

Javier Bobo-Pinilla^{1,2,} ^(b), Noemí López-González^{1,2} ^(b), Armando Caballero³ ^(b) & Julio Peñas⁴

Received: 5 March 2020 / Accepted: 23 July 2020 / Published online: 24 June 2021

Abstract. *Astragalus edulis* (Fabaceae) is an endangered annual plant species from the western Mediterranean region. It occurs in the Iberian Peninsula (SE), Morocco (two main locations in the NE and SW), and the easternmost Canary Islands (Lanzarote and Fuerteventura). After a thorough study assessing genetic diversity and phylogeographic patterns, and a proposal of a management plan to preserve the genetic diversity across the entire area of the species, it is about time to go further in the species conservation. It is necessary to recognize genetically impoverished populations to be able to successfully reinforce populations and find realistic places for translocations. For this, Nei's genetic diversity values were calculated for the three geographic areas and for their populations. Diversity values were further evaluated to calculate the optimal contribution for the populations within each area to maximize the genetic pool. To evaluate whether the optimum habitat for the species is expected to grow or decrease under a climate change scenario, the optimal model of current climatic suitability of *A. edulis* was projected into the IPCC (2070_RCP8.5) future scenario. The different geographical areas showed similar genetic diversity values. The populations that contribute to maximize the genetic pool in each geographic area were similar, but not equal, to those proposed in previous studies. The future potential habitat for the species shows a significant range decline, which will affect translocation efforts.

Keywords: Conservation; genetic diversity; species distribution models; western Mediterranean-Macaronesian area.

How to cite: Bobo-Pinilla, J., López-González, N., Caballero, A. & Peñas, J. 2021. Looking for a successful translocation: the case of *Astragalus edulis*. Mediterr. Bot. 42, e68048. https://dx.doi.org/10.5209/mbot.68048

Introduction

Acquiring knowledge on species potential habitat, genetic diversity, and intraspecific phylogenetic relationships is essential to appropriately address species conservation (Commander et al., 2018). The genetic patterns of biodiversity must be preserved as they hold the survival ability of the species (Pérez-Collazos et al., 2008). Human-mediated changes to ecosystems and the climate change threaten species survival (Leadley et al., 2010). These perturbations often promote habitat fragmentation leading to isolation and limitation of gene flow among populations. As a consequence, species genetic diversity will be negatively affected (Young & Clarke, 2000). This isolation additionally implies a decrease in the resilience of populations when facing environmental changes. Urgent conservation measures and restoration plans are required given the current scenario of biodiversity loss due to rapid environmental change (Christmas et al., 2016; IUCN, 1998).

Climate change forces species to adapt, move (migrate), or become extinct (Christmas *et al.*, 2016; Nogués-Bravo *et al.*, 2018). Species can respond plastically to environmental variation being phenotypic plasticity considered a major evolutionary force (Noel

et al., 2007). Phenotypic plasticity can be adaptive (Thompson, 1991; Valladares et al., 2006) being this essential for conservation biology. Despite this, the ability to adapt is often limited when the rate of environmental change is high, and in this context the dispersal abilities of species can be vital in preventing extinction (Tremlová & Münzbergová, 2007). These displacements occur naturally through the dispersive abilities of the species (dispersal of seeds and propagules). However, dispersive processes are generally stochastic, so the probability that the species will colonize habitats with climatic and edaphic suitable characteristics is also unlikely (Nathan, 2006). Species distribution models (SDMs) provide essential information about the ecological requirements of the species. This can help to find the optimal areas for the species in the future (Elith & Leathwick, 2009; Guisan et al., 2013) and consequently it could be vital to ensure the success of conservation proposals (Dawson et al., 2011; IUCN/SSC, 2013).

Regarding *in situ* conservation, several estimators have been proposed to answer how many and which populations should be conserved; all of which are based on the preservation of the higher amount of genetic diversity in the least number of populations and/or areas (Ryder, 1986; Moritz, 1994; Riddle &

¹ Department of Botany, University of Salamanca. 37007, Salamanca, Spain. Email: javicastronuevo@usal.es

² Biobanco de ADN Vegetal, Edificio Multiusos I+D+i. 37007, Salamanca, Spain.

³ Departamento de Bioquímica, Genética e Inmunología and Centro de Investigación Mariña (CIM-UVIGO), Universidade de Vigo. 36310 Vigo, Spain.

⁴ Plant Conservation Unit, Department of Botany, University of Granada. 18071, Granada, Spain.

Hafner, 1999; Maes et al., 2004; Pérez-Collazos et al., 2008; among others). The selection of relevant genetic units for conservation (RGUCs) has been successfully used to propose sampling strategies for plant species, such as Boleum asperum Desv. (Pérez-Collazos et al., 2008), Borderea pyrenaica Miégev. (Segarra-Moragues & Catalán, 2010), and Astragalus edulis Bunge (Peñas et al., 2016). This method is based on the idea that rare alleles are essential in conservation as they represent unique evolutionary products that could provide the species with the ability to adapt to environmental changes (Pérez-Collazos et al., 2008). However, the conservation proposals are often focused on passive protection, which are often inappropriate for reducing accelerated losses of natural species and habitats (Fenu et al., 2019). The main objective of in situ conservation is to establish a method that combines the conservation of the entire genetic diversity with the socioeconomic viability of the conservation proposal. Thus, ex situ conservation is not feasible at large scales due to the costs, but it is feasible to apply to the most threatened species, which are precisely those that require greater ex situ conservation effort (Fay & Krauss, 2003). The gathering of all the genetic diversity of the species is essential to propose viable translocation measures (Caujapé-Castells & Pedrola-Monfort, 2004; Pearse & Crandall, 2004), and represents the basis of the creation and use of seed banks, the core of ex situ conservation strategy.

The reinforcement of existing populations and the creation of new populations (through the introduction of propagules or seeds to a new location within or outside the range of distribution of the species) are two types of translocation (Akeroyd & Wyse Jackson, 1995). The reinforcement of rare and threatened species populations has become essential for biodiversity conservation (Armstrong & Seddon, 2008). When reinforcing populations or creating new ones, it is common to use only a few individuals from one population or from several small populations, which inevitably leads to the failure of the conservation proposals (Godefroid et al., 2011). Small populations of endemic species and/or of fragmented habitat species hold limited genetic variation; therefore, increasing the size of the population may end up decreasing the number of individuals capable of reproducing within the populations, this is due to inbreeding depression that eventually reduces their genetic diversity (Frankham et al., 2002; Friar et al., 2000). Outbreeding depression must also be considered as it may influence the survival in species that inhabit ecologically or geographically different areas (Fenster & Galloway, 2000). The number of successful reintroductions of plant species is still small (Fenu et al., 2019). In order to reduce the failure rates, reintroductions require knowledge of the life history, demography, reproductive biology, ecology, and geography of the focal species (Armstrong &

Seddon, 2008; Bajomi *et al.*, 2010). The failure of the translocations has probably diverse causes, but the reasons are partially unknown due to the fact that the studies are focused on monitoring the establishment and not on the explanation about the failure (Armstrong & Seddon, 2008).

Astragalus edulis (Fabaceae) is an endangered annual endemism from the western Mediterranean hotspot region. It occurs in South-Eastern (SE) Iberian Peninsula, Morocco (two main locations in the North East and South West), and the easternmost Canary Islands (Lanzarote and Fuerteventura). Previous studies have assessed intraspecific genetic diversity and phylogeographic patterns of A. edulis, which have led to propose an *in situ* management plan to preserve the genetic diversity across the entire distribution area of the species (Bobo-Pinilla et al., 2018; Peñas et al., 2016). As described by Peñas (2004), the dispersal ability of the species is very limited. The low values of diversity and rarity found have been interpreted as a consequence of recent colonization events or reduction of the effective population sizes (Bobo-Pinilla et al., 2018). Moreover, this species represents an ideal study system to test the utility of Species Distribution Modeling (SDMs) and diversity patterns as a tool to assess how climate change will affect the potential distribution of endangered species from semiarid ecosystems and a way to preserve taxa that have highly fragmented populations.

In the present study, we propose the reinforcement planning of the most degraded populations of *A. edulis* in order to preserve its genetic diversity. In addition, we will use SDMs to infer the areas that will present optimal climatic conditions for the survival of the species in the future and consequently propose realistic translocations.

Materials and Methods

Genetic diversity patterns

Amplified fragment length polymorphisms (AFLP) of Astragalus edulis obtained by Peñas et al. (2016) were used in this study as a source of genetic data. This data includes a total of 358 plants from 17 populations (6 from the Iberian Peninsula, 8 from Morocco, and 3 from the Canary Islands; Table 1 and Figure 1). AFLP-SURV 1.0 (Vekemans et al., 2002) was used to calculate the Nei (1987) gene diversity index for each population (H_{nei}). Nei's gene diversity was also calculated for the three main geographical areas separately (H.; Table 1). The multilevel b-spline tool (Conrad et al., 2015) implemented in QGIS (QGIS-Development-Team, 2017) software was used to draw the genetic diversity patterns. This tool creates a diversity gradient map, interpolating the specific values of the populations.



Figure 1. Location of the populations of Astragalus edulis studied.

Table 1. Geographic and genetic diversity features of the populations of *Astragalus edulis* (N, Number of individuals used for the AFLP analyses; H_{Nei} , Nei's gene diversity index; H_t , Nei's average gene diversity values per geographical area) and Metapop2 v2.2.1 results (ΔH_{nei} , proportional increment/decrement of the within-population gene diversity when the population data is removed in the analysis; ΔH_{dist} , proportional increment/decrement/decrement of Nei's average genetic distance between populations when the population data is removed in the analysis; C_x , expected proportion of seeds from the populations within the geographic areas in order to obtain the maximum diversity values in a synthetic population).

| Area | Population | Ν | Longitude | Latitude | H _{nei} | H | ΔH_{nei} | ΔH_{dist} | C _x |
|-------------------|------------|----|-----------|----------|------------------|-------|------------------|-------------------|----------------|
| Iberian Peninsula | AE1 | 16 | -2.60 | 37.10 | 0.098 | 0.115 | 1.68 | -2.97 | 8.5 |
| | AE2 | 24 | -2.46 | 37.13 | 0.096 | | 2.00 | 0.77 | 0.0 |
| | AE3 | 32 | -2.53 | 37.12 | 0.109 | | -1.82 | 1.28 | 0.0 |
| | AE4 | 24 | -2.47 | 37.10 | 0.131 | | -4.96 | 0.28 | 30.9 |
| | AE5 | 23 | -2.49 | 37.07 | 0.133 | | -5.37 | -0.54 | 60.6 |
| | AE6 | 33 | -2.36 | 37.12 | 0.076 | | 7.78 | -1.23 | 0.0 |
| Morocco | AE7 | 17 | -2.60 | 34.42 | 0.069 | 0.106 | 3.48 | -2.42 | 0.0 |
| | AE8 | 21 | -3.35 | 34.47 | 0.107 | | -2.98 | -0.54 | 44.1 |
| | AE9 | 22 | -8.19 | 31.87 | 0.107 | | -2.43 | 1.17 | 15.3 |
| | AE10 | 14 | -8.62 | 31.57 | 0.084 | | 1.55 | -0.26 | 0.0 |
| | AE11 | 18 | -8.48 | 30.18 | 0.097 | | -0.57 | -0.26 | 17.5 |
| | AE12 | 19 | -8.45 | 30.05 | 0.095 | | 0.16 | 0.10 | 0.0 |
| | AE13 | 25 | -8.86 | 29.74 | 0.096 | | -1.00 | -1.15 | 23.1 |
| | AE14 | 7 | -8.94 | 29.64 | 0.084 | | 1.19 | -0.41 | 0.0 |
| Canary Islands | AE15 | 29 | -13.73 | 28.93 | 0.079 | 0.111 | 15.20 | -1.61 | 0.0 |
| | AE16 | 14 | -14.03 | 28.36 | 0.134 | | -7.69 | -2.68 | 82.5 |
| | AE17 | 22 | -13.99 | 28.27 | 0.111 | | -6.96 | -0.68 | 17.5 |

Contribution of the populations to global genetic diversity

In order to calculate the relative contribution of the populations to a genetic pool of the species in the different areas, the proportional contribution of each population to Nei's gene diversity (ΔH_{nei} ; Nei, 1987) and the proportional contribution of the average Nei's (ΔH_{dist}) minimum genetic distance between populations was calculated using the software Metapop2 v2.2.1 (López-Cortegano et al., 2019). These contributions are obtained by disregarding each population from the analysis one by one in each area and calculating the proportional amount of diversity gained or lost after the removal of the population data. The software also calculates the proportion of gene diversity explained within and among populations in each area, giving an approximation of the contribution of each population to the total diversity of each area (Petit et al., 1998). Moreover, Metapop2 v2.2.1 was also used to obtain the expected proportional contribution (C_{i}) of each population (within the areas) to a theoretical synthetic pool (e.g. germplasm bank) with maximum global gene diversity (D_{max}) (Table 1). This was done by maximizing the function $D_{max=1-\sum_{i=1}^{n} f_{ij} c_i c_j}$, where f_{ij} is the average coancestry between populations *i* and *j*, and *c*, is the contribution of subpopulation i to the pool (Toro & Caballero, 2005).

Species distribution modeling

The optimal model of current climatic-suitability of A. edulis (Bobo-Pinilla et al., 2018) was projected into the future scenario (2070 RCP8.5) climatic layers (available at www.worldclim.com) using the package 'raster' (Hijmans et al., 2005); the most restrictive future scenario was chosen to ensure that the areas found will have the climatic characteristics matching the habitat suitability of the species. The climatic features found to have an influence on the ecology and range limits of A. edulis are: temperature seasonality (bio4), precipitation of the wettest quarter (bio16), and precipitation of the driest quarter (bio17) (Bobo-Pinilla et al., 2018). QGIS (QGIS-Development-Team, 2017) software was used to search for matching areas of optimal habitat in current and future times; the values of habitat suitability over 0.5 at the present were compared with those over 0.5 in the future scenario (2070 RCP8.5).

Results

Genetic diversity patterns

Genetic Diversity values (H_{Nei}) ranged from 0.069 in the *Astragalus edulis* population AE7 (Morocco) to 0.134 in the population AE16 (Canary Islands). The average genetic diversity values (H_t) for the three areas considered were similar (0.115 for the Iberian Peninsula, 0.106 for Morocco, and 0.111 for the Canary Islands; Table 1).

Regarding the genetic diversity patterns, the Iberian populations showed the highest values of diversity in the central distribution area (populations AE3, AE4, and AE5, with 0.109, 0.131, and 0.133 respectively), whereas the westernmost and easternmost areas showed the lowest values (0.098 in population AE1 and 0.076 in population AE6; Figure 2A and Table 1), being population AE6 the most impoverished one. The Moroccan populations showed low diversity values, with the exception of populations AE9 and AE8 (with both having a value of 0.107; Figure 2B and Table 1). Thus, all populations in the south of the Atlas Mountains showed a pattern of low genetic diversity. With respect to the Canary Islands, the population of Lanzarote was found to have a remarkably low genetic diversity (0.079;Figure 2C and Table 1).

Contribution of the populations to global genetic diversity

The intra- and inter-population contributions to the total genetic diversity were 90.98% and 9.02% respectively for the Iberian Peninsula, 81.93% and 18.07% respectively for Morocco, and 96.75% and 3.25% respectively for the Canary Islands. In the Iberian Peninsula, populations AE4 and AE5 showed the largest loss of within-population diversity (ΔH_{nei} of 4.96% and 5.37%, respectively) when they are disregarded in the analysis (Table 1), indicating that these are the populations with a more important contribution to the intra-population diversity in the area. Population AE1 showed the largest contribution to between-population diversity (a loss of 2.97% of variation when removed from the analysis, ΔH_{dist}). In Morocco, populations AE8 and AE9 showed the largest contribution to within-population diversity, whereas AE7 and AE13 showed the largest contribution to between-population diversity. Finally, in the Canary Islands, populations AE16 and AE17 showed a substantial contribution to both within- and between-population diversity. The net overall contribution of the populations to global genetic diversity is clearly illustrated by the expected contributions of the populations to a pool of maximum diversity ($C_{..}$, Table 1), with contributions of AE1, AE4, and AE5 in the Iberian Peninsula, AE8, AE9, AE11, and AE13 in the Moroccan area, and AE17 and, particularly, AE16 in the Canary Islands.

Species distribution models

The current distribution of *A. edulis* mostly coincided with that predicted by the potential distribution of the species (Figure 3A). In general, the suitable habitat decreased significantly (Figure 3B), especially in the SE of the Iberian Peninsula and in the NE of Morocco. Only small regions of Lanzarote and the northern area situated at the SW part of the Atlas Mountains remained suitable for the species in the future considering its current distribution (Figure 3C).



Figure 2. Nei's gene diversity pattern (red = low; yellow = medium; green = high) obtained with QGIS (QGIS-Development-Team, 2017); A, Iberian Peninsula; B, Morocco; C, Canary Islands.



Figure 3. A, Distribution model representing the habitat suitability for the species in the present (red-yellow, medium, green, high; Bobo-Pinilla *et al.*, 2018); B, Projected habitat suitability values for the species in the future scenario 2070_RCP8.5 (red-yellow, medium, green, high); C, red areas represent suitable areas for the species in the present and future scenarios, circle represent present populations. (slightly modified from Bobo-Pinilla *et al.*, 2018).

Discussion

The mere designation of protected areas is not enough to protect biodiversity (Volis, 2019; and references within). A method that unifies the different aspects regarding the viability of populations as part of conservation proposals is thus necessary; such unification is especially deficient for *ex situ* conservation (Volis & Blecher, 2010). The aim of a successful proposal should be the combination of preserving the maximum variability and viability of the species as well as the efficient search for suitable locations for translocations of the species in a changing world.

Reinforcement of the genetically impoverished populations

The relevant genetic units for conservation (RGUCs) proposed for Astragalus edulis (Peñas et al., 2016) could be underestimating the importance of some populations with low genetic diversity. The proposal of RGUCs in Morocco was to preserve the populations AE8 and AE9; the results obtained here show that the optimal contributions to the genetic diversity of the Moroccan area would include these two populations with 44.1% and 15.3% respectively, but also populations AE11 and AE13 with 17.5% and 23.1% of the total Moroccan optimal seed proportion. The optimal proportions of the populations in the Iberian Peninsula to maximize the genetic pool corresponds with AE5 (60.6%), AE4 (30.9%), and AE1 (8.5%), coinciding with the proposal of RGUCs priority selection. In addition, the RGUC chosen for the Canary Islands was population AE16; the optimal contribution of this population to the maximization of the genetic diversity is 82.5%. Our results suggest population AE17 should also be considered as its optimal contribution is 17.5% of the seed optimal proportion. The selection of population AE17 to the optimal contribution seems to be the result of its main contribution to within-population diversity (a decrease of the genetic diversity by 6.96% when this population is removed from the analysis) but also to between-population diversity (a decrease of 0.68%). Regarding the Moroccan populations, the value of diversity explained among populations seems to be also underestimated by the RGUCs method. The genetic and geographic distance among populations make it difficult to encompass the majority of the genetic diversity with the preservation of the two populations proposed by the RGUCs method in the Moroccan area. It is imperative to consider the contribution to the genetic pool of the genetic distance among the populations within the genetically structured areas.

The aim of translocation proposals is to increase the survival of a given species (Commander *et al.*, 2018; Volis & Blecher, 2010). Even considering that it is impossible to know the final implications of a translocation due to the multiple interacting variables (Friar *et al.*, 2000), it is important to consider the possible risks of inbreeding and outbreeding depression (Barrett & Kohn, 1991). Inbreeding depression is common when only a few random individuals are used in translocations. The inevitable result is the decrease of fitness and genetic diversity of the populations (Young et al., 2005; Young & Clarke, 2000), and consequently, a loss in their ability to adapt to changing environmental conditions (Frankham, 2005). The proposal of obtaining optimal proportions of seeds from each population for translocation seems appropriate, not only because of the intended maximization of genetic diversity, but also because of the fact that collecting large quantities of seeds in natural populations may have a negative impact on local population dynamics (Broadhurst et al., 2008). Outbreeding depression may be an important risk in species that inhabit ecologically or geographically different areas (Fenster & Galloway, 2000). Exposure of individuals to new environments and the phenotypic changes that this generates can decrease the survival and reproductive success of individuals and, therefore, the success of translocation. (Lema & Nevitt, 2006). As shown by Bobo-Pinilla et al. (2018), this species exhibits a genetic pattern of three geographic areas (Iberian Peninsula, Morocco, and Canary Islands). As it is considered that the seeds from the same area will not affect the genetic pattern, the transference of plant material within these areas reduces the risk of outbreeding (Kaulfuß & Reisch, 2017). The phenotypic plasticity that can provide the adaptive traits of the species is generally unknown, and therefore conservation biologists still rely on quantitative genetic approaches to study phenotypic variation of endangered species and design efficient management programs (Noel et al., 2007). Following the predicted optimal contributions of the populations to global diversity in each area, a complete seed bank should be created and the populations most genetically impoverished (AE1, AE2, and AE6 from the Iberian Peninsula; AE7 and AE10-14 from Morocco; and AE15 from the Canary Islands) should be reinforced with the population proportions calculated for each area.

The future potential habitat of Astragalus edulis

Species distribution models are a recommended conservation tool that allow the proposal of areas where to create new populations of endangered species that are suffering under the climate change scenario (IUCN/ SSC, 2013). Despite this, their use for such an inference has been limited to date (Keppel et al., 2012). The impact of climate change on biodiversity will depend on the ability of plant species to migrate to new sites with suitable habitat conditions (Normand et al., 2011). As the seeds of the species lack of evident adaptations for dispersal (Peñas, 2004), the geographic distance between present and future suitable areas for A. edulis (Figure 3) makes it unlikely that their seeds colonize the areas found to have optimal conditions for them in the future. This mismatch between the high rate of change in climatic conditions in the habitat of the species and the ability of species to follow these changes may affect species survival (Malcolm et al., 2002).

The model indicates that the desertification process favors the optimal conditions for the species in areas

with no frost influence, with optimal conditions decreasing in inland areas and shifting directionally to the North (Figure 3B). Notably, these areas are also highly affected by human pressure, which can become a major drawback when proposing specific areas for translocation. Moreover, the optimal areas in the future scenario are located mainly outside the actual areas for the species. This implies a significant challenge, as the translocations outside the actual range of the species could have unknown results (Seddon et al., 2015). Shared suitable areas between present and future scenarios (Figure 3C) could be used to introduce plants corresponding with the closest genetic pattern (i.e., Iberian Peninsula, Morocco or Canary Islands). However, further research about fine scale distribution models is needed to find the specific sites, with adequate edaphic characteristics of the habitats, where to create the new populations.

Conclusion

We propose an approach that integrates genetic and geographic data to be used in ex situ and in situ complementary conservation measures as a tool to design efficient solutions for the conservation of endangered plant species. Applied to Astragalus edulis, we used genetic diversity data to propose an optimal proportion of seeds from the populations within the geographic areas in order to maximize the genetic pool contributing to storage the highest variability. This will strengthen the viability of the reinforcement planning of the most degraded populations of the species. Also, we detected the populations from the three geographic areas studied (Iberian Peninsula, Morocco and Canary Islands) that should be reinforced. Finally, we inferred, using the species distribution model, which areas will present optimal conditions for the survival of A. edulis in the future. A general pattern of range shifts towards coastal and northern areas have been detected, so further research is needed in order to find fine scale edaphic suitable sites to propose the creation of new populations given the decrease of suitable areas for the species in the current distribution range. Finally, in order to have a way to check the success of the translocation proposal, further analyses regarding the genetic diversity of the created and reinforced populations must be made and a monitoring of the population dynamics must be carried out. This study should help to propose genetically diverse and spatially explicit translocations.

Acknowledgements

This work has been financed by the Spanish Ministerio de Ciencia e Innovación through the projects CGL2012-32574 and REN2003-09427, as well as by the Andalusian Consejería de Innovación, Ciencia y Tecnología through the project P05-RNM-01067. A.C. was funded by Agencia Estatal de Investigación (AEI) (CGL2016-75904-C2-1-P), Xunta de Galicia (ED431C 2016-037)

and Fondos Feder: "Unha maneira de facer Europa". The funders had no role in study design, data collection and analysis, decision to publish or preparation of the manuscript. We are also grateful to M.M. Martínez-Ortega for her work in all the projects.

References

- Akeroyd, J. & Wyse Jackson, P. 1995. A handbook for botanical gardens on the reintroduction of plants to the wild. Botanic Gardens Conservation International (BGCI), Richmond.
- Armstrong, D.P. & Seddon, P.J. 2008. Directions in reintroduction biology. Trends Ecol. Evol. 23(1): 20– 25. doi:10.1016/j.tree.2007.10.003
- Bajomi, B., Pullin, A.S., Stewart, G.B., & Takács-Sánta, A. 2010. Bias and dispersal in the animal reintroduction literature. ORYX 44(3): 358–365. doi: 10.1017/ S0030605310000281
- Barrett, S.C.H. & Kohn, J.R. 1991. Genetic and evolutionary consequences of small population sizes in plants: Implications for conservation. In: Falk, D.A., Holsinger, K.A. (Eds.). Genetics and conservation of rare plants. Pp. 3–30. Oxford University Press, New York.
- Bobo-Pinilla, J., Peñas, J., López-González, N., Mediavilla, S., & Martínez-Ortega, M.M. 2018. Phylogeography of an endangered disjunct herb: Long-distance dispersal, refugia and colonization routes. AoB Plants 10(5), ply047. doi: 10.1093/aobpla/ply047
- Broadhurst, L.M., Lowe, A., Coates, D.J., Cunningham, S.A., McDonald, M., Vesk, P.A., & Yates, C. 2008. Seed supply for broadscale restoration: maximizing evolutionary potential. Evol. Appl. 1(4): 587–597. doi: 10.1111/j.1752-4571.2008.00045.x
- Caujapé-Castells, J. & Pedrola-Monfort, J. 2004. Designing ex-situ conservation strategies through the assessment of neutral genetic markers: Application to the endangered Androcymbium gramineum. Conserv. Genet. 5(2): 131–144. doi: 10.1023/B:COGE.0000029997.59502.88
- Christmas, M.J., Breed, M.F. & Lowe, A.J. 2016. Constraints to and conservation implications for climate change adaptation in plants. Conserv. Genet. 17(2): 305–320. doi: 10.1007/s10592-015-0782-5
- Commander, L., Coates, D., Broadhurst, L., Offord, C.A., Makinson, R.O. & Matthes, M. (Eds.). 2018. Guidelines for the translocation of threatened plants in Australia. Australian Network for Plant Conservation, Canberra.
- Conrad, O., Bechtel, B., Bock, M., Dietrich, H., Fischer, E., Gerlitz, L., Wehberg, J., Wichmann, V., & Böhner, J. 2015. System for Automated Geoscientific Analyses (SAGA) v. 2.1.4. Geosci. Model Dev. 8: 1991–2007. doi: 10.5194/gmd-8-1991-2015
- Dawson, T.P., Jackson, S.T., House, J.I., Prentice, I.C., & Mace, G.M. 2011. Beyond predictions: Biodiversity conservation in a changing climate. Science 332(6025): 53–58.

- Elith, J. & Leathwick, J.R. 2009. Species distribution models: ecological explanation and prediction across space and time. Annu. Rev. Ecol. Evol. Syst. 40(1): 677–697.
- Fay, M.F. & Krauss, S.L. 2003. Orchid conservation genetic in the molecular age. In: Dixon, K.W., Kell, S.P., Barrett, R.L. & Cribb, P.J. (Eds.). Orchid Conservation. Pp. 91–112. Natural History Publications, Kota Kinabalu, Sabah.
- Fenster, C.B. & Galloway, L.F. 2000. Inbreeding and outbreeding depression in natural populations of Chamaecrista fasciculata (Fabaceae). Conserv. Biol. 14(5): 1406–1412. doi: 10.1046/j.1523-1739.2000.99234.x
- Fenu, G., Bacchetta, G., Charalambos, S.C., Fournaraki, C., Giusso del Galdo, G. Pietro, Gotsiou, P., Kyratzis, A., Piazza, C., Vicens, M., Pinna, M.S., & de Montmollin, B. 2019. An early evaluation of translocation actions for endangered plant species on Mediterranean islands. Plant Divers. 41(2): 94–104. doi: 10.1016/j. pld.2019.03.001
- Frankham, R. 2005. Genetics and extinction. Biol. Conserv. 126(2): 131–140. doi: 10.1016/j.biocon.2005.05.002
- Frankham, R., Ballou, J.D., Briscoe, D.A., & McInnes, K.H. 2002. Introduction to Conservation Genetics. Cambridge University Press, Cambridge. doi: 10.1017/ CBO9780511808999
- Friar, E.A., Ladoux, T., Roalson, E.H., & Robichaux, R.H. 2000. Microsatellite analysis of a population crash and bottleneck in the Mauna Kea silversword, Argyroxiphium sandwicense ssp. sandwicense (Asteraceae), and its implications for reintroduction. Mol. Ecol. 9(12): 2027–2034. doi: 10.1046/j.1365-294X.2000.01111.x
- Godefroid, S., Piazza, C., Rossi, G., Buord, S., Stevens, A.D., Aguraiuja, R., Cowell, C., Weekley C.W., Vogg, G., Iriondo, J.M., Johnson, I., Dixon, B., Gordon, D., Magnanon, S., Valentin, B., Bjureke, K., Koopman, R., Vicens, M., Virevaire, M. & Vanderborght, T. 2011. How successful are plant species reintroductions? Biol. Conserv. 144(2): 672–682. doi: 10.1161/ STROKEAHA.110.606277
- Guisan, A., Tingley, R., Baumgartner, J.B., Naujokaitis-Lewis, I., Sutcliffe, P.R., Tulloch, A.I.T., Regan, T.J., Brotons, L., Mcdonald-Madden, E., Mantyka-Pringle, C., Martin, T.G., Rhodes, J.R., Maggini, R., Setterfield, S.A., Elith, J., Schwartz, M.W., Wintle, B.A., Broennimann, O., Austin, M., Ferrier, S., Kearney, M.R., Possingham, H.P., & Buckley, Y.M. 2013. Predicting species distributions for conservation decisions. Ecol. Lett. 16(12): 1424–1435.
- Hijmans, R.J., Cameron, S.E., Parra, J.L., Jones, P.G., & Jarvis, A. 2005. Very high resolution interpolated climate surfaces for global land areas. Int. J. Climatol. 25(15): 1965–1978.
- Kaulfuß, F. & Reisch, C. 2017. Reintroduction of the endangered and endemic plant species Cochlearia bavarica—Implications from conservation genetics. Ecol. Evol. 7(24): 11100–11112. doi: 10.1002/ ece3.3596

- Keppel, G., Van Niel, K.P., Wardell-Johnson, G.W., Yates, C.J., Byrne, M., Mucina, L., Schut, A.G.T., Hopper, S.D., & Franklin, S.E. 2012. Refugia: Identifying and understanding safe havens for biodiversity under climate change. Glob. Ecol. Biogeogr. 21(4): 393–404. doi: 10.1111/j.1466-8238.2011.00686.x
- Leadley, P., Pereira, H.M., Alkemade, R., Fernández-Manjarrés, J.F., Proença, V., Scharlemann, J.P.W. & Walpole, M.J. 2010. Biodiversity scenarios: Projections of 21st Century change in biodiversity and associated ecosystem services. Secretariat of the Convention on Biological Diversity, Montreal.
- Lema, S.C. & Nevitt, G.A. 2006. Testing an ecophysiological mechanism of morphological plasticity in pupfish and its relevance to conservation efforts for endangered Devils Hole pupfish. J. Exp. Biol. 209(18): 3499–3509.
- López-Cortegano, E., Pérez-Figueroa, A., & Caballero, A. 2019. metapop2: Re-implementation of software for the analysis and management of subdivided populations using gene and allelic diversity. Mol. Ecol. Resour. 19(4): 1095–1100. doi: 10.1111/1755-0998.13015
- Maes, D., Vanreusel, W., Talloen, W., & Van Dyck, H. 2004. Functional conservation units for the endangered Alcon Blue butterfly Maculinea alcon in Belgium (Lepidoptera: Lycaenidae). Biol. Conserv. 120(2): 229–241. doi: 10.1016/j.biocon.2004.02.018
- Malcolm, J.R., Markham, A., Neilson, R.P., & Garaci, M. 2002. Estimated migration rates under scenarios of global climate change. J. Biogeogr. 29(7): 835–849. doi: 10.1046/j.1365-2699.2002.00702.x
- Moritz, C. 1994. Defining Evolutionarily Significant Units. Tree Genet. Genomes 9: 373–375.
- Nathan, R. 2006. Long-distance dispersal of plants. Science 313(5788): 786–788. doi: 10.1126/science.1124975
- Nei, M. 1987. Molecular evolutionary genetics. Columbia University Press, New York.
- Noel, F., Machon, N., & Porcher, E. 2007. No genetic diversity at molecular markers and strong phenotypic plasticity in populations of Ranunculus nodiflorus, an endangered plant species in France. Ann. Bot. 99(6): 1203–1212.
- Nogués-Bravo, D., Rodríguez-Sánchez, F., Orsini, L., de Boer, E., Jansson, R., Morlon, H., Fordham, D.A., & Jackson, S.T. 2018. Cracking the code of biodiversity responses to past climate change. Trends Ecol. Evol. 33(10): 765–776. doi: 10.1016/j.tree.2018.07.005
- Normand, S., Ricklefs, R.E., Skov, F., Bladt, J., Tackenberg, O., & Svenning, J.C. 2011. Postglacial migration supplements climate in determining plant species ranges in Europe. Proc. R. Soc. B. 278(1725): 3644–3653. doi: 10.1098/rspb.2010.2769
- Pearse, D.E. & Crandall, K.A. 2004. Beyond FST: Analysis of population genetic data for conservation. Conserv. Genet. 5(5): 585–602. doi: 10.1007/s10592-003-1863-4
- Peñas, J. 2004. Astragalus edulis Bunge. In: Bañares, Á., Blanca, G., Güemes, J., Moreno, J.C. & Ortiz, S. (Eds.). Atlas y libro rojo de la flora vascular amenazada de España. Pp. 614–615. Dirección General de Conservación de La Naturaleza. R. Jard. Bot., CSIC, Madrid

- Peñas, J., Barrios, S., Bobo-Pinilla, J., Lorite, J., & Martínez-Ortega, M.M. 2016. Designing conservation strategies to preserve the genetic diversity of Astragalus edulis Bunge, an endangered species from western Mediterranean region. PeerJ 4(1): e1474. doi: 10.7717/peerj.1474
- Pérez-Collazos, E., Segarra-Moragues, J.G. & Catalán, P. 2008. Two approaches for the selection of Relevant Genetic Units for Conservation in the narrow European endemic steppe plant Boleum asperum (Brassicaceae). Biol. J. Linn. Soc. 94(2): 341–354. doi: 10.1111/j.1095-8312.2008.00961.x
- Petit, R.J., El Mousadik, A., & Pons, O. 1998. Identifying populations for conservation on the basis of genetic markers. Conserv. Biol. 12(4): 844–855. doi: 10.1111/j.1523-1739.1998.96489.x
- Riddle, B.R. & Hafner, D.J. 1999. Species as units of analysis in ecology and biogeography: time to take the blinders off. Glob. Ecol. Biogeogr. 8(6): 433–441. doi: 10.1046/j.1365-2699.1999.00170.x
- Ryder, O.A. 1986. Species conservation and systematics: the dilemma of subspecies. Trends Ecol. Evol. 1(1): 9–10. doi: 10.1016/0169-5347(86)90059-5
- Seddon, A.W., Macias-Fauria, M. & Willis, K.J. 2015. Climate and abrupt vegetation change in Northern Europe since the last deglaciation. Holocene 25(1): 25–36. doi: 10.1177/0959683614556383
- Segarra-Moragues, J.G. & Catalán, P. 2010. The fewer and the better: Prioritization of populations for conservation under limited resources, a genetic study with Borderea pyrenaica (Dioscoreaceae) in the Pyrenean National Park. Genetica 138(3): 363–376.
- Thompson, J.D. 1991. Phenotypic plasticity as a component of evolutionary change. Trends Ecol. Evol. 6(8): 246-249.
- Toro, M.A. & Caballero, A. 2005. Characterization and conservation of genetic diversity in subdivided populations. Philos. Trans. R. Soc. B Biol. Sci. 360(1459): 1367–1378. doi: 10.1098/rstb.2005.1680
- Tremlová, K. & Münzbergová, Z. 2007. Importance of species traits for species distribution in fragmented landscapes. Ecology 88(4): 965–977. doi: 10.1890/06-0924

- Valladares, F., Sanchez-Gomez, D. & Zavala, M.A. 2006. Quantitative estimation of phenotypic plasticity: Bridging the gap between the evolutionary concept and its ecological applications. J. Ecol. 94: 1103–1116.
- Vekemans, X., Beauwens, T., Lemaire, M., & Roldán-Ruiz, I. 2002. Data from amplified fragment length polymorphism (AFLP) markers show indication of size homoplasy and of a relationship between degree of homoplasy and fragment size. Mol. Ecol. 11(1): 139–151. doi: 10.1046/j.0962-1083.2001.01415.x
- Volis, S. 2019. Conservation-oriented restoration a two for one method to restore both threatened species and their habitats. Plant Divers. 41(2): 50–58. doi: 10.1016/j.pld.2019.01.002
- Volis, S. & Blecher, M. 2010. Quasi in situ: a bridge between ex situ and in situ conservation of plants. Biodivers. Conserv. 19(9): 2441–2454. doi: 10.1007/ s10531-010-9849-2
- Young, A.G. & Clarke, G.M. 2000. Genetics, demography and viability of fragmented populations. Cambridge Univ. Press, New York. doi: 10.1017/cbo9780511623448
- Young, T.P., Petersen, D.A., & Clary, J.J. 2005. The ecology of restoration: Historical links, emerging issues and unexplored realms. Ecol. Lett. 8(6): 662– 673. doi: 10.1111/j.1461-0248.2005.00764.x

Websites

- IUCN. 1998. IUCN Guidelines for Re-introductions. Gland & Cambridge. https://www.iucn.org/es/content/iucnguidelines-re-introductions
- IUCN/SSC. 2013. Guidelines for reintroductions and other conservation translocations. Species Survival Commission, Gland. Available at https://www.iucn. org/content/guidelines-reintroductions-and-otherconservation-translocations
- QGIS-Development-Team. 2017. QGIS un Sistema de Información Geográfica libre y de Código Abierto. htpps://qgis.org/es/site