EDITORIAL

NG2 antigen is expressed in CD34 + HPCs and plasmacytoid dendritic cell precursors: is NG2 expression in leukemia dependent on the target cell where leukemogenesis is triggered?

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The neuron-glial antigen 2 (NG2) molecule and its human homolog was first reported on oligodendrocyte progenitor cells.¹ NG2 is recognized by the 7.1 monoclonal antibody.² The physiological role of this molecule remains to be elucidated.³ The expression pattern of NG2 in leukemia is controversial. NG2 expression was initially claimed to be specifically associated with 11q23/mixed-lineage leukemia (MLL) gene rearrangements.⁴ In fact, over the past years, NG2 has been incorporated in diagnostic panels for immunophenotyping of leukemic patients because of its potential predictive value for MLL rearrangements in childhood and adult acute myeloid leukemias.^{2,4-9} However, it has been suggested that 7.1 expression could be specifically associated with only two specific subtypes of leukemia harboring either the translocations t(4;11) (q21;q23) or t(9;11) (p13;q23), which encode for the leukemic fusion genes MLL-AF4 and MLL-AF9, respectively, but not to other MLL rearrangements.¹⁰ Moreover, in the clinic, many leukemic patients harboring MLL rearrangements but lacking NG2 expression are commonly seen (Supplementary Table 1). In turn, we and many others have reported the existence of acute leukemias and plasmacytoid dendritic cell (pDC) leukemias (>50%) lacking MLL rearrangements but expressing NG2¹¹ (Supplementary Table 1).

Over 80 different partners of the human MLL gene have been identified so far, most being associated with a poor clinical outcome.^{12–14} Clearly, this renders the characterization of MLL fusion alleles and its correlation with NG2 expression a very demanding task. Intriguingly, it has been suggested that the expression of NG2 antigen may be dependent on the particular gene(s) paired to MLL when it is rearranged, explaining, at least in part, why some patients with 11q23 balanced translocations can be NG2 negative.^{5,10}

On the basis of the controversial data about the clinical relevance of NG2 expression and its inclusion in diagnostic immunophenotypic panels as well as the existence of NG2-expressing acute leukemias lacking MLL rearrangements, in particular pDC leukemias,¹¹ we aimed (i) to gain further insights into the biological association between NG2 expression and MLL rearrangements; (ii) to analyze whether the expression of NG2 may depend on the particular gene(s) paired to MLL when it is rearranged and (iii) to explore the hypothesis that the expression of NG2 in leukemias lacking MLL rearrangements, such as NG2 + pDC-leukemias, may be due to the existence of a minor subset of CD34 + hematopoietic stem/progenitor cells readily coexpressing NG2 where the leukomogenesis process may be initially triggered.

The following cell lines, chosen based on the status of the MLL locus were used in this study: KG1a (stem cell like-AML), REH (TEL-AML1 + pre-B ALL), 293T (kidney embryonic epithelial cell line), MV4;11 (MLL-AF4 + pro-B ALL), RS4;11 (MLL-AF4 + pro-B ALL) and THP-1 (MLL-AF9 + AML). Three

out of the six (KG1a, REH and 293T) are cell lines without MLL translocations, whereas the other three cell lines (MV4;11, RS4;11 and THP-1) harbor balanced MLL translocations. Importantly, AF4 is the partner gene paired to rearranged MLL in both MV4;11 and RS4;11 cell lines, whereas AF9 is paired to MLL in THP-1. The cell lines were maintained in standard culture conditions: RPMI or DMEM with 10% fetal bovine serum supplemented with nonessential amino acids, 2 mM L-glutamine and antibiotics (all from Gibco, Invitrogen, Grand Island, NY, USA). Genetically stable (manuscript submitted) and fully characterized pluripotent undifferentiated human embryonic stem cells (hESCs) HS181 (kindly provided by Professor O Hovatta, Karolinska Institute) were maintained on mitotically inactivated human embryonic fibroblasts as described previously.¹⁵ Cord blood samples (CB; n = 5) from healthy newborns were obtained from local hospitals with Ethics Board approval. Normal bone marrow (n=6) and granulocyte-colony stimulating factor (G-CSF)-mobilized peripheral blood (n = 1) were obtained from healthy volunteers upon informed consent. The mononuclear cells were isolated using Ficoll-Hypaque (GE Healthcare, Bucks, UK), washed with phosphate-buffered saline +3% fetal bovine serum and the red blood cells lysed (Red Blood Cell-Lysis Solution, BD Biosciences, Erembodegem, Belgium). Then, the mononuclear cells were stained by direct immunofluorescence (see below) and ready for multicolor high-speed flow cytometry analysis.16

The expression of NG2 in CD34 + pDC precursors¹⁶ was assessed by flow cytometry using the following combination of monoclonal antibody in a six-color staining: CD38-FITC/NG2-PE/HLADR-PerCP/CD123-APC/CD34-PECy7/CD45-APC-Cy7. All monoclonal antibody were purchased from Becton Dickinson (San Jose, CA, USA) except the NG2 that is from Immunotech (Marseille, France) and CD123-APC from Miltenyi (Bergisch, Gladbach, Germany). Of note, as many as 2×10^5 CD34 + hematopoietic stem/progenitor cells were gated and analyzed. Plasmacytoid DC precursors were identified as being CD34⁺/CD45⁺/CD38⁺/CD123^{high}/HLADR⁺, as described recently.¹⁶ From an ontogeny standpoint, the expression of NG2 was assessed in hESCs (prenatal stage of in utero development) and CB samples (neonatal stage of hematopoietic development), because a high frequency of primary leukemias with rearranged MLL gene occurs during infancy/childhood.¹⁷ NG2 expression was also analyzed in adult bone marrow samples.

Among the cell lines without MLL rearrangements, the myeloid cell line KG1a lacked NG2 expression, whereas REH was NG2 + (Figure 1a). Interestingly, the nonhematopoietic cell line 293T was also clearly positive for NG2. This data confirm the expression of NG2 in hematopoietic and nonhematopoietic cell lines without MLL rearrangements, further supporting the existence of human leukemias lacking MLL translocations but expressing the NG2 antigen¹¹ (Figure 1a and Supplementary Table 1).

Among the MLL translocation-harboring cell lines, THP-1 displayed clear expression of NG2. Intriguingly, both the

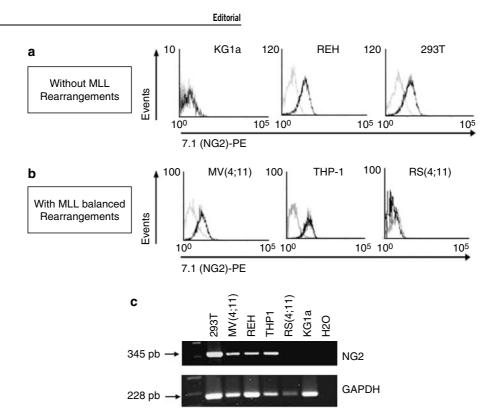


Figure 1 Flow cytometry analysis of NG2 (7.1) antigen in transformed hematopoietic cell lines. (**a**) Expression of NG2 in transformed cell lines lacking MLL gene rearrangements. (**b**) Expression of NG2 in transformed cell lines carrying balanced MLL gene rearrangements. Gray lines represent the irrelevant isotype-matched control. (**c**) reverse transcriptase showing the NG2 mRNA expression in different leukemic cell lines. MLL, mixed-lineage leukemia; NG2, neuron-glial antigen 2.

MV4;11 and RS4;11 cell lines, while sharing MLL gene rearrangements fused to the same partner gene (AF4), displayed a different profile of NG2 expression: MV4;11 cells were NG2-positive, whereas RS4;11 were negative (Figure 1b). To add more mechanistic insight, NG2 expression was also analyzed at the RNA level by reverse transcriptase-PCR. As shown in Figure 1c, NG2 mRNA is present in all the NG2 + leukemic cell lines and absent in all the NG2 – leukemic cell lines, indicating a full correlation between NG2 mRNA detection and NG2 transcribed protein.

Our experimental data support the clinical finding of both primary human leukemias with balanced MLL rearrangements coexpressing NG2 and human leukemia cases harboring balanced MLL gene translocations but lacking NG2 expression (Supplementary Table 1). This data not only highlight the lack of association between NG2 antigen and MLL rearrangements but also provide preliminary experimental evidence against the hypothesis that NG2 expression in human leukemia may be dependent on the particular gene(s) paired to MLL when rearranged, as we demonstrate that two pro-B ALL cell lines (MV4;11 and RS4;11) with the MLL gene fused to the same partner, AF4, have a completely different NG2 antigen expression profile (Figure 1).

Several cellular and molecular mechanisms, intrinsic molecular determinants and extrinsic signals may contribute to the controversial correlation between MLL rearrangements and NG2 regulation. The confused association observed in actual leukemic samples makes an explanation based on a potential role for '*in vivo*' (bone marrow, CB and so on) occurring cell–cell interactions unlikely. Similarly, our '*in vitro*' data, rule out the possibility that potential hits/mutations secondary to MLL translocations are required for triggering NG2 expression, as we used fully transformed/immortalized

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cell lines derived from patients with overt disease, therefore carrying a paramount of cooperating mutations and genetic insults. The possibility that NG2 expression could be associated with nonbalanced MLL rearrangements such as deletions or inversions has previously been ruled out.^{4,5} Although unlikely, the possibility that NG2 expression could be linked to MLL internal duplications, which are not prospectively analyzed in human leukemias at diagnosis, should not be excluded.

Here, we hypothesize that NG2 expression may be dependent on the cell of origin where a specific leukemic abnormality initially occurs. For instance, NG2 might only be regulated when the leukemic abnormality arises either in a lineagespecific progenitor (hematopoietic progenitor cell; HPC) or in a more immature, less committed stem cell (hematopoietic stem cell; HSC). When a large number $(2 \times 10^5 \text{ cells})$ of CD34 + hematopoietic stem/progenitor cells are gated and analyzed by flow cytometry, coexpression of NG2 is readily observed in a subset of CD34 + CD38 + HPCs from CB (2.1 ± 2.4%; Figure 2a), bone marrow $(0.83 \pm 2.2\%)$ and mobilized peripheral blood (1.3%; Supplementary Figure 1a), suggesting that HPCs rather than HSCs may be the target cell for transformation. To verify that this CD34+CD38+NG2+ cell subset represents truly HPCs, this population was enriched (74% post-sort purity) by flurescence-activated cell sorting from CB and the cells plated in methylcellulose assays. Importantly, multilineage (CFU-G, CFU-M; CFU-Mix, BFU-E) hematopoietic colonies were obtained in *in vitro* colony-forming unit (CFU) assays¹⁸ (Supplementary Figure 1a), suggesting that HPCs rather than HSCs may be the target cell for transformation. Recent studies from Vormoor laboratory^{19,20} support our data as they found that NG2 is not expressed in MLL-rearranged HSC populations (CD34+CD38-) but was up-regulated in differentiated MLL-

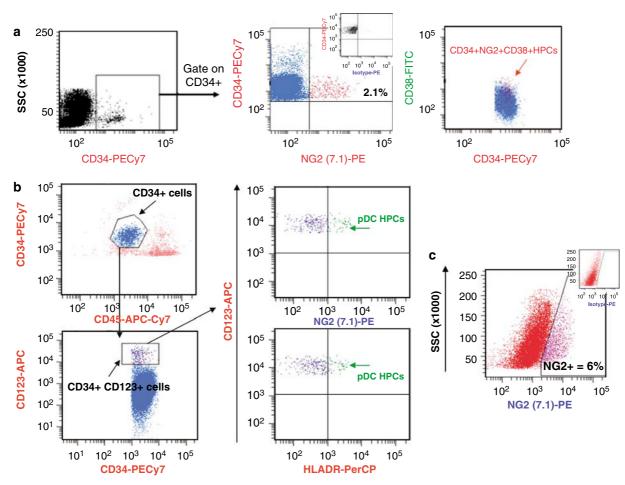


Figure 2 Representative flow cytometry analysis of NG2 (7.1) antigen in normal CD34 + progenitors, CD34 + pDC precursors and hESCs. (a) Expression of NG2 in gated CB-derived CD34 + cells. Analyses of as many as 2×10^5 CD34 + cells revealed that $2.1 \pm 2.4\%$ of the CD34 + cells coexpress NG2 + cells. All these CD34 + NG2 + cells are CD38 + , therefore, representing HPCs. An irrelevant isotype-matched antibody was used as a negative control (inset panel). (b) Six-color high-speed flow cytometry analysis showing the expression of NG2 (green dots) in a population of pDC precursors (CD45 + CD34 + CD38 + CD123 + + and HLADR +).¹⁶ (c) Representative NG2 expression in genetically stable hESCs. An irrelevant isotype-matched antibody was used as a negative control (inset panel). CB, cord blood; HPCs, hematopoietic progenitor cells; NG2, neuron-glial antigen 2; pDC, plasmacytoid dendritic cell.

rearranged leukemic blasts (CD19+ B-lineage blasts and CD33+ myeloid blasts) in pediatric MLL-rearranged AMLs and MLL-AF4+ ALLs.

The expression of NG2 in 60% of pDC leukemias lacking MLL rearrangements have recently been reported¹¹ (our unpublished observations). We, therefore, next addressed whether pDC $CD34 + precursors^{16}$ coexpress NG2. Interestingly, we found that $12.2 \pm 13\%$ of the pDC precursors (CD34⁺/CD45⁺/ CD38⁺/CD123^{high}/HLADR⁺) readily coexpress NG₂ (Figure 2b). This suggests that, regardless of the status of the MLL locus, the NG2 antigen may be expressed in pDC leukemias, if the leukomogenesis process is initially triggered in a pDC CD34 + precursor readily expressing NG2, which might act as a leukemic-initiating cell. In fact, based on the existence of NG2 + cell lines and NG2 + human primary leukemias ¹¹(our unpublished observations) lacking MLL rearrangements our data illustrate that the leukemic abnormality underlying NG2 expression does not necessarily need to be an MLL rearrangement.

Unlike other MLL fusion proteins, MLL-AF4 fusion gene resulting from the t(4;11) is always found in infant pro-B-ALL with a dismal prognosis and arises prenatally.¹⁷ From an ontogeny standpoint, two possible scenarios have been proposed: (i) MLL-AF4 arises and has its preleukemic impact during

early human embryonic hematopoiesis or (ii) MLL-AF4 arises during early hematopoiesis but has its preleukemic impact only in later fetal hematopoiesis.¹⁷ We analyzed the expression of NG2 in undifferentiated hESCs, a potential prenatal target cell for MLL-AF4 occurrence. Intriguingly, NG2 expression is also readily found in 5–6% of genetically stable hESCs (Figure 2d).

Despite NG2 and its human homolog was first reported on oligodendrocyte progenitor cells, the possibility that the cell subset CD34 + NG2 + might represent a potential circulating neural progenitors should be ruled out since, as shown by sixcolor high-speed flow cytometry, the NG2-expressing cells coexpress the panhematopoietic marker CD45 and the HPCs markers CD34 and CD38. More importantly, NG2 expression in CB was confined to the CD45 + CD34 + cell subset, indicating that, NG2 is solely expressed in hematopoietic cells, especially in committed HPCs; pDC-progenitors (CD45+ CD34+ CD38 + CD123 + + and $HLADR +)^{16}$ among others, in line with the occurrence of NG2 + pDC-leukemias lacking MLL.¹¹ Taken together, we throw more light on the association between NG2 expression and MLL rearrangements and report preliminary data supporting that NG2 expression does not depend on the particular gene(s) paired to MLL. The fact that CD34+ progenitors, pDC precursors and hESCs express NG2 indicates that NG2 in leukemia might uniquely be expressed in those 1477

patients where the leukemogenesis process is triggered in a NG2-expressing cell subset, regardless the type of leukemia. More comprehensive clinical, functional, molecular, cytogenetic and immunophenotypic studies are warranted to resolve the current puzzling and confusing significance of NG2 expression in human leukemias.

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Author contribution:

CB and RM designed and perfomed experiments and analyzed data. LM performed and analyzed experiments. IP and MCH provided CB samples and clinical data. AO contributed key clinical data, analyzed data and revised the manuscript. PM conceived, supported and supervised the work and analyzed the data. CB and PM wrote the paper.

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Supplementary Information accompanies the paper on the Leukemia website (http://www.nature.com/leu)

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