1	Estimation of non-shivering thermogenesis and cold-induced nutrient oxidation
2	rates: Impact of method for data selection and analysis
3	Guillermo Sanchez-Delgado <sup>a</sup> , Juan M.A. Alcantara <sup>a</sup> , Francisco M. Acosta <sup>a</sup> , Borja
4	Martinez-Tellez <sup>a,b</sup> , Francisco J. Amaro-Gahete <sup>a,c</sup> , Lourdes Ortiz-Alvarez <sup>a</sup> , Marie Löf
5	<sup>d,e</sup> , Idoia Labayen <sup>f</sup> , Jonatan R. Ruiz <sup>a</sup>
6	
7	<sup>a</sup> PROFITH (PROmoting FITness and Health through Physical Activity) Research Group,
8	Department of Physical Education and Sport, Faculty of Sport Sciences, University of
9	Granada, Granada, Spain.
10	<sup>b</sup> Division of Endocrinology, and Einthoven Laboratory for Experimental Vascular
11	Medicine, Department of Medicine, Leiden University Medical Center, Leiden,
12	Netherlands.
13	<sup>c</sup> Department of Medical Physiology. School of Medicine. University of Granada,
14	Granada, Spain.
15	<sup>d</sup> Department of Biosciences and Nutrition, Karolinska Institutet, NOVUM, Huddinge,
16	Sweden
17	<sup>e</sup> Department of Medical and Health Sciences, Faculty of Medicine and Health, Linköping
18	University, 581 83 Linköping, Sweden
19	<sup>f</sup> Department of Health Sciences, Public University of Navarra, Avda. Barañáisn s/n,
20	31008, Pamplona, Spain
21	
22	Corresponding author:
23	Guillermo Sanchez-Delgado. Department of Physical Education and Sport, Faculty of
24	Sport Sciences. University of Granada, Granada, Spain. Crta de Alfacar s/n C.P. 18071.
25	E-mail: gsanchezdelgado@ugr.es; tel.: 0034 958242754; Fax: 0034 958244369

#### 27 ABSTRACT

Background & Aims: Since the discovery of active brown adipose tissue in human adults, non-shivering cold-induced thermogenesis (CIT) has been regarded as a promising tool to combat obesity. However, there is a lack of consensus regarding the method of choice to analyze indirect calorimetry data from a CIT study. We analyzed the impact of methods for data selection and methods for data analysis on measures of cold-induced energy expenditure and nutrient oxidation rates.

Methods: Forty-four young healthy adults Caucasians (22.1±2.1 years old, 25.6±5.2 34 kg/m<sup>2</sup>, 29 women) participated in the study. RMR, CIT, and cold-induced nutrient 35 oxidation rates were estimated by indirect calorimetry under fasting conditions during 36 one hour of cold exposure combining air conditioning (19.5-20°C) and a water perfused 37 38 cooling vest set at a temperature of 4°C above the individual shivering threshold. We applied three methods for data selection: (i) time intervals every 5 minutes (5min-TI), (ii) 39 the most stable 5-minute period of every forth part of the cold exposure (5min-SS-4P), 40 41 and (iii) the most stable 5-minute period of every half part of the cold exposure (5min-42 SS-2P). Lately we applied two methods for data analysis: (i) area under the curve as a percentage of the baseline RMR (AUC) and; (ii) the difference between EE at the end of 43 44 the cold exposure and baseline RMR (Last-RMR).

**Results:** Mean overall CIT estimation ranged from  $11.6\pm10.0$  to  $20.1\pm17.2$  %RMR depending on the methods for data selection and analysis used. Regarding methods for data selection, *5min-SS-2P* did not allow to observe physiologically relevant phenomena (e.g. metabolic shift in fuel oxidation; P=0.547) due to a lack of resolution. The *5min-TI* and *5min-SS-4P* methods for data selection seemed to be accurate enough to observe physiologically relevant phenomena (all P<0.014), but not comparable for estimating over-all CIT and cold-induced nutrient oxidation rates (P<0.01). Regarding methods for data analysis, the AUC seemed to be less affected for data artefacts and to be morerepresentative in participants with a non-stable energy expenditure during cold exposure.

Conclusions: The methods for data selection and analysis can have a profound impact on CIT and cold-induced nutrient oxidation rates estimations, and therefore, it is mandatory to unify it across scientific community to allow inter-study comparisons. Based on our findings, *5min-TI* should be considered the method of choice to study dynamics (i.e. changes across time) of CIT and cold-induced nutrient oxidation rates, while *5min-SS-4P* and AUC should be the method of choice when computing CIT and cold-induced nutrient oxidation rates as a single value.

Keywords: Cold-induced thermogenesis, adaptive thermogenesis, indirect calorimetry,
metabolic rate, energy balance, obesity.

## 63 ABBREVIATIONS

- 64 BAT: Brown adipose tissue.
- 65 CIT: Cold-induced thermogenesis.
- 66 RMR: Resting metabolic Rate.
- 67 SS: Steady state.
- 68 TI: Time interval.
- 69 EE: Energy expenditure.
- 70 AUC: Area under the curve.
- 71 CCM: CCM Express (Medgraphics Corp, Minnesota, USA).
- 72 MGU: Ultima CardiO2 (Medgraphics Corp, Minnesota, USA).
- 73 DXA: Dual X-ray absorciometry.
- 74 RER: Respiratory exchange ratio.
- 75 CHOox: Carbohydrates oxidation.
- 76 FATox: Fat oxidation.
- 77 ANOVA: Analyses of variance.
- 78 5min-TI: mean values of every consecutive 5-minute period.
- 5 5min-SS-4P: The most stable 5-minute period of every forth part of the cold exposure.
- 5min-SS-2P: The most stable 5-minute period of every half part of the cold exposure.

### 81 INTRODUCTION

82 In simple terms, obesity results from a positive energy balance (i.e. lower energy expenditure than energy intake), and thus, weight loss would be easily achieved inducing 83 84 a negative energy balance. However, many physiological and behavioral adaptations occur in parallel to caloric restriction, making weight loss unsuccessful in long-term [1]. 85 Currently there are no non-invasive successful strategies to achieve sustainable weight 86 loss, and new strategies have to be explored [1]. During the last decade, brown adipose 87 tissue (BAT) activation has been regarded as a possible solution to the obesity problem 88 [2]. 89

BAT was confirmed to be present and active in adult humans in 2009 [3–6]. Since them, 90 91 BAT has been considered a promising therapeutic target due to its capacity to oxidize 92 glucose and lipids for heat producing purposes, in a process known as non-shivering 93 thermogenesis. In murine models, BAT thermogenesis can account up to 60% of total 94 energy expenditure [7]. In humans, BAT is much more scarce than in murine [8] and there 95 is an open debate on whether BAT activity can significantly influence human energy expenditure [9,10]. Noteworthy, even assuming the most pessimistic views of BAT 96 potential to contribute to energy expenditure [11–14], non-shivering thermogenesis can 97 be mediated by other tissues, such as withe adipose tissue and skeletal muscle [12,13,15], 98 and seems to be relevant enough to be considered a possible solution to the weight loss 99 100 maintenance problem [1].

101 Cold-induced thermogenesis (CIT) can be broadly divided into shivering and non-102 shivering thermogenesis [16], although both processes can occur concomitantly [12]. CIT 103 and the associated changes in nutrient oxidation rates are commonly measured by indirect 104 calorimetry [17]. Indirect calorimetry data are quite variable minute by minute, and 105 methods for data selection based on steady state (SS) periods are often necessary to

minimize individuals' and instrument's variability [17-22]. Alternatively, selection of 106 107 predefined (i.e. not considering data variability) time intervals (TI) is commonly made 108 [18]. Besides how to select data, it is often necessary to compute CIT as a unique value 109 to be used in cross-sectional studies, such as to study the association between BAT and CIT [23]. Therefore, investigators have used area under the curve calculations (AUC) 110 111 and/or the difference between EE at the end of cold exposure and baseline RMR (Last-112 RMR) [20,24,25]. These methods for data selection (i.e. SS or TI) and analysis (i.e. AUC or Last-RMR) significantly impact on RMR or meal-induced thermogenesis estimations 113 [18–21]. However, the impact of the chosen method for data selection and data analysis 114 115 on the overall measure of CIT and cold-induced nutrient oxidation rates is largely 116 unknown.

The aim of the present study was to analyze the impact of methods for data selection (TI and SS) and methods for data analysis (AUC and Last-RMR) on measures of coldinduced energy expenditure and nutrient oxidation rates. Despite large scientific interest in cold-induced thermogenesis during the last decade [1,15,26], to our knowledge, there are no studies evaluating the impact of various methodologies on the measurement of cold-induced energy expenditure in healthy human.

#### 123 MATERIAL AND METHODS

#### 124 Participants

A total of 63 participants (45 women) participated in the study. The participants were part of the ACTIBATE study, an exercise-based randomized controlled trial (clinicaltrial.gov: NCT02365129) [27]. All participants were young (18-25 years old), healthy, sedentary (<20 min physical activity on <3 days/week), did not smoke or take any medication, had a stable body weight for the past 3 months (<3 Kg change), and were not regularly exposed to cold. The evaluations were performed between October 11<sup>th</sup> and November 29<sup>th</sup>, 2016.

The study protocol and informed consent were performed in accordance with the last revision of the Declaration of Helsinki. The study was approved by the Human Research Ethics Committee of the University of Granada (n°924) and of the Servicio Andaluz de Salud (Centro de Granada, CEI-Granada).

### 136 Previous conditions to the study days

Participants came to the lab on two separate occasions (5-7 days apart). They were asked to come by bus or by car, under fasting conditions (at least 6 hours), to sleep as usual, to refrain from any moderate (in the previous 24 hours) or vigorous (in the previous 48 hours) physical activity, and not consume alcoholic or stimulant beverages over the past for 6 hours. The participants were evaluated between 8.30 and 19.15hrs. For nutrient oxidation rates analysis, only the participants with a fasting time between 6 and 8 hours were considered [28,29].

## 144 Shivering threshold test

During the first study day, we determined the individual shivering threshold. The 145 146 procedure for the shivering threshold determination has been extensively described elsewhere [30,31]. In brief, participants dressed-up with standardized clothes and stayed 147 148 seated in a warm room (22.1±1.6°C) for 30 minutes, before entering the cold (air cooled) room (19.8±0.5°C), where they were dressed in a temperature-controlled water perfused 149 150 cooling vest (Polar Products Inc., Ohio, USA) and seated again. The water temperature 151 decreased progressively from 16.6°C until 3.8°C or until shivering occurred. Shivering was determined visually and by asking the participants if they were experiencing 152 shivering. 153

## 154 CIT and cold-induced nutrient oxidation rates determination

In the second day, the participants performed the CIT test at the approximate same time 155 of the day at which the shivering threshold test was performed. They dressed-up with the 156 157 same standardized clothes as in the shivering threshold test. Later, they were moved into 158 the warm room (23.2±0.7°C). Before being evaluated, all participants lay down on a reclined bed, in a supine position, and were covered by a sheet for 20 minutes. They were 159 instructed to breathe normally, and not to talk, fidget, or sleep. Thereafter, we assessed 160 the RMR maintaining the same standardized conditions [17] during 30 minutes (Figure 161 162 1).

After assessing RMR, the participants were moved into the cold room (19.7±0.4°C) and they put on the temperature-controlled water perfused cooling vest (Polar Products Inc., Ohio, USA) set 4°C above the individual shivering threshold temperature. Once they had the cooling vest on, they lay down on another reclined bed Indirect calorimetry measurement was performed during two consecutive 30-minute periods, separated by a 5-minute pause to recalibrate the metabolic cart (Figure 1).

The indirect calorimetry measurements for both RMR and CIT were performed with the 169 170 CCM Express (CCM) or with the Ultima CardiO2 (MGU) (Medgraphics Corp, Minnesota, USA), using a neoprene face-mask equipped with a directconnect<sup>TM</sup> 171 172 metabolic flow sensor (Medgraphics Corp, Minnesota, USA) [21]. Flow calibration was performed by a 3-L calibration syringe at the beginning of every testing day, and gas 173 174 analyzers were calibrated using 2 standard gas concentrations following the 175 manufacturer's instructions before every 30 minutes of indirect calorimetry measurement. We used the same metabolic cart for the RMR and CIT measurements in every 176 participant. 177

178 Body composition was measured by a DXA scanner (Discovery Wi, Hologic, Inc.,

179 Bedford, MA, USA) and data were extracted from the Hologic APEX 4.0.2. (Hologic,

180 Inc., Bedford, MA, USA), and weight and height were measured with a Seca scale and a

181 stadiometer (model 799, Electronic Column Scale, Hamburg, Germany).

### 182 Methods for data selection and analysis

Indirect calorimetry data were averaged every minute and downloaded from the Breeze Suite (8.1.0.54 SP7) software. For RMR, we selected the most stable 5-minute period (i.e. the one with the lowest average of coefficients of variance for oxygen consumption, carbon dioxide production, minute ventilation, and respiratory exchange ratio (RER)) [21].

For CIT, we applied different methods for data selection and analysis. Methods for data selection refer to the way of processing the data obtained from the continuous indirect calorimetry instrument. After excluding the first 5 minutes of every 30-minute record [18], we used three different methods for data selection (Figure 1): i) TI every 5 minutes (*5min-TI*): mean values of every consecutive 5-minute period (i.e. from the 6<sup>th</sup> to the 10<sup>th</sup>,

193	from the 11 <sup>th</sup> to the 15 <sup>th</sup> , etc.); ii) The most stable 5-minute period of every forth part of
194	the cold exposure (i.e. after dividing the cold exposure into 4 parts equal in length) (5min-
195	SS-4P); iii) The most stable 5-minute period of every half part of the cold exposure (i.e.
196	after dividing the cold exposure into 2 parts equal in length) (5min-SS-2P).

- 197 In order to express the CIT as a single value, we used two different methods for data
- analysis (Figure 1): i) The AUC following the trapezoidal rule; and ii) the Last-RMR.
- Both methods for data analysis were expressed as a percentage of the baseline RMR.



#### 200

Figure 1. Cooling protocol, methods for data selection, and methods for data analysis. White rectangles in cooling protocol represent every 30 minutes of recorded gas exchange. Gray squares represent the 5-minute selected period within a specific recorded time (i.e. in the time interval method: average of every consecutive 5-minute period; in the steady state method: the 5 minute-period with the lowest average of coefficient of variances of: oxygen consumption, carbon dioxide production, respiratory exchange ratio, and minute ventilation). Crosses represent excluded gas exchange data (i.e. first 5 minutes of every 30-minute record). Vertical lines within white rectangles represent divisions of recorded data for the selection of the representative 5-minute period. TI: Time interval; SS: Steady State; min: minutes; 4P: Four periods; 2P: Two periods; RMR: Resting metabolic rate; AUC: Area under the curve. Oxygen consumption and carbon dioxide production for each selected data point were used to estimate EE, and carbohydrates (CHOox) and fat oxidation (FATox). EE was estimated through Weir's abbreviated equation, not considering urinary nitrogen concentration [32]. For carbohydrates and fat oxidation estimations, we used Frayn's equation, not considering urinary nitrogen concentration [33].

#### 213 Statistical analysis

Results are presented as means  $\pm$  standard deviation, unless otherwise stated. The analyses were conducted using the Statistical Package for Social Sciences (SPSS, v. 21.0,

216 IBM SPSS Statistics, IBM Corporation), and the level of significance was set at <0.05.

A repeated-measures analysis of variance (ANOVA) was used to test differences in EE and nutrient oxidation rates across the selected data points following the different methods for data selection and analysis. To compare CIT, cold-induced CHOox, and FATox estimations obtained with different combinations of methods for data selection and analysis, we conducted a two-factor (method for data selection \* method for data analysis) ANOVA. Bonferroni corrections (automatically performed by the SPSS) were used to perform post hoc comparisons.

#### 224 **RESULTS**

225 During the CIT, visually detected and auto-reported shivering was recorded in 17 226 participants (n=16 women) who were therefore excluded from further analysis. In 227 addition, participants with RER values higher than 1.1 or lower than 0.7 in any measure point, or a RER higher than 1.0 in RMR assessment, were also excluded from the analysis 228 (n=2) [17]. Finally, a total of 44 participants were included in the energy expenditure 229 230 analysis (Table 1). Mean fasting time was 9±3.7 hours. Of this sample, a total of 18 (n=13 231 women) strictly met the fasting time criterion for assessing nutrient oxidation rates (i.e. a fasting time of 6-8 hours) and were included in the nutrient oxidation rate analysis (Table 232 233 S1).

Table 1. Descriptive characteristics of the participants included in the energy expenditureanalysis.

	All (1	All (n=44)		Male (n=15)		Female (n=29)	
Age (years)	22.1	(2.1)	22.4	(2.2)	22.0	(2.2)	
BMI (kg/m <sup>2</sup> )	25.6	(5.2)	27.9	(6.0)	24.4	(4.4)	
Lean mass (kg)	42.7	(10.4)	54.6	(6.8)	36.4	(5.2)	
Fat mass (kg)	27.2	(10.6)	29.9	(13.5)	25.8	(8.8)	
Fat mass (%)	37.0	(8.0)	32.7	(8.5)	39.2	(6.9)	
VO <sub>2</sub> (ml/min)	223	(39)	252	(45)	208	(23)	
VCO <sub>2</sub> (ml/min)	192	(33)	216	(37)	178	(20)	
RMR (kcal/day)	1564	(277)	1769	(324)	1459	(178)	
RER	0.862	(0.054)	0.863	(0.048)	0.861	(0.057)	

236 Data are presented as means (standard deviation). BMI: Body mass index; VO<sub>2</sub>: resting oxygen

consumption; VCO<sub>2</sub>: resting carbon dioxide production; RMR: Resting Metabolic Rate; RER:
resting respiratory exchange ratio.

239

240 Cold-induced thermogenesis

Figure 2 shows the EE dynamics during a mild cold exposure by method for data selection. EE was significantly increased by mild cold exposure, which was detected regardless of the method for data selection used (All P<0.001). Post-hoc comparisons showed that for all methods for data selection, EE was increased just after starting the cold-exposure (i.e. first data point analyzed) and remained unchanged until the end of themild cold exposure.





248 Figure 2. Energy expenditure (EE) during mild cold exposure by methods for data selection. Represented values are mean ± standard error. Panel A presents data obtained 249 250 with the 5-minute time interval (TI) method; Panel B presents data obtained with the steady state (SS) method after dividing the cold exposure into 4 periods; Panel C presents 251 data obtained with the steady state method after dividing the cold exposure into 2 periods. 252 Min 0 represents the value obtained in the resting metabolic rate (RMR) period (baseline). 253 \*: Significantly different to baseline value. P value for repeated measures ANOVA. Min: 254 minutes; Kcal: Kilocalories. 255

Mean overall CIT estimation ranged from 11.6±10.0 to 20.1±17.2 %RMR depending on
the methods for data selection and analysis used.

258 Figure S1 shows the individual data for the over-all estimation of CIT by different 259 combinations of methods for data selection and methods for data analyses. Figure 3 260 compares the mean overall CIT estimation obtained by the different methods for data 261 selection and analysis. Both main effects (methods for data selection and methods for data 262 analysis) were significant (all P<0.01) and no significant interaction effect (method for data selection \* method for data analysis) was found (P=0.3). Mean overall CIT 263 estimation was consistently higher with the Last-RMR than with the AUC in all methods 264 265 for data selection (all paired comparisons P≤0.043). No differences in mean over-all CIT estimation were found between different methods for data selection when using the Last-266 RMR (all P=0.6). However, mean over-all CIT estimation varied across methods for data 267 268 selection when using the AUC (P<0.001).





Figure 3. Comparisons between mean over-all cold-induced thermogenesis (CIT) 270 obtained with different methods for data selection and analysis. Represented values are 271 mean  $\pm$  standard error. P values for paired *t*-test. \*: significant differences with the rest of 272 the AUC results after Bonferroni correction. RMR: Resting metabolic rate; AUC: Area 273 274 under the curve; Last-RMR: Last period value minus RMR value.5min-TI: mean values of every consecutive 5-minute period; 5min-SS-4P: The most stable 5-minute period of 275 every forth part of cold exposure; 5min-SS-2P: The most stable 5-minute period of every 276 277 half part of cold exposure.

278

### 280 *Cold-induced nutrient oxidation rates*

- Figure 4 shows CHOox and FATox dynamics during the mild cold exposure by method
- for data selection. There were significant changes in CHOox when selecting the data by
- 283 5min-TI and 5min-SS-4P (all P<0.015), but not with 5min-SS-2P (P=0.547). Of note,
- differences between CHOox at 30 minutes and at the end of the mild cold exposure were
- only detected with *5min-TI*. FATox changes during the mild cold exposure were detected
- with all methods for data selection (All P<0.002). The highest FATox rate was observed
- at 30 minutes with 5min-TI and 5min-SS-4P, but not with 5min-SS-2P. A reduction on
- 288 FATox after minute 30 was only detected by 5min-TI.

CHOox

FATox



290 Figure 4. Nutrient oxidation rates during cold exposure by method for data selection. 291 Panels A and B present data obtained with the 5-minute time interval (TI) method; panels C and D present data obtained with the steady state (SS) method after dividing the cold 292 exposure into 4 periods; and panels E and F present data obtained with the steady state 293 294 method after dividing the cold exposure into 2 periods. Represented values are mean  $\pm$ standard error. P value for repeated measures ANOVA. Equal lower-case letters indicate 295 296 significant differences after Bonferroni correction. CHO: Carbohydrates. min: minutes; 297 g: grams.

- 298 Regarding mean overall cold-induced nutrient oxidation rates estimation, no differences
- 299 were found when comparing the methods for data selection (P=0.181), nor when
- 300 comparing the methods for data analysis (P=0.328) (Figure S2).

#### 301 **DISCUSSION**

302

303 5min-SS-2P) in combination with two different methods for data analysis (AUC and Last-304 RMR) on estimations of cold-induced energy expenditure and nutrient oxidation rates 305 during a 60-minute individualized mild cold exposure, designed to elicit maximum non-306 shivering thermogenesis. The 5min-TI and 5min-SS-4P methods for data selection seemed 307 to be accurate enough to observe physiologically relevant phenomena, but not comparable 308 for estimating over-all CIT and cold-induced nutrient oxidation rates. Regarding methods for data analysis, the AUC seemed to be less affected for data artefacts and be more 309 310 representative in participants with a non-stable energy expenditure during cold exposure. 311 The selection of the method for data selection and analysis influences the estimations of RMR and meal-induced thermogenesis [17-22]. Therefore, it is expected that the 312 313 selection of the method for data selection and analysis also influences the estimation of 314 CIT and cold-induced nutrient oxidation rates. Regarding the methods for data selection, 315 5min-SS-2P may not be an appropriate method, since, as a consequence of a lack of resolution, it does not allow to detect relevant physiological changes. In contrast, 5min-316 317 TI allows to detect changes that no other method is able to detect (see Figure 4). However, 318 for the RMR data selection, there is a consensus on the need of using a method based on the selection of a SS, as it is supposed not to be affected by artefacts, and to ensure a more 319 320 valid measure [19,21,22]. Therefore, 5min-SS-4P could be the method of choice. Our data 321 supports that selection, especially when an over-all CIT estimation is made. In this case, the outcome obtained with the 5min-TI method might be affected by artefacts (see Figure 322 323 S2A), as previously argued [19,21,22]. Indeed, we observed a wider range of CIT values

This study analyzed the impact of methods for data selection (5min-TI, 5min-SS-4P and

324 applying *5min-TI* (-14.9/46.2 %RMR) than *5min-SS-4P* (-14.8/39.9 %RMR) (see Figure

S1). On the other hand, *5min-TI* might be the method of preference when studying the

dynamics (i.e. changes during time of cold-exposure) of CIT or cold-induced nutrient
oxidation rates, as it allows a more detailed insight (Figure 2 and 5). Standardizing the
methods for data selection and analysis would allow between-studies comparability.

329 In relation to the methods for data analysis, the AUC resulted in a lower inter-individual variability than the Last-RMR (see figure S1). We observed a stable EE during the mild 330 331 cold exposure, and consequently one could expect no differences between the AUC and the Last-RMR in over-all CIT estimation. In contrast, we observed large differences 332 333 between the AUC and the Last-RMR over-all CIT estimation, with the Last-RMR reporting higher values (Figure 3). This could be explained by the fact that energy 334 335 expenditure progressively increases during the mild cold exposure in some individuals while in others did not. This, together with the possibility of the Last-RMR methods to 336 be influenced by artefacts (see outlier in Figure S1) would point to the AUC as the method 337 of choice for over-all CIT estimation. 338

339 Observations on humans' CIT have reported huge inter-individual variability [26,34,35]. 340 This is congruent with our results, where some individuals showed negative values of CIT (i.e. lower EE in cold than in RMR) while others even get more than 100% increase 341 342 over RMR with some methods for data analysis. Many factors have been reported to 343 contribute to inter-individual CIT difference. [26]. Here, we show that the method for 344 data selection and analysis could have an important impact on inter-individual CIT 345 variability estimations. This is in line with observations about the impact of the method 346 for data selection and analysis on RMR estimations [17–19,21].

347 Limitations

Our results should be considered with caution due to the presence of limitations. Firstly,we did not analyze urine nitrogen excretion, and therefore we could not correct the

nutrient oxidation rates for protein oxidation. Although protein oxidation correction 350 351 would have been desirable, it is not plausible to obtain different urine nitrogen concentration in short intervals such as the periods that we have studied (i.e.  $\leq 60$  min). 352 Secondly, although we selected a cooling protocol thought to ensure maximum non-353 shivering thermogenesis, we cannot be sure of the relative contribution to CIT of 354 355 shivering thermogenesis [12]. However, we excluded from the analysis participants who 356 reported shivering or whose shivering was visually detected, and therefore it is probable than the contribution of non-shivering thermogenesis is predominant in the included 357 participants. Thirdly, we used two different metabolic carts which are not comparable and 358 359 have relatively low reliability [36–38]. However, the within-subject design applied in this 360 study reduce the impact of this limitation. Finally, our results only apply to young healthy individuals, and further studies are needed to confirm whether this also applies to older 361 362 and unhealthy individuals.

363 Conclusions

The methods for data selection and analysis can have a profound impact on CIT and coldinduced nutrient oxidation rates estimations, and therefore, it is mandatory to unify it across scientific community to allow inter-study comparisons. Based on our findings, *5min-TI* should be considered the method of choice to study dynamics (i.e. changes across time) of CIT and cold-induced nutrient oxidation rates, while *5min-SS-4P* and AUC should be the method of choice when computing CIT and cold-induced nutrient oxidation rates as a single value.

### 371 ACKNOWLEDGEMENTS

372 The study was supported by the Spanish Ministry of Economy and Competitiveness, Fondo de Investigación Sanitaria del Instituto de Salud Carlos III (PI13/01393), and Retos 373 374 de la Sociedad (DEP2016-79512-R), Fondos Estructurales de la Unión Europea (FEDER), by the Spanish Ministry of Education (FPU 13/04365, FPU14/04172, and 375 15/04059), by the Fundación Iberoamericana de Nutrición (FINUT), by the Redes 376 temáticas de investigación cooperativa RETIC (Red SAMID RD16/0022), by 377 AstraZeneca HealthCare Foundation and by the University of Granada, Plan Propio de 378 Investigación 2016, Excellence actions: Units of Excellence; Unit of Excellence on 379 380 Exercise and Health (UCEES). This study is part of a Ph.D. Thesis conducted in the Biomedicine Doctoral Studies of the University of Granada, Spain. We are grateful to 381 Ms. Carmen Sainz-Quinn for assistance with the English language. 382

## 383 AUTHOR'S CONTRIBUTION

GSD, JMA, and JRR conceived the study; GSD, JMA, ML, IL and JRR designed the
study; GSD, JMA, FMA, BMT, FAG and LOA did the data collection; GSD performed
the statistical analyses and drafted the manuscript. All authors read and approved the final
manuscript.

### 388 CONFLICT OF INTEREST SOURCES

389 The authors confirm that there are no conflicts of interest.

#### **390 REFERENCES**

- 391 [1] Palmer BF, Clegg DJ. Non-shivering thermogenesis as a mechanism to facilitate
  392 sustainable weight loss. Obes Rev 2017:1–13. doi:10.1111/obr.12563.
- Lee P, Swarbrick MM, Ho KKY. Brown adipose tissue in adult humans: a
- metabolic renaissance. Endocr Rev 2013;34:413–38. doi:10.1210/er.2012-1081.
- 395 [3] Cypess AM, Lehman S, Williams G, Tal I, Rodman D, Goldfine AB, et al.
- Identification and importance of brown adipose tissue in adult humans. N Engl J
  Med 2009;360:1509–17. doi:10.1056/NEJMoa0810780.
- 398 [4] Saito M, Okamatsu-Ogura Y, Matsushita M, Watanabe K, Yoneshiro T, Nio-
- Kobayashi J, et al. High incidence of metabolically active brown adipose tissue
  in healthy adult humans: effects of cold exposure and adiposity. Diabetes
  2009;58:1526–31. doi:10.2337/db09-0530.

2009,50.1520,51.401.10.2557/4009,0550.

- 402 [5] van Marken Lichtenbelt WD, Vanhommerig JW, Smulders NM, Drossaerts JM a
- 403 FL, Kemerink GJ, Bouvy ND, et al. Cold-activated brown adipose tissue in
- 404 healthy men. N Engl J Med 2009;360:1500–8. doi:10.1056/NEJMoa0808718.
- 405 [6] Virtanen KA, Lidell ME, Orava J, Heglind M, Westergren R, Niemi T, et al.
- 406 Functional brown adipose tissue in healthy adults. N Engl J Med 2009;360:1518–
- 407 25. doi:10.1056/NEJMoa0808949.
- 408 [7] Heldmaier G, Buchberger A. Sources of heat during nonshivering thermogenesis
- 409 in Djungarian hamsters: a dominant role of brown adipose tissue during cold
  410 adaptation. J Comp Physiol B 1985;156:237–45.
- 411 [8] Cypess AM, White AP, Vernochet C, Schulz TJ, Xue R, Sass CA, et al.
- 412 Anatomical localization, gene expression profiling and functional

413		characterization of adult human neck brown fat. Nat Med 2013;19:635–9.
414		doi:10.1038/nm.3112.
415	[9]	Blondin DP, Carpentier AC. The role of BAT in cardiometabolic disorders and
416		aging. Best Pract Res Clin Endocrinol Metab 2016;30:497–513.
417		doi:10.1016/j.beem.2016.09.002.
418	[10]	Leitner BP, Huang S, Brychta RJ, Duckworth CJ, Baskin AS, McGehee S, et al.
419		Mapping of human brown adipose tissue in lean and obese young men. Proc Natl
420		Acad Sci U S A 2017:6–11. doi:10.1073/pnas.1705287114.
421	[11]	Jensen MD. Brown adipose tissuenot as hot as we thought. J Physiol
422		2015;593:489. doi:10.1113/jphysiol.2014.287979.
423	[12]	Blondin DP, Labbé SM, Phoenix S, Guérin B, Turcotte ÉE, Richard D, et al.
424		Contributions of white and brown adipose tissues and skeletal muscles to acute
425		cold-induced metabolic responses in healthy men. J Physiol 2015;593:701-14.
426		doi:10.1113/jphysiol.2014.283598.
427	[13]	U Din M, Raiko J, Saari T, Kudomi N, Tolvanen T, Oikonen V, et al. Human
428		brown adipose tissue [(15)O]O2 PET imaging in the presence and absence of
429		cold stimulus. Eur J Nucl Med Mol Imaging 2016;43:1878–86.
430		doi:10.1007/s00259-016-3364-y.
431	[14]	Muzik O, Mangner TJ, Leonard WR, Kumar A, Janisse J, Granneman JG. 150
432		PET measurement of blood flow and oxygen consumption in cold-activated
433		human brown fat. J Nucl Med 2013;54:523–31. doi:10.2967/jnumed.112.111336.
434	[15]	Betz MJ, Enerbäck S. Targeting thermogenesis in brown fat and muscle to treat
435		obesity and metabolic disease. Nat Rev Endocrinol 2017.

436

doi:10.1038/nrendo.2017.132.

437	[16]	Blondin DP, Tingelstad HC, Mantha OL, Gosselin C, Haman F. Maintaining
438		thermogenesis in cold exposed humans: Relying on multiple metabolic pathways.
439		Compr Physiol 2014;4:1383–402. doi:10.1002/cphy.c130043.
440	[17]	Fullmer S, Benson-Davies S, Earthman CP, Frankenfield DC, Gradwell E, Lee
441		PSP, et al. Evidence analysis library review of best practices for performing
442		indirect calorimetry in healthy and non-critically ill individuals. J Acad Nutr Diet
443		2015;115:1417-1446.e2. doi:10.1016/j.jand.2015.04.003.
444	[18]	Borges JH, Langer RD, Cirolini VX, Páscoa MA, Guerra-Júnior G, Gonçalves
445		EM. Minimum Time to Achieve the Steady State and Optimum Abbreviated
446		Period to Estimate the Resting Energy Expenditure by Indirect Calorimetry in
447		Healthy Young Adults. Nutr Clin Pract 2016:0884533615627268
448		doi:10.1177/0884533615627268.
449	[19]	Irving CJ, Eggett DL, Fullmer S. Comparing Steady State to Time Interval and
450		Non-Steady State Measurements of Resting Metabolic Rate. Nutr Clin Pract
451		2016. doi:10.1177/0884533616672064.
452	[20]	Ruddick-Collins LC, King N a, Byrne NM, Wood RE. Methodological
453		considerations for meal-induced thermogenesis: measurement duration and
454		reproducibility. Br J Nutr 2013;110:1978-86. doi:10.1017/S0007114513001451.
455	[21]	Sanchez-Delgado G, Alcantara JMA, Ortiz-Alvarez L, Xu H, Martinez-Tellez B,
456		Labayen I, et al. Reliability of resting metabolic rate measurements in young
457		adults: Impact of methods for data analysis. Clin Nutr 2017.
458		doi:10.1016/j.clnu.2017.07.026.

- 459 [22] McClave S a, Spain D a, Skolnick JL, Lowen CC, Kieber MJ, Wickerham PS, et
  460 al. Achievement of steady state optimizes results when performing indirect
  461 calorimetry. JPEN J Parenter Enteral Nutr 2003;27:16–20.
- 462 doi:10.1177/014860710302700116.
- 463 [23] van Marken Lichtenbelt WD, Kingma B, van der Lans A, Schellen L. Cold
  464 exposure--an approach to increasing energy expenditure in humans. Trends
  465 Endocrinol Metab 2014;25:165–7. doi:10.1016/j.tem.2014.01.001.
- Labayen I, Forga L, Martínez J a. Nutrient oxidation and metabolic rate as
  affected by meals containing different proportions of carbohydrate and fat, in
  healthy young women. Eur J Nutr 1999;38:158–66.
- 469 [25] Yoneshiro T, Matsushita M, Nakae S, Kameya T, Sugie H, Tanaka S, et al.
- 470 Brown adipose tissue is involved in the seasonal variation of cold-induced

471 thermogenesis in humans. Am J Physiol Regul Integr Comp Physiol

472 2016;310:ajpregu.00057.2015. doi:10.1152/ajpregu.00057.2015.

473 [26] Brychta RJ, Chen KY. Cold-induced thermogenesis in humans. Eur J Clin Nutr
474 2017;71:345–52. doi:10.1038/ejcn.2016.223.

475 [27] Sanchez-Delgado G, Martinez-Tellez B, Olza J, Aguilera CM, Labayen I, Ortega

476 FB, et al. Activating brown adipose tissue through exercise (ACTIBATE) in

- 477 young adults: Rationale, design and methodology. Contemp Clin Trials
- 478 2015;45:416–25. doi:10.1016/j.cct.2015.11.004.
- 479 [28] Jeukendrup AE, Wallis GA. Measurement of substrate oxidation during exercise
  480 by means of gas exchange measurements. Int J Sport Med Suppl 2005;26.

481 doi:10.1055/s-2004-830512.

- 482 [29] Venables MC, Achten J, Jeukendrup AE. Determinants of fat oxidation during
  483 exercise in healthy men and women: a cross-sectional study. J Appl Physiol
  484 2005;98:160–7. doi:10.1152/japplphysiol.00662.2003.
- 485 [30] Martinez-Tellez B, Sanchez-Delgado G, Garcia-Rivero Y, Alcantara JMA,
- 486 Martinez-Avila WD, Muñoz-Hernandez M V., et al. A new personalized cooling
- 487 protocol to activate brown adipose tissue in young adults. Front Physiol
- 488 2017;8:1–10. doi:10.3389/fphys.2017.00863.
- 489 [31] Martinez-Tellez B, Sanchez-Delgado G, Acosta FM, Alcantara JMA, Boon MR,
- 490 Rensen PCN, et al. Differences between the most used equations in BAT-human
- 491 studies to estimate parameters of skin temperature in young lean men. Sci Rep
- 492 2017;7:10530. doi:10.1038/s41598-017-10444-5.
- 493 [32] Weir JBDB. New methods for calculating metabolic rate with special reference to
  494 protein metabolism. J Physiol 1949;109:1–9.
- 495 [33] Frayn KN. Calculation of substrate oxidation rates in vivo from gaseous
- 496 exchange. J Appl Physiol 1983;55:628–34. doi:0161-7567/83.
- 497 [34] van Marken Lichtenbelt WD, Daanen H a M. Cold-induced metabolism. Curr
- 498 Opin Clin Nutr Metab Care 2003;6:469–75.
- doi:10.1097/01.mco.0000078992.96795.5f.
- 500 [35] van Marken Lichtenbelt WD, Schrauwen P, van de Kerckhove S, Westerterp-
- 501 Plantenga MS. Individual variation in body temperature and energy expenditure
- in response to mild cold. Am J Physiol Metab 2002;282:E1077–83.
- 503 doi:10.1152/ajpendo.00020.2001.
- 504 [36] Alcantara JMA, Sanchez-Delgado G, Martinez-Tellez B, Merchan-Ramirez E,

505		Labayen I, Ruiz JR. Congruent validity and inter-day reliability of two breath by
506		breath metabolic carts to measure resting metabolic rate in young adults. Nutr
507		Metab Cardiovasc Dis 2018. doi:10.1016/j.numecd.2018.03.010.
508	[37]	Graf S, Karsegard VL, Viatte V, Maisonneuve N, Pichard C, Genton L.
509		Comparison of three indirect calorimetry devices and three methods of gas
510		collection: A prospective observational study. Clin Nutr 2013;32:1067–72.
511		doi:10.1016/j.clnu.2013.08.012.
512	[38]	Oshima T, Berger MM, De Waele E, Guttormsen AB, Heidegger C-P, Hiesmayr
513		M, et al. Indirect calorimetry in nutritional therapy. A position paper by the
514		ICALIC study group. Clin Nutr 2017;36:651-62.
515		doi:10.1016/j.clnu.2016.06.010.
516		

# SUPPLEMENTARY MATERIAL

•	All (n=18)		Male (n=5)		Female (n=13)	
Age (years)	21.9	(2.0)	21.6	(2.2)	22.0	(2.0)
BMI (kg/m <sup>2</sup> )	24.3	(4.6)	26.5	(5.3)	23.5	(4.1)
Lean mass (kg)	40.4	(8.0)	50.7	(5.7)	36.5	(4.5)
Fat mass (kg)	25.0	(9.6)	28.4	(13.0)	23.7	(8.2)
Fat mass (%)	36.1	(7.0)	33.5	(7.3)	37.1	(6.9)
VO <sub>2</sub> (ml/min)	222	(33)	248	(43)	211	(22)
VCO <sub>2</sub> (ml/min)	186	(27)	206	(31)	178	(23)
RMR (kcal/day)	1554	(226)	1738	(292)	1483	(157)
RER	0.842	(0.048)	0.835	(0.025)	0.846	(0.055)

**Table S1.** Descriptive characteristics of the participants included in the nutrient oxidation rate analysis.

Data are presented as means (standard deviation). BMI: Body mass index; VO<sub>2</sub>: resting oxygen consumption; VCO<sub>2</sub>: resting carbon dioxide production; RMR: Resting Metabolic Rate; RER: resting respiratory exchange ratio.



**Figure S1.** Individual data for over-all cold-induced thermogenesis (CIT) obtained with different methods for data selection and analysis. Panels A and B represent data obtained with the 5-minute time interval (TI) method for data selection. Panels C and D represent data obtained with the steady state (SS) method for data selection after dividing the cold exposure into 4 periods. Panels E and F represent data obtained with the SS method for data selection after dividing the cold exposure into 2 periods. Regarding the methods for data analysis, panels A, C, and E represent data obtained following the area under the curve method for data analysis, and panels B, D, and F represent data obtained from the difference between the last period of the cold exposure and the warm value. RMR: Resting metabolic rate.



**Figure S2.** Comparisons between mean over-all cold-induced nutrient (carbohydrates (CHO) and fat (FAT)) oxidation rates obtained with different methods for data selection and analysis. TI: Time interval; SS: Steady State; RMR: Resting metabolic rate; AUC: Area under the curve; Last-RMR: Last period value minus RMR value. 5min-TI: mean values of every consecutive 5-minute period; 5min-SS-4P: The most stable 5-minute period of every forth part of the cold exposure; 5min-SS-2P: The most stable 5-minute period of every half part of the cold exposure.