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### Research report

# Consistent rewarding or aversive effects of the electrical stimulation of the lateral parabrachial complex

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#### Abstract

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Electrical stimulation of the external lateral parabrachial subnucleus (LPBe) may induce rewarding or aversive behaviors in animals subjected to two different learning discrimination tasks. Statistical analysis found no significant differences between the group receiving electrical stimulation of the brain and the non-stimulated control group. However, rewarding or aversive behaviors were consistent and positively correlated between the two discrimination tasks in the stimulated group.

Thus, these tests differed in the gustatory stimuli used, in the right/left position of stimulation-associated/non-associated flavors, and in the cage in which experiments were performed. This behavioral consistency and corresponding correlation were not observed in the non-stimulated control group. These results suggest the existence of aversive and reward systems that are differentiated but anatomically very close. Therefore, the activation of aversive or rewarding systems may depend on the precise location of the electrode implanted in the LPBe of each animal. © 2008 Elsevier B.V. All rights reserved.

16 Keywords: Electrical brain stimulation; Parabrachial nucleus; Reward; Taste preference

#### 1. Introduction

The external lateral parabrachial subnucleus (LPBe) is at the ventral lateral end of the parabrachial complex and has been implicated in the processing of visceral and gustatory information [1,2]. The LPBe receives gustatory information through the rostral nucleus of the solitary tract (NST) [2–4], whereas it receives both vagal and visceral information [5] by means of its connections with the caudal NTS and area postrema (AP) [1,6,7].

The LPBe has been related to taste aversion learning 26 induced by aversive agents (e.g., abdominal irritants, body 27 rotation, or copper sulfate) or even by drugs of abuse (e.g., 28 cocaine, amphetamines, or morphine) [8–12], mostly when these 29 gustatory-visceral stimuli are contiguously presented [9,13]. 30 Furthermore, this LPBe subnucleus has also been related to 31 the spino(trigemino) pontoamygdaloid system and would be involved in the transmission of nociceptive information and in 33 the affective-emotional, autonomic, and visceral processing of 34 these negative events [14-24]. 35

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The LPBe may also participate in the processing of rewarding substances. Thus, the intraduodenal injection of glucose [25] or intragastric administration of lactose or sucrose, among others, generates C-Fos immunoreactivity in this area [11,12]. Moreover, lesions in this parabrachial subnucleus block taste preferences induced by the intragastric administration of rewarding food [26]. In this regard, some substances related to food intake and nutritional metabolism may exert their functions through this region, as in the case of cholecystokinin (CCK) [27,28], galanin, neuropeptide Y [29–31], and leptin [32]. The same has been found for certain drugs that increase the hedonic value of intake, e.g., benzodiazepines [33]; agents that reduce food intake, e.g., mercaptoacetate, 2.5-anhydro-D-mannitol, methyl palmoxirate [37–40,75].

Finally, it has been demonstrated that some drugs of abuse implicated in the reduction or increase of food intake [41–44], e.g., amphetamines [10] or opiates [10,27,35,45,46], may also be processed *via* the LPBe. In relation to the opiate system, it has been shown that the number of opiate receptors of this parabrachial area can be modified by food restriction [47] and, conversely, that drug manipulation of opiate receptors in this area can modulate intake [48,49].

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Taken together, the above data suggest that the LPBe nucleus may be related to both aversive and rewarding motivational 60 processes. For this reason, we expected that LPBe electrical 61 stimulation, an appropriate procedure for inducing prefer-62 ence/rewarding or aversive behaviors [50-52], might generate 63 these appetitive and/or aversive responses in a random but con-64 sistent manner. These results might also serve as a behavioral 65 criterion to identify the aversive or rewarding effects of LPBe 66 electrical stimulation, especially the latter (preferences), given 67 that lever-press self-stimulation has not been obtained with these 68 animals [53], or at least not as readily as in the lateral hypothala-69 mus, for example [54]. Therefore, in this study, it was decided to 70 use two different learning discriminative tasks by which the ani-71 mals could associate a stimulus with its rewarding or aversive 72 consequences [26,51,52,55–59]. Furthermore, the two flavors 73 presented, the right/left position of the stimulation-associated 74 flavor, and the cages used by each animal were modified in these 75 tests. According to our hypothesis, the animals would still prefer 76 or avoid, in each individual case, the stimulus associated with 77 intracerebral electrical stimulation, despite the modifications 78 introduced and possible interferences among tasks, although 79 global differences might not be found between stimulated and 80 non-stimulated groups. 81

#### 2. Materials and methods

#### 2.1. Subjects

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Thirty-one male Wistar rats (255-315 g each at time of surgery) from 84 the University of Granada animalarium were used in this study. They were 85 O2 randomly assigned to one of two groups: (a) ES-LPBe: intermittent (30' 86 on/30" off) (n = 10) or continuous (n = 12) electrical stimulation and (b) US-87 control: unstimulated control (n=9). In the former group, the total duration 88 of stimulation at each session was 15' in all cases. Since no significant dif-89 ferences were found between the two subgroups, the results were analyzed 90 91 together.

On arrival at the laboratory, animals were individually housed in  $30 \text{ cm} \times 15 \text{ cm} \times 30 \text{ cm}$  methacrylate cages that served as training chambers. 93 The room temperature was maintained between 21 and 24 °C, and light-dark periods lasted 12 h each, with lights on at 8:30 a.m. All experimental and test procedures were conducted during the light phase.

Subjects were allowed a 5-day adaptation period during which they remained 97 in their home cage with food and water available ad libitum. All behavioral 98 procedures and surgical techniques were conducted in agreement with animal 99 care guidelines established by Spanish Royal Law 23/1988. 100

Day 1

Strawberry L + St. (15 min)

Lemon L. + St. (15 min)

Lemon L + Non-St. (15 min)

Strawberry L + Non-St. (15 min)

#### Table 1

#### Q6 Behavioral procedure

Phase I

Phase I

Phase II

Random distribution

Group A 50% of animals

Group B 50% of animals

Group A 50% of animals

Group B 50% of animals

#### 2.2. Surgery

#### Under general anesthesia (sodium pentothal, 50 mg/kg., B. Braun Medical S.A. Barcelona, Spain), the ES-LPBe group was stereotaxically implanted (Stoelting Co. Stereotaxic 51600, USA) with a 00 stainless steel electrode aimed at the LPBe {Coordinates: AP = -0.16; V = 3; L = $\pm 2.5$ ; [76]}. After surgery, subjects were returned to their home cages where they stayed for at least 7 days of recovery with water and food ad libitum.

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Animals from the US-control group had similar periods of adaptation and recovery but did not undergo surgical intervention.

#### 2.3. Apparatus

As mentioned above, experimental procedures were conducted in the same methacrylate chambers that served as home cages. The front part of each cage had two 1.6-cm holes, each at the same distance from the center and sides and at the same height from the floor. Through those orifices, the animal had access to spouts attached to graduated burettes in which the flavors were offered.

Electrical stimulation was delivered using a LI12100 stimulator (Letica, Barcelona, Spain). At a current selected to be below the threshold for producing undesired behavioral effects [60], 66.6 Hz, 0.1 ms, 57-115 µA, cathodal rectangular pulses were applied to the LPBe. The stimulation process was moni- Q3 tored with a DM63 oscilloscope (Textronic Ltd., London, U.K.), which allowed constant visualization of the electrical pulses administered to animals during experimental sessions.

#### 2.4. Behavioral procedure

#### 2.4.1. Pretraining

During two pretraining sessions, all animals were water deprived for 23 h 50 min and allowed to drink tap water for 10 min from graduated burettes. The position of the burettes was alternated across sessions to avoid development of positional preferences. Once the water was withdrawn, animals had access to 15 g of food.

The experiment began after the 2-day pretraining period and comprised two phases separated by a 14-day interval. In each phase, animals underwent a discriminative learning task. The gustatory stimuli used were strawberry 'S' and coconut 'C' for phase I and lemon 'L' and vanilla 'V' for phase II (0.5% diluted in water, McCormick & Co. Inc. San Francisco, CA), which were always presented in the same right/left position.

In each phase, four learning sessions were performed in which the two different gustatory stimuli were offered on alternate days. For half of the animals, intake of one flavor (for 7 min) was immediately followed by 15 min of electrical stimulation of the LPBe, whereas intake of the other flavor was not followed by stimulation. For the remaining animals, the opposite process was carried out (for details, see Table 1). On day 5, a two-bottle free-choice test was conducted by placing both burettes in the cage simultaneously. Animals were allowed to freely drink the flavored solutions for 7 min and the total amount ingested was recorded.

Day 4

=2nd day

=2nd day

=2nd day

=2nd day

Test

Strawberry L, coconut R (7 min)

Strawberry L, coconut R (7 min)

Lemon L, vanilla R (7 min)

Lemon L, vanilla R (7 min)

Day 3

=1st day

=1st day

\*0 Day

\*# Day

Diagram of the balanced experimental conditions used in the two phases of this experiment. Strawberry and coconut were used as gustatory stimuli in the first phase and vanilla and lemon in the second phase. The unstimulated control animals (US-control) were also assigned one flavor with sham stimulation and another with non-"stimulation", although control animals never received electrical stimulation.

Vanilla R + St. (15 min)

Coconut R+Non-St. (15 min)

Vanilla R + Non-St. (15 min)

Coconut R + St. (15 min)

Day 2

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Table 2								
Design of position	ns occupied by ex	perimental and con	trol animals					
Phase 1								
Subject	1	2	3	4	5	6	7	8
Position	Cage 1		Cage 2		Cage 3		Cage 4	
Phase II								
Subject	4	3	8	7	2	1	6	5
Position	Cage 1		Cage 2		Cage 3		Cage 4	

Procedure for the distribution of the animals in different cages during phases I and II of the experiment. This procedure (amplified) was used for the 18 animals in the ES-LPBe group and the 9 in the US-control group.

In the case of the unstimulated control group (US-control), intake of the two gustatory stimuli was never followed by brain electrical stimulation.

In the interval between phases, animals received food and water ad libitum. At the start of the second phase, conditions of the new discrimination test were modified with regard to the gustatory stimuli used, the right/left position of the electrical stimulation-associated stimulus, and the cage in which experiments were conducted (Table 2).

#### 2.5. Histology

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After conclusion of the experiment, all animals were deeply anesthetized 153 with an overdose of sodium pentothal and intracardially perfused with isotonic 154 saline and 4% paraformaldehyde. Electrolytic lesions (0.5 mA of cathodic cur-155 rent for 5 s) were made to verify placement of electrodes in the LPBe. Their brains 156 were removed, stored in paraformaldehyde, and laminated in 50-µ sections. 157 Sections were mounted, stained with cresyl violet, and photographed (VMZ-4F 158 stereoscopic magnifying glass and PM-6 camera, Olympus, Tokyo, Japan) (see 159 Figs. 1 and 2). 160

#### 2.6. Data analysis 161

Statistica 6.0 software (Statsoft Inc., OK) was used for the statistical analysis. 162 A mixed bifactorial (group × flavor) ANOVA was used to analyze differ-163 ences between stimulated and control groups. 164

To demonstrate the intra-subject consistency of data, comparisons were per-165 formed using correlation coefficients of the differential data (difference between 166



Fig. 1. Histological preparation stained with cresyl violet, showing the localization (as small lesion) of the area occupied by the end of the electrode, representative of observations in the animals in this experiment.



Interaural .0.16mm



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Fig. 2. Map of the localization of electrode tip in some experimental group animals (diamond, animal 1; inverted triangle, animal 2; four-point star, animal 6; five-point star, animal 8; square, animal 13; rounded square, animal 16; triangle, animal 17).

intake of electrical LPBe stimulation-associated flavor [Flavor + St.] and intake of the flavor not associated with this stimulation [Flavor+Non-St.]) obtained for each animal in phases I and II tests. In the US-control group, one of the flavors was randomly selected before each learning phase to be sham-associated with the electrical LPBe stimulation (animals in this group never received this stimulation).

Classification and selection criteria can be developed from these data according to the behavior of these animals. Thus, animals were grouped into: those with a >50% consumption of the stimulation-associated flavor in the two tests, designated "positive" group, those with <30% consumption of the stimulationassociated flavor in the two tests, designated "negative" group; and those with 30-50% consumption of this flavor in one or both phases, designated "neutral" group

#### 3. Results

Three animals in the ES-LPBe group were excluded from the statistical analysis because the implant became detached during 167

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the behavioral procedure. Another subject of this same group was excluded for showing circling behavior.

There was wide intersubject variability in the intake of one or other flavor (Flavor + St. or Flavor + Non-St.) by animals in both groups (ES-LPBe and US-control), and the mixed bifactorial (group × flavor) ANOVA found no differences in phase 1 between groups [ $F(1,25)=0.2230 \ p<0.6408$ ] in either flavor intake [ $F(1,25)=0.2490 \ p<0.6221$ ] or interaction of the two factors [ $F(1,25)=0.2724 \ p<0.6063$ ). Similar results were found for phase II [group:  $F(1,25)=1.0083 \ p<0.3249$ ; flavor:  $F(1,25)=2.0877 \ p<0.1609$ ; interaction:  $F(1,25)=0.0010 \ p<0.9748$ ] (Fig. 3).

However, the comparison using Pearson correlation coefficient data for each animal in phases I and II showed a significant correlation (r = 0.70508;  $p > 0.001^{**}$ ) for the intake of stimulation-associated flavor values in both phases by the ES-LPBe group (Fig. 4). This was not the case for the UScontrol group, in which the correlation was not significant (r = -0.07176; p < 0.8544) (Fig. 5). When the criteria proposed above were applied to the 18 animals in the ES-LPBe group, 6 were assigned to the "positive" group (preference for stimulation-associated flavor) [mean current intensity of 87.1 μA], 7 to the "negative" group (aversion to stimulationassociated flavor) [mean intensity of  $81.5 \,\mu$ A], and 5 to the "neutral" group [mean =  $100 \mu$ A]. Hence, whereas 72% of animals in the stimulated group were consistent in their behavior, only two members of the "US-control" group showed a repeated preference for one of the gustatory stimuli presented, while the



Fig. 3. Mean intake (in cc) by ES-LPBe (above) and US-control (below) of the flavor associated with electrical stimulation of the LPBe and the flavor not associated with this stimulation (Flavor + St., Flavor + Non-St.) during phases I and II of the experimental procedure in this experimental series.



Fig. 4. Correlation matrix for the data of each ES-LPBE subject in phases I and II of the experiment (r = 0.7050,  $p > 0.001^{**}$ ). The area between the two curves represents the estimated surface area predicted to contain these data points from the performance of each subject in the two phases of the experiment, with a confidence interval of 95%.



Fig. 5. Correlation matrix for the data of each subject of the US-control group in phases I and II of the experiment (r = -0.0717, p < 0.8544).

remaining 7 [77.7%] met the criterion established for the neutral group. Q4

#### 4. Discussion

The results of this experiment show that LPBe stimulation may induce preferences or aversions towards associated stimuli in discrimination learning tasks. Analysis of the preferences shown for the flavor associated with the electrical brain stimulation in each task showed no significant results. However, a significant correlation was found between data obtained in the two tests by the stimulated animals [ES-LPBe] but not by the control animals [US-control]. This finding suggests that most animals in the stimulated group developed a preference for either the electrical stimulation-associated flavor or for the other one - and only a minority showed alternating behavior. Hence, this effect was consistent in the majority of the electrically stimulated animals, despite modifications introduced into the two discrimination learning tasks with regard to the gustatory stimuli presented, the left/right localization of the stimulationassociated flavor, and the experimental cage used, with the 213

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consequent egocentric and allocentric re-organization that these
tests might have required. Therefore, these data appear to indicate that the effect is consistent within an individual but not
among individuals.

The lateral end of the parabrachial nucleus, which includes 234 the LPBe, receives visceral information from the caudal NTS 235 and area postrema [1,2,6] and gustatory information from its 236 connections with the rostral NTS [2,7]. This sensory informa-237 tion can be rewarding or aversive. Thus, it has been reported that 238 the LPBe appears to participate in the development of learned 239 preferences and aversions [9,26]. In this context, electrical stim-240 ulation of the LPBe can act as an adequate stimulus to induce one 241 or other biological process (positive or negative reinforcement), 242 as also found in other brain regions [50–52,61]. 243

Previously published data have shown that a single anatomi-244 cal structure can be the substrate for both appetitive and aversive 245 motivational processes [62-65]: The same can be observed with 246 some substances (e.g., corticotropin-releasing factor or κ ago-247 nists), whose rewarding or aversive effects may depend on the 248 dose, the experimental situation, or the anatomic localization 249 of the neurochemical systems involved [66,67]. In the present 250 study, however, while the stimulated anatomical area appeared 251 to be critical, no differences were found in preference/aversion 252 behavior as a function of the electrical current parameters used. 253

In other words, it appears possible that the electrical stim-254 ulation of the LPBe could have affected positive or negative 255 cells according to the precise localization of the electrode. This 256 would modify the quality of the associated stimulus and would 257 mean that the stimulus, initially neutral, acquires appetitive or 258 aversive motivational properties. This interpretation is compat-259 ible with the identification in this LPBe area of cells that react 260 to the hedonic properties of different gustatory stimuli [4], and 261 of mutually inhibitory interactions between activity patterns of 262 hedonic-positive and hedonic-negative cells [2,7,68,69].

Furthermore, various authors have detected C-Fos immunoreactivity in the LPBe after the intragastric administration of nutrients and of chemical substances that generate learned preferences (e.g., glucose, sucrose, lactose, maltose, polycose) [12,25] or aversions (e.g., cocaine, morphine, methamphetamine, hypertonic saline, LiCl, copper sulfate) [10,11].

The present observation of animals that found electrical 270 stimulation of the LPBe to be aversive is also consistent 271 with reports by other authors that the administration of some 272 immune system-activating toxic agents (bacterial lipopolysac-273 charide) can produce immunoreactivity in the external part of 274 the LPBe [70]. This aversive effect can also be explained as 275 a consequence of the activation of an important relay of the 276 spino-parabrachial pathway in the LPBe, whose participation 277 in the processing of noxious information is well documented 278 [14-24].279

Finally, various authors have demonstrated that the behavioral effect of opiates may depend on the type of receptor on which they act. Thus, agonists of  $\mu$  and  $\delta$  receptors usually generate reinforcement, whereas substances that act on  $\kappa$  receptors do not favor its self-administration and may also induce aversive behaviors [71,72]. Some of these receptor types, i.e.,  $\mu$  and  $\kappa$ , have been identified in the LPBe [47]. It is likely that the electrical stimulation could have produced an activation of some or other receptors, generating positive or negative effects or even both simultaneously, which would explain the behavior of the "neutral" animals. Indeed, this explanation is compatible with data reported by Moufid-Bellancourt et al. [71], who achieved both rewarding and aversive effects from the intraparabrachial infusion of  $\mu$  and  $\kappa$  agonists, respectively. Moreover, it cannot be ruled out that the electrical stimulation of the LPBe might also have activated a motivational system associated with deficit/satiation states [64] or brain reward.

To summarize, the present results show that electrical stimulation of the LPBe can generate, in different animals, consistent preferences or aversions to gustatory stimuli with which it is associated. These results may possibly depend on slight variations in the position of the end of the electrode, differentially activating some of the motivational subsystems present in this region. These different reward/aversion mechanisms could be anatomically very close together, allowing the establishment of learned associations between a stimulus and its positive or negative consequences, analogous to observations in humans [64,73,74].

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#### Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.bbr.2008.02.036.

#### References

- De Lacalle S, Saper CB. Calcitonin gene-related peptide-like immunoreactivity marks putative visceral sensory pathways in human brain. Neuroscience 2000;100(1):115–30.
- [2] Karimnamazi H, Travers SP, Travers JB. Oral and gastric input to the parabrachial nucleus of the rat. Brain Res 2002;957(2):193–206.
- [3] Hermann GE, Rogers RC. Convergence of vagal and gustatory afferent input within the parabrachial nucleus of the rat. J Auton Nerv Syst 1985;13(1):1–17.
- [4] Yamamoto T, Shimura T, Sakai N, Ozaki N. Representation of hedonics and quality of taste stimuli in the parabrachial nucleus of the rat. Physiol Behav 1994;56(6):1197–202.
- [5] Yuan CS, Barber WD. Parabrachial nucleus: neuronal evoked responses to gastric vagal and greater splanchnic nerve stimulation. Brain Res Bull 1991;27(6):797–803.
- [6] Papas S, Ferguson AV. Electrophysiological characterization of reciprocal connections between the parabrachial nucleus and the area postrema in the rat. Brain Res Bull 1990;24(4):577–82.
- [7] Halsell CB, Travers SP. Anterior and posterior oral cavity responsive neurons are differentially distributed among parabrachial subnuclei in rat. J Neurophysiol 1997;78(2):920–38.
- [8] Yamamoto T, Shimura T, Sako N, Azuma S, Bai WZ, Wakisaka S. C-Fos expression in the rat brain after intraperitoneal injection of lithium chloride. Neuroreport 1992;3(12):1049–52.

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#### M.J. Simon et al. / Behavioural Brain Research xxx (2008) xxx-xxx

- [9] Mediavilla C, Molina F, Puerto A. The role of the lateral parabrachial nuclei in concurrent and sequential taste aversion learning in rats. Exp Brain Res 2000;134(4):497–505.
- [10] Sakai N, Yamamoto T. Conditioned taste aversion and C-Fos expression in the rat brainstem after administration of various USs. Neuroreport 1997;8(9–10):2215–20.
- [11] Yamamoto T, Sawa K. C-Fos- like immunoreactivity in the brainstem following gastric loads of various chemical solutions in rats. Brain Res 2000;866(1–2):135–43.
- [12] Yamamoto T, Sawa K. Comparison of C-Fos- like immunoreactivity in the brainstem following intraoral and intragastric infusions of chemical solutions in rats. Brain Res 2000;866(1–2):144–51.
- [13] Mediavilla C, Molina F, Puerto A. Concurrent conditioned taste aversion: a learning mechanism based on rapid neural versus flexible humoral processing of visceral noxious substances. Neurosci Biobehav Rev 2005;29(7):1107–18.
- [14] Bernard JF, Carroué J, Besson JM. Efferent projections from the external parabrachial area to the forebrain: a *Phaseolus vulgaris* leucoagglutinin study in the rat. Neurosci Lett 1991;122(2):257–60.
- [15] Bernard JF, Huang GF, Besson JM. The parabrachial area: electrophysiological evidence for an involvement in visceral nociceptive processes. J Neurophysiol 1994;71(5):1646–60.
- [16] Bernard JF, Dallel R, Raboisson P, Villanueva L, Le Bars D. Organization of the efferent projections from the spinal cervical enlargement to the parabrachial area and periaqueductal gray: a PHA-L study in the rat. J Comp Neurol 1995;353(4):480–505.
- [17] Bester H, Menendez L, Besson JM, Bernard JF. Spino-(trigemino-) parabrachiohypothalamic pathway: electrophysiological evidence for an involvement in pain processes. J Neurophysiol 1995;73(2):568–85.
- [18] Bester H, Matsumoto N, Besson JM, Bernard JF. Further evidence for the involvement of the spinoparabrachial pathway in nociceptive processes: a C-Fos study in the rat. J Comp Neurol 1997;383(4):439–58.
- [19] Light AR, Sedivec MJ, Casale EJ, Jones SL. Physiological and morphological characteristics of spinal neurons projecting to the parabrachial region of the cat. Somatosens Mot Res 1993;10(3):309–25.
- [20] Saper CB. The spinoparabrachial pathway: shedding new light on an old path. J Comp Neurol 1995;353(4):477–9.
- [21] Jasmin L, Burkey AR, Card JP, Basbaum AI. Transneuronal labeling of a nociceptive pathway, the spino-(trigemino)–arabrachio-amigdaloid, in the rat. J Neurosci 1997;17(10):3751–65.
- [22] Craig AD, Dostrosvsky JO. Medulla to thalamus. In: Wall PD, Melzack R, editors. Textbook of Pain. 4th ed. New York: Churchill Livingstone; 1999. p. 183–214.
- [23] Wang LG, Li HM, Li JS. Formalin induced fos-like inmunoreactive neurons in the trigeminal spinal caudal subnucleus project to contralateral parabrachial nucleus in the rat. Brain Res 1994;649(1–2):62–70.
- [24] Buritova J, Besson JM, Bernard JM. Involvement of the spinoparabrachial pathway in inflamatory nociceptive processes: a C-Fos protein study in the awake rat. J Comp Neurol 1998;397(1):10–28.
- [25] Wang L, Cardin S, Martinez V, Tache I, Lloyd CK. Duodenal loading with glucose induces Fos expression in rat brain: selective blockade by devazepide. Am J Physiol 1999;277(3):R667–74.
- [26] Zafra MA, Simon MJ, Molina F, Puerto A. The role of the external lateral parabrachial subnucleus in flavor preferences induced by pre-digested food administered intragastrically. Brain Res 2002;950(1–2):155–64.
- [27] Li BH, Rowland NE. Effects of vagotomy on cholecystokinin- and dexfenfluramine-induced fos-like immunoreactivity in the rat brain. Brain Res Bull 1995;37(6):589–93.
- [28] Trifunovic R, Reilly S. Medial versus lateral parabrachial nucleus lesions in the rat: effects cholecystokinin- and D-fenfluramine-induced anorexia. Brain Res 2001;894(2):288–96.
- [29] Smith GP, Gibbs J, Kulkosky PJ. Relationships between brain-gut peptides and neurons in the control of food Intake. In: Hoebel BG, Novin D, editors. The Neural Basis of Feeding and Reward. Haer Institute for Electrophysiology Research; 1982. p. 149–66.
- [30] Petrov T, Jhamandas JH, Krukoff TL. Characterization of peptidergic efferents from the lateral parabrachial nucleus to identified neurons in the rat dorsal rafe nucleus. J Chem Neuroanat 1992;5(5):367–73.

- [31] Petrov T, Krukoff TL, Jhamandas JH. The hypothalamic paraventricular and lateral parabrachial nuclei receive collaterals from raphe nucleus neurons: a combined double retrograde and immunocytochemical study. J Comp Neurol 1992;318(1):18–26.
- [32] Elias CF, Kelly F, Lee CE, Ahima RS, Drucker DJ, Saper CB, et al. Chemical characterization of leptin-activated neurons in the rat brain. J Comp Neurol 2000;423(2):261–81.
- [33] Söderpalm AH, Berridge KC. The hedonic impact and intake of food are increased by midazolam microinjection in the parabrachial nucleus. Brain Res 2000;877(2):288–97.
- [34] Li BH, Spector AC, Rowland NE. Reversal of dexfenfluramine-induced anorexia and C-Fos/C-Jun expression by lesion in the lateral parabrachial nucleus. Brain Res 1994;640(1–2):255–67.
- [35] Li BH, Rowland NE. Dexfenfluramine induces fos-like immunoreactivity in discrete brain regions in rats. Brain Res Bull 1993;31(1–2):43–8.
- [36] Simansky KJ, Nicklous DM. Parabrachial infusion of D-fenfluramine reduces food intake blockade by the 5-HT1B antagonist SB-216641. Pharmacol Biochem Behav 2002;71(4):681–90.
- [37] Calingasan NY, Ritter S. Lateral Parabrachial subnucleus lesions abolish feeding induced by mercaptoacetate but not by 2-deoxy-D-glucose. Am J Physiol 1993;265(34):R1168–78.
- [38] Ritter S, Dinh TT, Friedman MI. Induction of Fos-like immunoreactivity (Fos-li) and stimulation of feeding by 2,5-anhydro-D-mannitol (2,5-AM) require the vagus nerve. Brain Res 1994;646(1):53–64.
- [39] Horn CC, Friedman MI. 2,5-Anhydro-D-mannitol induces Fos-like immunoreactivity in hindbrain and forebrain: relationship to eating behavior. Brain Res 1998;779(1–2):17–25.
- [40] Horn CC, Friedman MI. Methyl palmoxirate increases eating and brain Fos-like immunoreactivity in rats. Brain Res 1998;781(1–2):8–14.
- [41] Le Magnen J. Neurobiology of feeding and nutrition. San Diego: Academic Press; 1992.
- [42] Drewnowski A. Taste preferences and food intake. Annu Rev Nutr 1997;17:237–53.
- [43] Carr KD. Augmentation of drug reward by chronic food restriction: behavioral evidence and underlying mechanism. Physiol Behav 2002;76(3):353–64.
- [44] Carr KD. Chronic food restriction: enhancing effects on drug reward and striatal cell signaling. Physiol Behav 2007;91(5):459–72, doi:10.1016/j.physbeh.2006.09.021.
- [45] Ding YQ, Kaneko T, Nomura S, Mizuno N. Immunohistochemical localization of μ-opioid receptors in the central nervous system of the rat. J Comp Neurol 1996;367(3):375–402.
- [46] Chamberlin NL, Mansour A, Watson SJ, Saper CB. Localization of muopioid receptors on amygdaloid projection neurons in the parabrachial nucleus of the rat. Brain Res 1999;827(1–2):198–204.
- [47] Wolinsky TS, Carr KD, Hiller JM, Simon EJ. Chronic food restriction alters μ and κ opioid receptor binding in the parabrachial nucleus of the rat: a quantitative autoradiographic study. Brain Res 1996;706(2):333–6.
- [48] Wilson JD, Nicklous DM, Aloyo VJ, Simansky KJ. Peptides that regulate food intake. An orexigenic role for μ-opioid receptors in the lateral parabrachial nucleus. Am J Physiol Regul Integr Comp Physiol 2003;285(5):R1055–65.
- [49] Ward HG, Simansky KJ. Chronic prevention of μ-opioid receptor (MOR) G-protein coupling in the pontine parabrachial nucleus persistently decreases consumption of standard but not palatable food. Psychopharmacology 2006;187(4):435–46.
- [50] Simon MJ, García R, Zafra MA, Molina F, Puerto A. Learned preferences induced by electrical stimulation of a food-related area of the parabrachial complex: effects of naloxone. Neurobiol Learn Mem 2007;87(3):332–42.
- [51] Gallo M, Arnedo M, Agüero A, Puerto A. Electrical intracerebral stimulation of the area postrema on TAL. Behav Brain Res 1988;30(3):289–96.
- [52] Agüero A, Arnedo M, Gallo M, Puerto A. Lesions of the lateral parabrachial nuclei disrupt aversion learning induced by electrical stimulation of the area postrema. Brain Res Bull 1993;30(5–6):585–92.
- [53] Simon MJ. Efectos comportamentales de la activación del Complejo Parabraquial Troncoencefálico: Relevancia del subnúcleo Lateral Externo en el Aprendizaje Espacial y Gustativo inducido por estimulación eléctrica o administración enteral de nutrientes. [Behavioral effects of activation

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- of the brainstem parabrachial complex: relevance of the external lateral parabrachial subnucleus in place and taste learning induced by electrical stimulation or enteral administration of nutrients]. Unpublished PhD thesis, 2003, University of Granada, Spain.
- [54] Hawkins RD, Roll PL, Puerto A, Yeomans JS. Refractory periods of neurons mediating stimulation-elicited eating and brain stimulation reward: interval scale measurement and test of a model of neural integration. Behav Neurosci 1983;97(3):416–32.
- [55] Puerto A, Deutsch JA, Molina F, Roll PL. Rapid discrimination of rewarding nutrient by the upper gastrointestinal tract. Science 1976;192:485–7.
- [56] Puerto A, Deutsch JA, Molina F, Roll PL. Rapid rewarding effects of intragastric injections. Behav Biol 1976;18(1):123–4.
- [57] Perez C, Lucas F, Sclafani A. Increased flavor acceptance and preference conditioned by the postingestive actions of glucose. Physiol Behav 1998;64(4):483–92.
- [58] Parker L, Failor A, Weidman K. Conditioned preferences in the rat with an unnatural need state: morphine withdrawal. J Comp Physiol Psychol 1973;82(2):294–300.
- [59] Yeomans MR, Jackson A, Lee MD, Nesic J, Durlach PJ. Expression of flavour preferences conditioned by caffeine is dependent on caffeine deprivation state. Psychopharmacology 2000;150(2):208–15.
- [60] Gallistel CR, Karras D. Pimozide and amphetamine have opposing effects on the reward summation function. Pharmacol Biochem Behav 1984;20(1):73–7.
- [61] Cubero I, Puerto A. Electrical stimulation of the insular cortex induces flavor-preferences in rats. Brain Res 2000;872(1–2):134–40.
- [62] Salamone JD. The involvement of nucleus accumbens dopamine in appetitive and aversive motivation. Behav Brain Res 1994;61(2):117–33.
- [63] Di Chiara G. Drug addiction as dopamine-dependent associative learning disorder. Eur J Pharmacol 1999;375(1–3):13–30.
- [64] Small DM, Gregory MD, Mak YE, Gitelman D, Mesulam MM, Parrish T. Dissociation of neural representation of intensity and affective valuation in human gustation. Neuron 2001;39(4):701–11.

- [65] Reynolds SM, Berridge KC. Positive and negative motivation in nucleus accumbens shell: bivalent rostrocaudal gradients for GABA-elicited eating. Taste "liking"/"disliking" reactions, place preference/avoidance, and fear. J Neurosci 2002;22(16):7308–20.
- [66] Heinrichs SC, Britton KT, Koob GF. Both conditioned taste preference and aversion induced by corticotropin-releasing factor. Pharmacol Biochem Behav 1991;40(4):717–21.
- [67] Kanarek R, Przypek J, D'ancik E, Marks-Kaufman R. Dietary modulation of Mu and kappa opioid receptor-mediated analgesia. Pharmacol Biochem Behav 1997;58(1):43–9.
- [68] Smith DV, Liu H, Vogt MB. Neural coding of aversive and appetitive gustatory stimuli: interactions in the hamster brain stem. Physiol Behav 1994;56(6);1189–96.
- [69] Sewards TV. Dual separate pathways for sensory and hedonic aspects of taste. Brain Res Bull 2004;62(4):271–3.
- [70] Tkacs NC, Li J. Immune stimulation induces Fos expression in brainstem amygdala afferents. Brain Res Bull 1999;48(2):223–31.
- [71] Moufid-Bellancourt S, Razafimanalina R, Velley L. Interaction between mu and kappa receptors located in the parabrachial area in the opioid control of preference threshold for saccharin: modulatory role of lateral hypothalamic neurons. Behav Pharmacol 1996;7:798–809.
- [72] Shippenberg TS, Elmer GI. The neurobiology of opiate reinforcement. Crit Rev Neurobiol 1998;12(4):267–303.
- [73] Rolls ET. Memory systems in the brain. Ann Rev Psychol 2000;51: 599–630.
- [74] O'Doherty JO, Kringelbach ML, Rolls ET, Hornak J, Andrews C. Abstract reward and punishment representations in the human orbitofrontal cortex. Nat Neurosci 2001;4(1):95–102.
- [75] Trifunovic R, Reilly S. Medial versus lateral parabrachial nucleus lesions in the rat: effects on mercaptoacetate-induced feeding and conditioned taste aversion. Brain Res Bull 2002;58(1):107–13.
- [76] Paxinos G, Watson C. The rat brain in stereotaxic coordinates. 4th ed. San Diego: Academic Press; 1998.