTICLE



Lipidomic signatures from physically frail and robust older adults at hospital admission

Robinson Ramírez-Vélez · Nicolás Martínez-Velilla · María Correa-Rodríguez · Mikel L. Sáez de Asteasu · Fabricio Zambom-Ferraresi · Sara Palomino-Echeverria · Antonio García-Hermoso · Mikel Izquierdo

Received: 30 March 2021 / Accepted: 22 December 2021 © The Author(s) 2022

Abstract Identifying serum biomarkers that can predict physical frailty in older adults would have tremendous clinical value for primary care, as this condition is inherently related to poor quality of life and premature mortality. We compared the serum lipid profile of physically frail and robust older adults to identify specific lipid biomarkers that could be used to assess physical frailty in older patients at hospital admission. Fortythree older adults (58.1% male), mean (range) age 86.4 (78–100 years) years, were classified as physically frail (*n*=18) or robust (*n*=25) based on scores from the Short Physical Performance Battery (≤ 6 points). Nontargeted metabolomic study by ultra-high performance liquid chromatography coupled to mass spectrometry

Supplementary Information The online version contains supplementary material available at https://doi.org/10.1007/s11357-021-00511-1.

R. Ramírez-Vélez · N. Martínez-Velilla ·

M. L. Sáez de Asteasu · F. Zambom-Ferraresi ·

S. Palomino-Echeverria · A. García-Hermoso ·

M. Izquierdo (🖂)

Navarrabiomed, Hospital Universitario de Navarra (HUN), Navarra Institute for Health Research (IdiSNA), Universidad Pública de Navarra (UPNA), Pamplona, Spain e-mail: mikel.izquierdo@gmail.com; robin640@hotmail. com; nicolas.martinez.velilla@navarra.es; macoro@ugr.es; mikel.lopez.saezdeasteasu@gmail.com; fabriciogigante@ hotmail.com; spalomie@navarra.es; antonio.garciah@ unavarra.es

R. Ramírez-Vélez · N. Martínez-Velilla ·

M. L. Sáez de Asteasu · F. Zambom-Ferraresi ·

(UHPLC-MS) analysis with later bioinformatics data analysis. Once the significantly different metabolites were identified, the KEGG database was used on them to establish which were the metabolic pathways mainly involved. Area under receiver-operating curve (AUROC) analysis was used to test the discriminatory ability of lipid biomarkers for frailty based on the Short Physical Performance Battery. We identified a panel of five metabolites including ceramides Cer (40:2), Cer (d18:1/20:0), Cer (d18:1/23:0), cholesterol, and phosphatidylcholine (PC) (14:0/20:4) that were significantly increased in physically frail older adults compared with robust older adults at hospital admission. The most interesting in the physically frail metabolome study found with the KEGG database were the metabolic pathways, vitamin digestion and absorption, AGE-RAGE signaling pathway in diabetic complications,

M. Izquierdo

CIBER of Frailty and Healthy Aging (CIBERFES), Instituto de Salud Carlos III, Madrid, Spain

M. Correa-Rodríguez

Department of Nursing, Health Sciences Faculty, University of Granada, Avda. De la Ilustración 60, 18016 Granada, Spain

A. García-Hermoso Laboratorio de Ciencias de La Actividad Física, El Deporte Y La Salud, Universidad de Santiago de Chile, USACH, Santiago, Chile and insulin resistance. In addition, Cer (40:2) (AUROC 0.747), Cer (d18:1/23:0) (AUROC 0.720), and cholesterol (AUROC 0.784) were identified as higher values of physically frail at hospital admission. The non-targeted metabolomic study can open a wide view of the physically frail features changes at the plasma level, which would be linked to the physical frailty phenotype at hospital admission. Also, we propose that metabolome analysis will have a suitable niche in personalized medicine for physically frail older adults.

Keywords Frailty · Lipidomic · Ceramides · Cholesterol · Phosphatidylcholines · Biomarker · Older adults

Introduction

The concept of frailty is gaining traction as an independent clinical syndrome to explain differential outcomes in older adults ¹. Weakness, slowness, and poor balance are some of the main physical manifestations of frailty, which is associated with adverse events that can lead to a dramatic decline in the quality of life and independence ². It is estimated that 25-50% of all people over the age of 85 are frail and are, consequently, at significantly higher risk of falls, disability, long-term care, and death ³.

Frailty is characterized by a host of manifestations, including metabolic abnormalities, neuroendocrine dysregulation, and impaired immunity, and may also be influenced by underlying genetic and environmental factors (e.g., physical inactivity, smoking, and unhealthy diet). Identifying populations at-risk and exploratively identifying potential early and/or predictive biomarkers of age-associated changes that could guide effective health promotion interventions are especially relevant ⁴. One of these strategies has been the investigation of the metabolome, the complete set of molecular weight molecules (metabolites) present in a biological sample. Pujos-Guillot et al. recently used an untargeted metabolomic approach to identify several circulating metabolites that were predictive of pre-frailty ⁵. Metabolomic approaches that analyze changes in metabolite levels thus seem promising for the development of non-invasive diagnostic biomarkers of the frailty phenotype, yet data from older adults at hospital admission are sparse.

The multidimensional nature of frailty presents challenges in accurately measuring the degree of

frailty, and several instruments have been developed in recent years to assess the frailty phenotype, although most are focused on different subpopulations of the elderly ⁶. Given the established relationship between functional decline, disability, and comorbidity, an objective clinical approach to assess physical status is to have a "proxy-frailty" measure based on, for example, the Short Physical Performance Battery (SPPB) ⁷. The predictive role of this clinical tool for future disabilities, frailty, and other adverse outcomes including institutionalization, hospital admissions, and mortality has been confirmed in several clinical settings ^{7–9}. The SPPB is, however, essentially a descriptive clinical tool and is therefore unable to assess the molecular/biochemical modifications underlying the loss of functionality.

While the causal processes involved in the pathogenesis of frailty are not fully understood, growing evidence supports a role for lipid metabolism in the risk for frailty by modulating skeletal muscle mass and function ¹⁰. To date, however, few studies have conducted broad lipidomic analyses to assess the relationship between frailty and blood lipids ^{11–13}. In the present study, we explored the potential of a targeted lipidomic approach to better characterize the biochemistry underlying physical frailty and to identify molecular biomarkers that might be useful to improve the clinical assessment of frailty in acutely hospitalized older patients at admission. To do this, we compared plasma lipidomic profile differences between physically frail and robust older adults at hospital admission.

Materials and methods

Patients and study design

In this study, we enrolled 43 elderly patients from 2018 to 2020 admitted from 2018 to 2020 to an Acute Geriatric Ward of a tertiary Hospital in Spain. Inclusion criteria were age \geq 70 years, Barthel Index score \geq 60 points, and ability to ambulate (with/without assistance) and to communicate and collaborate with the research team. Exclusion criteria were very severe cognitive decline (a score of 7 on the Global Deterioration Scale), terminal illness, uncontrolled arrhythmias, acute pulmonary embolism or myocardial infarction, or extremity bone fracture in the past

3 months. Participants signed an informed consent approved by the ethics committee before entering the study. The study followed the principles of the Declaration of Helsinki and was approved by the local Research Ethics Committee (ID Pyto2018/7; 15 May 2018). Patients were evaluated by a staff of 8 geriatricians at the moment of admission and were classified as physically frail (n=18) or robust (n=25) based to their performance on the SPPB.

Clinical and functional parameters

Height was measured to the nearest 0.1 cm and body mass was measured to the nearest 100 g. Body mass index (BMI) was computed as weight in kilograms divided by height in meters squared. Handgrip strength was measured in the seated position using a Takei 5401 digital dynamometer (Takei Scientific Instruments Co., Tokyo, Japan). Patients were asked to perform 2 maximum force trials for each hand, with the dynamometer beside but not against their body, and the maximum value was used as the final score ¹⁴.

Physical frailty was measured by the SPPB, which combines balance, gait velocity, and leg strength (five times sit-to-stand test) in a single score from 0 (worst) to 12 (best). Gait speed was calculated for each participant using distance in meters and time in seconds, and was obtained by dividing the distance traveled (4 m) on a flat and unobstructed path by the time to cover that distance. The total SPPB score was obtained through summing the scores obtained from each component. The total score was categorized as follows: 0-6 points = physically frail and 7-12 points = robust. We based this cut-off on two studies. Guimarães Rocco et al.¹⁵ highlighted an SPBB cutoff point < 6 as the most suitable for screening physically frail older people in their sample, with a sensitivity of 0.52, a specificity of 0.70, and an accuracy 0.88. Another study, carried out in Spain, also found that the best cut-off point was ≤ 6 , with an area under the curve (AUC) of 0.956 and sensitivity and specificity of 0.88^{9} .

Cognitive function was assessed with the Mini-Mental State Examination (MMSE, 30-point questionnaire; scale of 0 [worst] to 30 [best]) ¹⁶; and activities of daily living (ADLs) was assessed with the Barthel Index, with a scale of 0 (severe functional dependence) to 100 (functional independence) ¹⁷. Data related to number of diseases, cumulative illness rating scale score for geriatrics (CIRS-G), and length of hospital stay were collected from clinical records.

Lipid extraction and UHPLC-MS analysis

Fasting venous blood samples were collected on the next morning after admission to the hospital from the antecubital vein (08:00 to 09:00 am). Blood was inverted five times and allowed to sit for 30 min for clotting. Samples were then centrifuged at $2,000 \times g$ for 10 min at 4 °C, and plasma was aliquoted and stored at - 80 °C until use. Metabolite extraction has been previously described in detail ²². Briefly, plasma extracts were mixed with sodium chloride (50 mM) and chloroform/methanol (2:1) in 1.5 mL microtubes on ice. After brief vortex mixing, the samples were incubated for 1 h at-20 °C. After centrifugation at $16,000 \times g$ for 15 min at 4 °C, the organic phase was collected and dried in a speed vacuum dryer. Dried extracts were reconstituted in acetonitrile/isopropanol (1:1), centrifuged $(18,000 \times \text{g for 5 min at 4 }^\circ\text{C})$, and transferred to vials for ultra-high performance liquid chromatography coupled to mass spectrometry (UHPLC-MS) analysis. Chromatographic separation and mass spectrometric detection conditions have been previously described ²². The UHPLC-MS-based platform was developed for optimal profiling of glycerolipids (di- and tri-glycerides), cholesteryl esters, sphingolipids (ceramides and sphingomyelins), and glycerophospholipids (diacylglycerophospholipids and 1-ether, 2-acylglycerophospholipids).

Data pre-processing

Data pre-processing was carried out using the Target-Lynx application manager for MassLynx 4.1 (Waters Corp., Milford, MA). The chromatography/MS features (as defined by retention time and mass-to-charge ratio pairs, Rt-m/z) used in the study were identified prior to the analysis, either by comparison of their accurate mass spectra and chromatographic Rt with those of available reference standards, or when these were not available, by accurate mass MS/MS fragment ion analysis. Lipid nomenclature and classification follow the LIPID MAPS convention (https:// www.lipidmaps.org). Additionally, once the features with significantly different levels ($P_{adjus} \leq 0.05$) between the study groups were identified, it was possible to establish the most involved metabolic pathways in the study. This approach was performed using the Client-side REST access to KEGG (KEG-GREST) package (https://www.genome.jp/kegg/).

Statistical analysis

Clinical and functional characteristic data for continuous variables are presented as mean (SD). Data normalization was performed by inclusion of multiple internal standards and pool calibration samples following a previously described procedure ²³. Briefly, logarithmic transformation of the lipidomic data was applied to transform non-normal data into data that follow a normal distribution. *Limma* package (3.34.9) was used to compute the differential expression on physically frail and robust samples ¹⁸. In addition, data was adjusted for age, sex, and BMI, in order to avoid the influence of confounding variables. Furthermore, ImFit function was used to perform the empirical Bayes (eBayes) statistics and a false discovery rate (FDR). The P-values obtained were adjusted according to the Benjamini and Hochberg method ¹⁹. The adjusted P-values < 0.05 were considered significant. Principal component analyses (PCA) between the study group were performed. Later, for each of these compounds, the FoldChangeLog was calculated to select the features with higher differences in plasma levels between the groups. These calculations were performed using the statistical software package R v.3.4.1 (R Development Core Team, 2017; http:// cran.r-project.org).

Area under receiver-operating curve (AUROC) analysis was used to test the discriminatory ability of lipid biomarkers for frailty based on the SPPB. Cut-off points were chosen based on the Youden index (J), which uses the point on the ROC curve that is farthest from the line of equality ²⁰. Validity measurements were calculated for the cut-off point of the SPPB scale and served as a basis for calculating the prevalence of physical frailty and the post-test probability. The positive likelihood ratio was also determined. AUROCs were compared by a non-parametric approach suggested by DeLong et al.²¹. Percent improvement in prediction error was calculated according to the following formula: 100×(AUROC_{Test-Score-AUROCReference-Score}) / 1-AUROC_{Reference-Score}), where Test-Score refers to the first (superior) and Reference-Score to the second (inferior) score. A P-value <0.05 was generally considered statistically significant. The STATA statistical package (Version 13.0, College Station, TX) was used to perform clinical data management and AUROC analyses.

Results

The clinical and functional characteristics of the study group are shown in Table 1. The study included 43 older adults (58.1% male) with a mean (SD) age of 86.4 (4.2) years (range, 78-100 years). As expected, the scores for SPPB, gait speed, and five times sitto-stand test were all significantly lower in the physically frail group than in the robust group, whereas the opposite was seen for functional dependence.

The untargeted metabolomic study of the physically frail and robust plasma samples performed by UHPLC-MS identified 250 common features among the study individuals (Electronic Supplementary Material 1), of which had statistically significant differences between the study groups, after "FDR" adjustment. PCA by the study group dispersion is shown in Fig. 1.

The outcome obtained by this selection criterion is presented in the heatmap displayed in Fig. 2, in which it was possible to isolate five features that were highly differentiated between the study groups. Changes were observed in both neutral lipids and phospholipids, including sphingolipids (ceramides [Cer]), cholesterol, and phosphatidylcholine (PC) which are also displayed in the boxplots of Fig. 3, and were possible to observe in the HMDB and LIPID MAPS database.

Regarding the most involved metabolic pathways in the physically frail metabolome study, it was possible to link features with significant differences between the study groups with 27 different metabolic pathways of the KEGG/HMDB databases. However, only 15% (n=4) of the metabolic pathways were linked by four metabolites; these metabolic pathways are shown in Table 2.

The AUROC analyses of five plasma metabolites in identification of physically frail at hospital admission are shown in Table 3. The AUROC of the cholesterol with regard to presence of physically frail was similar to Cer (40:2) biomarkers (Fig. 4); however, no differences in AUROCs were also observed.

Table 1 Clinical and functional characteristics at hospital admission

Variable	Full sample $n=43$	Physically frail $n = 18$	Robust $n=25$	<i>P</i> for group
Sex (male) (n (%))*	25 (58.1)	15 (83.3)	10 (40.0)	0.472
Age (years)	86.4 (4.3)	87.8 (3.9)	85.4 (4.3)	0.075
Body mass (kg)	69.0 (15.9)	71.4 (12.9)	67.2 (17.8)	0.401
BMI (kg/m ²)	27.5 (5.4)	27.2 (5.2)	27.7 (5.6)	0.790
CIRS-G score ^b	11.9 (6.3)	11.0 (7.0)	12.9 (5.4)	0.405
Length of hospital stay (days)	7.2 (2.0)	7.2 (1.2)	7.0 (1.6)	0.560
SPPB (score) ^c	6.1 (2.9)	3.4 (1.3)	8.1 (2.0)	< 0.001
Gait speed (s (6 m))	10.8 (5.1)	13.7 (6.1)	8.8 (2.8)	< 0.001
Five times sit-to-stand test (s)	17.9 (8.5)	24.9 (11.9)	15.1 (4.4)	0.002
MMSE (score ^d)	23.0 (4.2)	22.8 (3.8)	23.1 (4.6)	0.829
Barthel Index (ADL) (score) ^e	76.3 (17.4)	69.7 (16.4)	81.5 (16.7)	0.030
Handgrip strength (kg)	17.6 (5.4)	17.4 (5.4)	17.7 (5.5)	0.853

Data are presented as the mean (SD), except for the sex group*. *BMI*, body mass index; *CIRS-G*, Cumulative Illness Rating Scale for Geriatrics; *SPPB*, Short Physical Performance Battery. ^aThe most prevalent diseases were coronary, pulmonary, genitourinary, and neurologic diseases. ^bThe CIRS-G scale evaluates individual body systems, ranging from 0 (best) to 56 (worst). ^cThe SPPB scale ranges from 0 (worst) to 12 (best). ^d*MMSE*, The Mini-Mental State Examination ranges from 0 (worst) to 30 (best). ^eThe Barthel Index ranges from 0 (severe functional dependence) to 100 (functional independence)



Fig. 1 Principal component analysis (PCA) between the study groups. Score plot of PCA using 250 features identified by the metabolomic study between the study groups. The orange points correspond to physically frail subjects, and the green points correspond to and robust subjects

Discussion

Frail older people are highly vulnerable to negative health outcomes such as falls, fractures, disability, and dementia, which is associated with poor quality of life ²². Consequently, there is a need for developing new biomarkers to predict frailty in the older population. In the present study, we characterized the plasma lipidomic profiles of physically frail and robust older

adults at hospital admission using targeted UHPLC-MS. We found that the circulating plasma levels of five metabolites including ceramides, cholesterol, and phosphatidylcholines were significantly lower in physically frail older adults. Among them, cholesterol and ceramides had acceptable levels of accuracy to discriminate physically frail at hospital admission and, accordingly, might be a useful biomarker in clinical practice.

The development of serum biomarkers for frailty in older adults would be enormously valuable in clinical practice because of its association with premature mortality ²³ and the inevitable increased cost and use of healthcare resources ²⁴. Our analysis revealed a signature of five metabolites including ceramides Cer (40:2), Cer (d18:1/20:0), Cer (d18:1/23:0), cholesterol, and PC (14:0/20:4) that were significantly increased in physically frail compared to robust older adults at hospital admission. Interestingly, a recent metabolome study also showed that some features were linked to some pathophysiological alterations of sarcopenia ²⁵.

We identified differential expression of ceramides Cer (40:2), Cer (d18:1/20:0), and Cer (d18:1/23:0), which were all significantly higher in physically frail older adults. Sphingolipids, including ceramides, have their essential biological roles to influence aging



Fig. 2 Heatmap of Cer (40:2), Cer (d18:1/20:0), Cer (d18:1/23:0), cholesterol, and PC (14:0/20:4) after logarithmic transformation of the data

²⁶. Ceramides increase in concentration with age in mammals and have been linked to various age-related ailments, including cancer, type 2 diabetes, neurode-generation, immune dysfunction, and cardiovascular disease ²⁶. Huang et al. also have showed that ceramide accumulation is also correlated with increased insulin resistance and oxidative stress ²⁷. Interestingly, Chaurasia et al. found that the deletion of dihy-droceramide desaturase 1 improves insulin resistance and hepatic steatosis in mice ²⁸. Thus, it has been proposed that clinical therapies that reduce ceramide concentrations may delay or ameliorate symptoms of aging in humans.

Our study also supports that cholesterol, which had a ROC AUC value greater than 0.75 (defined as being of good diagnostic value), might be useful to discriminate physically frail from robust older adults. Several studies have examined the usefulness of different lipids or lipid-related proteins as biomarkers of aging or age-related disease ²⁶. Interestingly, this metabolite has been previously associated with hyper-cholesterolemia and hyperlipoproteinemia ^{29,30}, which are well-known cardiovascular risk factors related to atherosclerosis. In addition, due to the role of cholesterol in myelin maintenance and cognition in schiz-ophrenia, it has been considered being potentially relevant in neurological disorders. Al Awam et al.

identified cholesterol as a good biomarker to discriminate between schizophrenia patients and unaffected controls ³¹. Among the same line, previous studies have suggested a role for steroid analysis as a potential diagnostic tool ^{32–34}. Our findings suggest that cholesterol might be relevant for frailty development. However, since the pathophysiological alterations of cholesterol metabolism associated with frailty are still incompletely understood, further studies are needed to understand the underlying mechanisms for the breakdown in the cholesterol machinery.

We also identified differential expression of PC (14:0/20:4), which was significantly higher in physically frail older adults. Note that this phosphatidyl-choline has been previously proposed as an obesity biomarker ^{35–37}. Phosphatidylcholine is a major phospholipid of cellular membrane and is a well-known marker of age-related membrane degeneration ³⁸. Moreover, it is worth noting that deregulation of lipid metabolism markers such as phosphatidylcholine has been reported to contribute to the link between physical frailty and cognitive decline ³⁹.

The results of the present study may have important implications for clinical management and public health surveillance. We identify five biomarkers that could facilitate the stratification of physically frail older adults, providing a clinical resource for



Fig. 3 Boxplots of Cer (40:2) (upper left panel), Cer (d18:1/20:0) (upper right panel), Cer (d18:1/23:0) and cholesterol (middle panel), and PC (14:0/20:4) (bottom panel)

after logarithmic transformation of the data. FoldChangeLog value for each metabolite was 0.94, 0.87, 0.87, 0.80, and 0.79, respectively

 Table 2
 The metabolic pathways most involved in the physically frail metabolome study

Metabolite (common name)	Chemical formula	KEGG code	KEGG pathways/biological process	KEGG map code
Cholesterol	C ₂₇ H ₄₆ O	C00187	Lipid and atherosclerosis	map05417
			Vitamin digestion and absorption	map04977
			Steroid hormone biosynthesis	map00140/map00100
			Metabolic pathways	map01100
			Fat digestion and absorption	map04975
			Cholesterol metabolism	map04979
			Overview of biosynthetic pathways	map01010
			Bile secretion	map04976
			Primary bile acid biosynthesis	map00120
			Steroid degradation	map00984
			Microbial metabolism in diverse envi- ronments	map01120
			Ovarian steroidogenesis	map04913
			Aldosterone synthesis and secretion	map04925
			Cortisol synthesis and secretion	map04927
PC (14:0/20:4)	C42H76NO8P	C00157	Biosynthesis of secondary metabolites	map01110
			Metabolic pathways	map01100
			alpha-Linolenic acid metabolism	map00592
			Linoleic acid metabolism	map00591
			Arachidonic acid metabolism	map00590
			Glycerophospholipid metabolism	map00564
Cer (d18:1/20:0)	C ₃₈ H ₇₅ NO ₃	C00195	Sphingolipid signaling pathway	map04071
			Insulin resistance	map04931
			Sphingolipid metabolism	map00600
			Metabolic pathways	map01100
			Neurotrophin signaling pathway	map04722
			Adipocytokine signaling pathway	map04920
			AGE-RAGE signaling pathway in diabetic complications	map04933
			Diabetic cardiomyopathy	map05415
Cer (d18:1/23:0)	$C_{41}H_{81}NO_3$	HMDB0000950*	Lipid peroxidation	-
			Insulin signaling pathway	-
			Apoptosis	_
			Lipid metabolism pathway	_
			Phospholipid metabolism	_
			Lipid transport	_
			Lipid metabolism	_
			Fatty acid metabolism	-

We used the metabolic pathways by the Kyoto Encyclopedia of Genes and Genomes (KEGG) database were used for searches metabolites by their chemical name in the database and their KEGG codes were registered. This code locates the pathways where the metabolite is involved. Then, the script program obtains the most involved metabolic KEGG pathways by counting the number of metabolites (significantly differentiated) involved in each pathway (https://www.genome.jp/kegg/). *Lipid nomenclature and classification follow the LIPID MAPS convention (https://www.lipidmaps.org) from The Human Metabolome Database (HMDB) ID. –, no informed

Parameter	Cer (40:2)	Cer (d18:1/20:0)	Cer (d18:1/23:0)	Cholesterol	PC (14:0/20:4)
AUC (SE)	0.747 (0.0751)	0.689 (0.0825)	0.720 (0.0791)	0.784 (0.0732)	0.611 (0.0885)
95% CI	0.591 to 0.867	0.530 to 0.821	0.562 to 0.846	0.633 to 0.895	0.450 to 0.756
P-value	< 0.0001	< 0.0001	< 0.0001	< 0.0001	< 0.0001
Youden index	0.5044	0.3689	0.3689	0.6089	0.2489
Cut-off ^a	0.457	0.4724	0.3862	0.6653	0.3386
Sensitivity	56.00	48.00	48.00	72.00	36.00
Specificity	94.44	88.89	88.89	88.89	88.89
+LR	10.08	4.32	4.32	6.48	3.24
-LR	0.47	0.59	0.59	0.32	0.72
+ PV	93.3	85.7	85.7	90.0	81.8
-PV	60.7	55.2	55.2	69.6	50.0

Table 3 Performance of ROC-derived cut-off values for Cer (40:2), Cer (d18:1/20:0), Cer (d18:1/23:0), cholesterol, and PC (14:0/20:4) in identification of physically frail at hospital admission

AUC, area under the curve; SE, standard error; CI, confidence interval; +PV, positive predictive value; -PV, negative predictive value; +LR, likelihood ratio positive; -LR, likelihood ratio negative. ^aMost suitable threshold according to ROC analysis and Youden's J statistic



Fig. 4 Performance of five plasma metabolites in prediction of physically frail. Receiver operating characteristic (AUROC, left panel) and precision-recall cures (AUPRC, right panel) for

diagnosis that supports the importance of lipidomic evaluation in this population. Future longitudinal studies should investigate the potential of these biomarkers as predictors for frailty development. Likewise, given the complexity of frailty phenotype, exploratory approaches will be needed to allow the identification of specific signatures relevant to distinguish frailty status. Of note, metabolic pathways, vitamin digestion and absorption, the AGE-RAGE signaling pathway in diabetic complications, and



different plasma metabolite scores in prediction of physically frail at hospital admission

insulin resistance were linked by four of the metabolites identified in this study. More research is needed to determine the main consequences of the aberrant lipidomic profile in frail older adults and to provide more insight into the metabolic pathways in frailty. The identification of metabolomic profiles of physically frail older adults may allow the design of strategic interventions and approaches for delaying the progression of frailty in older populations, and the demonstration of changes in some lipid metabolomic biomarkers prior to the onset of frailty could provide a basis for future preventative interventions.

Our present study is not without limitations. First, our analysis was cross-sectional, and so causative relationships could not be made. Longitudinal studies will be needed to determine temporal relationship between changes in the lipidomic profile and the development of physical frailty. Second, the study population comprised a cohort including only Caucasians, which impedes the generalization of our findings to other ethnic groups. Additionally, although the cohort was extensively characterized, it was relatively small and analyses involved a large set of variables. Even considering these limitations, the study included 250 metabolic features and UHPLC-MS was used for the analyses, which has been established as a sensitive and highly reliable method for lipidomic analysis ⁴⁰. Also, we investigated, for the first time to our knowledge, the correlation between serum lipid changes in physically frail and robust patients using a targeted lipidomic method, which could be a good starting point to further investigate the predictive value of the specific lipidomic signatures of frailty.

Conclusion

In summary, the present targeted lipidomic approach found that five circulating metabolites including ceramides, cholesterol, and phosphatidylcholines were significantly increased in physically frail compared with robust older adults at hospital admission, pointing to the existence of a lipidomic profile of older adults with physical frailty. Moreover, cholesterol and ceramides had acceptable levels of accuracy to discriminate physically frail at hospital admission and, therefore, might be useful biomarkers in clinical practice. The non-targeted metabolomic study can open a wide view of the physically frail features changes at the plasma level, which would be linked to the physical frailty phenotype at hospital admission. Also, we propose that metabolome analysis will have a suitable niche in personalized medicine for physically frail older adults. Larger-scale longitudinal studies will be needed as a next step to provide novel insights about aberrant lipid metabolism and the development of physical frailty in older adults.

Acknowledgements We thank Fundacion Miguel Servet (Navarrabiomed) for its support during the implementation of the trial, as well as Fundación Caja Navarra and Fundación La Caixa. Finally, we thank our patients and their families for their confidence in the research team. We are grateful to Drs Cristina Alonso and Irantzu Pérez from the OWL Metabolomics for assistance with statistical analysis.

Funding Open Access funding provided thanks to the CRUE-CSIC agreement with Springer Nature. This study was funded by a Gobierno de Navarra project Resolución grant 2186/2014 and acknowledged with the "Beca Ortiz de Landázuri" as the best research clinical project in 2014, as well as by a research grant PI17/01814 of the Ministerio de Economía, Industria y Competitividad (ISCIII, FEDER). A.G.-H. is a Miguel Servet Fellow (Instituto de Salud Carlos III – CP18/0150). N.M.-V. received funding from "la Caixa" Foundation (ID 100010434), under agreement LCF/PR/PR15/51100006. R.R.-V. is funded in part by a Postdoctoral Fellowship Resolution ID 420/2019 of the Universidad Pública de Navarra.

Data availability The data that support the findings of this study are available from the corresponding author upon reasonable request.

Declarations

Ethics approval The study followed the principles of the Declaration of Helsinki and was approved by the local Research Ethics Committee (ID Pyto2018/7; 15 May 2018).

Conflict of interest The authors declare no competing interests.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

References

 Hoogendijk EO, Afilalo J, Ensrud KE, Kowal P, Onder G, Fried LP. Frailty: implications for clinical practice and public health. Lancet. 2019;394(10206):1365–75. https:// doi.org/10.1016/S0140-6736(19)31786-6.

- Clegg A, Young J, Iliffe S, Rikkert MO, Rockwood K. Frailty in elderly people. In: *The Lancet*. Vol 381. Lancet Publishing Group; 2013:752–762. https://doi.org/10.1016/ S0140-6736(12)62167-9
- Song X, Mitnitski A, Rockwood K. Prevalence and 10-year outcomes of frailty in older adults in relation to deficit accumulation. J Am Geriatr Soc. 2010;58(4):681– 7. https://doi.org/10.1111/j.1532-5415.2010.02764.x.
- Yin X, Willinger CM, Keefe J, et al. Lipidomic profiling identifies signatures of metabolic risk. *EBioMedicine*. 2020;51. https://doi.org/10.1016/j.ebiom.2019.10.046
- Pujos-Guillot E, Pétéra M, Jacquemin J, et al. Identification of pre-frailty sub-phenotypes in elderly using metabolomics. *Front Physiol*. 2019;10(JAN).https://doi.org/10. 3389/fphys.2018.01903
- de Vries NM, Staal JB, van Ravensberg CD, Hobbelen JSM, Olde Rikkert MGM, Nijhuis-van der Sanden MWG. Outcome instruments to measure frailty: a systematic review. Ageing Res Rev. 2011;10(1):104–114. https://doi. org/10.1016/j.arr.2010.09.001
- Guralnik JM, Ferrucci L, Simonsick EM, Salive ME, Wallace RB. Lower-extremity function in persons over the age of 70 years as a predictor of subsequent disability. N Engl J Med. 1995;332(9):556–62. https://doi.org/10.1056/ nejm199503023320902.
- da Câmara SMA, Alvarado BE, Guralnik JM, Guerra RO, Maciel ÁCC. Using the Short Physical Performance Battery to screen for frailty in young-old adults with distinct socioeconomic conditions. Geriatr Gerontol Int. 2013;13(2):421–8. https://doi.org/10.1111/j.1447-0594. 2012.00920.x.
- Abizanda P, Romero L, Sánchez-Jurado PM, Atienzar-Núñez P, Esquinas-Requena JL, García-Nogueras I. Association between functional assessment instruments and frailty in older adults: the FRADEA study. *J frailty aging*. 2012;1(4):162–168. https://doi.org/10.14283/jfa.2012.25
- Lipina C, Hundal HS. Lipid modulation of skeletal muscle mass and function. J Cachexia Sarcopenia Muscle. 2017;8(2):190–201. https://doi.org/10.1002/jcsm.12144.
- Corsetto PA, Montorfano G, Klersy C, et al. Fatty acid profile and antioxidant status fingerprint in sarcopenic elderly patients: role of diet and exercise. *Nutrients*. 2019;11(11). https://doi.org/10.3390/nu11112569
- Corona G, Polesel J, Fratino L, et al. Metabolomics biomarkers of frailty in elderly breast cancer patients. J Cell Physiol. 2014;229(7):898–902. https://doi.org/10.1002/ jcp.24520.
- Liu Z, Burgess S, Wang Z, et al. Associations of triglyceride levels with longevity and frailty: a Mendelian randomization analysis. *Sci Rep.* 2017;7:41579–41579. https:// doi.org/10.1038/SREP41579
- Ramírez-Vélez R, Sáez de Asteasu ML, Martínez-Velilla N, Zambom-Ferraresi F, García-Hermoso A, Izquierdo M. Handgrip strength as a complementary test for mobility limitations assessment in acutely hospitalized oldest old. *Rejuvenation Res.* Published online January 7, 2021. https://doi.org/10.1089/rej.2020.2344
- Rocco LLG, Fernandes TG. Validity of the short physical performance battery for screening for frailty syndrome among older people in the brazilian amazon region. A cross-sectional study. Sao Paulo Med J.

2020;138(6):537-544. https://doi.org/10.1590/1516-3180. 2020.0264.r1.14092020

- Folstein MF, Folstein SE, McHugh PR. "Mini-mental state". A practical method for grading the cognitive state of patients for the clinician. J Psychiatr Res. 1975;12(3):189–198. https://doi.org/10.1016/0022-3956(75)90026-6
- Shah S, Vanclay F, Cooper B. Improving the sensitivity of the Barthel Index for stroke rehabilitation. J Clin Epidemiol. 1989;42(8):703–9. https://doi.org/10.1016/0895-4356(89)90065-6.
- Ritchie ME, Phipson B, Wu D, et al. limma powers differential expression analyses for RNA-sequencing and microarray studies. Nucleic Acids Res. 2015;43(7):e47– e47. https://doi.org/10.1093/NAR/GKV007.
- Benjamini Y, Hochberg Y. Controlling the false discovery rate: a practical and powerful approach to multiple testing. J R Stat Soc Ser B. 1995;57(1):289–300. https://doi.org/ 10.1111/J.2517-6161.1995.TB02031.X.
- Fluss R, Faraggi D, Reiser B. Estimation of the Youden Index and its associated cutoff point. *Biom J*. 2005;47(4):458–472. Accessed January 12, 2018. http:// www.ncbi.nlm.nih.gov/pubmed/16161804
- DeLong ER, DeLong DM, Clarke-Pearson DL. Comparing the areas under two or more correlated receiver operating characteristic curves: a nonparametric approach. *Biometrics*. 1988;44(3):837–845. Accessed April 24, 2018. http://www.ncbi.nlm.nih.gov/pubmed/3203132
- Kojima G, Liljas AEM, Iliffe S. Frailty syndrome: implications and challenges for health care policy. Risk Manag Healthc Policy. 2019;12:23–30. https://doi.org/10.2147/ RMHP.S168750.
- Stow D, Matthews FE, Barclay S, et al. Evaluating frailty scores to predict mortality in older adults using data from population based electronic health records: Case control study. Age Ageing. 2018;47(4):564–9. https://doi.org/10. 1093/ageing/afy022.
- García-Nogueras I, Aranda-Reneo I, Peña-Longobardo LM, Oliva-Moreno J, Abizanda P. Use of health resources and healthcare costs associated with frailty: the FRADEA study. J Nutr Heal Aging. 2017;21(2):207–14. https://doi. org/10.1007/s12603-016-0727-9.
- Opazo R, Angel B, Márquez C, et al. Sarcopenic metabolomic profile reflected a sarcopenic phenotype associated with amino acid and essential fatty acid changes. *Metabolomics* 2021 179. 2021;17(9):1–12. https://doi.org/10. 1007/S11306-021-01832-0
- Johnson AA, Stolzing A. The role of lipid metabolism in aging, lifespan regulation, and age-related disease. *Aging Cell*. 2019;18(6). https://doi.org/10.1111/acel.13048
- Huang X, Withers BR, Dickson RC. Sphingolipids and lifespan regulation. Biochim Biophys Acta - Mol Cell Biol Lipids. 2014;1841(5):657–64. https://doi.org/10.1016/J. BBALIP.2013.08.006.
- Chaurasia B, Tippetts TS, Monibas RM, et al. Targeting a ceramide double bond improves insulin resistance and hepatic steatosis. *Science (80-)*. 2019;365(6451):386– 392. https://doi.org/10.1126/SCIENCE.AAV3722
- Nelson RH. Hyperlipidemia as a risk factor for cardiovascular disease. Prim Care. 2013;40(1):195. https://doi.org/ 10.1016/J.POP.2012.11.003.

- Thelen KM, Laaksonen R, Päivä H, Lehtimäki T, Lütjohann D. High-dose statin treatment does not alter plasma marker for brain cholesterol metabolism in patients with moderately elevated plasma cholesterol levels. J Clin Pharmacol. 2006;46(7):812–6. https://doi.org/10.1177/ 0091270006289851.
- Al Awam K, Haußleiter IS, Dudley E, et al. Multiplatform metabolome and proteome profiling identifies serum metabolite and protein signatures as prospective biomarkers for schizophrenia. *J Neural Transm 2014 1221*. 2014;122(1):111–122. https://doi.org/10.1007/ S00702-014-1224-0
- Vila-Rodriguez F, Honer WG, Innis SM, Wellington CL, Beasley CL. ApoE and cholesterol in schizophrenia and bipolar disorder: comparison of grey and white matter and relation with APOE genotype. J Psychiatry Neurosci. 2011;36(1):47. https://doi.org/10.1503/JPN.090116.
- Woods AG, Sokolowska I, Taurines R, et al. Potential biomarkers in psychiatry: focus on the cholesterol system. J Cell Mol Med. 2012;16(6):1184. https://doi.org/10. 1111/J.1582-4934.2012.01543.X.
- Bicikova M, Hill M, Ripova D, Mohr P, Hampl R. Determination of steroid metabolome as a possible tool for laboratory diagnosis of schizophrenia. J Steroid Biochem Mol Biol. 2013;133(1):77–83. https://doi.org/10.1016/J. JSBMB.2012.08.009.
- Cho K, Moon JS, Kang J-H, et al. Combined untargeted and targeted metabolomic profiling reveals urinary biomarkers for discriminating obese from normal-weight adolescents. Pediatr Obes. 2017;12(2):93–101. https://doi. org/10.1111/ijpo.12114.

- Haro C, Montes-Borrego M, Rangel-Zúñiga OA, et al. Two healthy diets modulate gut microbial community improving insulin sensitivity in a human obese population. J Clin Endocrinol Metab. 2016;101(1):233–42. https://doi.org/10.1210/JC.2015-3351.
- Reinehr T, Wolters B, Knop C, et al. Changes in the serum metabolite profile in obese children with weight loss. *Eur J Nutr 2014 542*. 2014;54(2):173–181. https://doi.org/10. 1007/S00394-014-0698-8
- Henneberry AL, Wright MM, McMaster CR. The major sites of cellular phospholipid synthesis and molecular determinants of fatty acid and lipid head group specificity. Mol Biol Cell. 2002;13(9):3148–61. https://doi.org/10. 1091/mbc.01-11-0540.
- Ma L, Chan P. Understanding the physiological links between physical frailty and cognitive decline. *Aging Dis*. 2020;11(2):405–418. https://doi.org/10.14336/AD.2019. 0521
- Triebl A, Trötzmüller M, Hartler J, Stojakovic T, Köfeler HC. Lipidomics by ultrahigh performance liquid chromatography-high resolution mass spectrometry and its application to complex biological samples. J Chromatogr B Anal Technol Biomed Life Sci. 2017;1053:72–80. https:// doi.org/10.1016/j.jchromb.2017.03.027.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.