

1 **Solar treatment (H<sub>2</sub>O<sub>2</sub>, TiO<sub>2</sub>-P25 and GO-TiO<sub>2</sub> photocatalysis, photo-Fenton)**  
2 **of organic micropollutants, human pathogen indicators, antibiotic resistant**  
3 **bacteria and related genes in urban wastewater**

4 Nuno F.F. Moreira<sup>1,2</sup>, Carlos Narciso-da-Rocha<sup>3</sup>, M. Inmaculada Polo-López<sup>4</sup>, Luisa M. Pastrana-  
5 Martínez<sup>1</sup>, Joaquim L. Faria<sup>1</sup>, Célia M. Manaia<sup>3</sup>, Pilar Fernández-Ibáñez<sup>4,5,\*</sup>, Olga C. Nunes<sup>2</sup>,  
6 Adrián M.T. Silva<sup>1,\*</sup>

7  
8 <sup>1</sup>*Laboratory of Separation and Reaction Engineering - Laboratory of Catalysis and Materials*  
9 *(LSRE-LCM), Faculdade de Engenharia, Universidade do Porto, Rua Dr. Roberto Frias, 4200-*  
10 *465 Porto, Portugal*

11 <sup>2</sup>*LEPABE – Laboratory for Process Engineering, Environment, Biotechnology and Energy,*  
12 *Faculdade de Engenharia, Universidade do Porto, Rua Dr. Roberto Frias, 4200-465 Porto,*  
13 *Portugal*

14 <sup>3</sup>*Universidade Católica Portuguesa, CBQF - Centro de Biotecnologia e Química Fina –*  
15 *Laboratório Associado, Escola Superior de Biotecnologia, Rua Arquiteto Lobão Vital, Apartado*  
16 *2511, 4202-401 Porto, Portugal*

17 <sup>4</sup>*Plataforma Solar de Almeria – CIEMAT, P.O. Box 22, 04200 Tabernas, Almeria, Spain*

18 <sup>5</sup>*Nanotechnology and Integrated BioEngineering Centre, School of Engineering, University of*  
19 *Ulster, Newtownabbey, Northern Ireland, BT37 0QB, United Kingdom*

20  
21 \*Corresponding authors e-mail addresses:

22 [adrian@fe.up.pt](mailto:adrian@fe.up.pt) (A.M.T. Silva); [p.fernandez@ulster.ac.uk](mailto:p.fernandez@ulster.ac.uk) (P. Fernández-Ibáñez).

23  
24 **Abstract**

25 Solar-driven advanced oxidation processes were studied in a pilot-scale photoreactor, as tertiary  
26 treatments of effluents from an urban wastewater treatment plant. Solar-H<sub>2</sub>O<sub>2</sub>, heterogeneous

27 photocatalysis (with and/or without the addition of H<sub>2</sub>O<sub>2</sub> and employing three different  
28 photocatalysts) and the photo-Fenton process were investigated. Chemical (sulfamethoxazole,  
29 carbamazepine, and diclofenac) and biological contaminants (human pathogen indicators, their  
30 antibiotic resistant counterparts, 16S rRNA and antibiotic resistance genes), as well as the whole  
31 bacterial community, were characterized.

32 Heterogeneous photocatalysis using TiO<sub>2</sub>-P25 and assisted with H<sub>2</sub>O<sub>2</sub> (P25/H<sub>2</sub>O<sub>2</sub>) was the most  
33 efficient process on the degradation of the chemical organic micropollutants, attaining levels below  
34 the limits of quantification in less than 4 hours of treatment (corresponding to Q<sub>UV</sub> < 40 kJ L<sup>-1</sup>).  
35 This performance was followed by the same process without H<sub>2</sub>O<sub>2</sub>, using TiO<sub>2</sub>-P25 or a composite  
36 material based on graphene oxide and TiO<sub>2</sub>.

37 Regarding the biological indicators, total faecal coliforms and enterococci and their antibiotic  
38 resistant (tetracycline and ciprofloxacin) counterparts were reduced to values close, or beneath, the  
39 detection limit (1 CFU 100 mL<sup>-1</sup>) for all treatments employing H<sub>2</sub>O<sub>2</sub>, even upon storage of the  
40 treated wastewater for 3-days. Moreover, P25/H<sub>2</sub>O<sub>2</sub> and solar-H<sub>2</sub>O<sub>2</sub> were the most efficient  
41 processes in the reduction of the abundance (gene copy number per volume of wastewater) of the  
42 analysed genes. However, this reduction was transient for 16S rRNA, *intI1* and *sulI* genes, since  
43 after 3-days storage of the treated wastewater their abundance increased to values close to pre-  
44 treatment levels. Similar behaviour was observed for the genes *qnrS* (using TiO<sub>2</sub>-P25), *bla<sub>CTX-M</sub>*  
45 and *bla<sub>TEM</sub>* (using TiO<sub>2</sub>-P25 and TiO<sub>2</sub>-P25/H<sub>2</sub>O<sub>2</sub>). Interestingly, higher proportions of sequence  
46 reads affiliated to the phylum *Proteobacteria* (*Beta*- and *Gammaproteobacteria*) were found after  
47 3-days storage of treated wastewater than before its treatment. Members of the genera  
48 *Pseudomonas*, *Rheinheimera* and *Methylothera* were among those with overgrowth.

49  
50 **Keywords:** Solar advanced oxidation processes; urban wastewater; human pathogen indicators;  
51 antibiotic resistance genes; bacterial community composition.

52

## 53 **1. Introduction**

54 Conventional urban wastewater treatment plants (UWWTPs) are not specifically designed for the  
55 removal of organic micropollutants, neither for disinfection which considers the inactivation of  
56 bacteria that can contribute to the spread of antibiotic resistant bacteria (ARB) and antibiotic  
57 resistant genes (ARG) into the environment. These contaminants can reach the natural waters, such  
58 as surface and ground waters that are serving as drinking water sources (Fatta-Kassinos et al. 2011,  
59 Manaia et al. 2016). Moreover, the continuous disposal of antibiotics and related products into the  
60 environment can lead to the development and proliferation of ARB, decreasing the efficiency of  
61 these antibiotics when supplied to human and animals (Rizzo et al. 2013, Ferro et al. 2016). Since  
62 high bacterial density can be found in effluents of UWWTPs (i.e., bacterial cells are close to each  
63 other), horizontal gene transfer and selection of ARB can be considered important mechanisms for  
64 ARG enrichment (Davison 1999). The dissemination of these contaminants urges the development  
65 of new technologies able to improve the simultaneous removal of organic micropollutants and  
66 microorganisms of concern.

67 Advanced oxidation processes (AOPs) are conceptually based on the generation of the highly  
68 reactive hydroxyl radicals ( $\text{HO}^\bullet$ ), but other reactive species can also be formed. AOPs have been  
69 widely studied, mainly with synthetic matrices, and they are seen as viable solutions for the removal  
70 of organic micropollutants from urban wastewaters (Ribeiro et al. 2015). Additionally, the formed  
71 radicals can also act as disinfectant species leading to effective reduction of high bacterial loads  
72 (Moreira et al. 2016, Malato et al. 2009, Sousa et al. 2017, Polo-López et al. 2014, Pablos et al.  
73 2013, Yang et al. 2014). The operating cost of AOPs is one of its main drawbacks holding-off their  
74 application. In the case of photo-driven AOPs, sunlight technologies are being developed in an  
75 effort to decrease the treatment cost. For instance, solar disinfection showed high efficiency on the  
76 reduction of microbial loads, and its efficiency can be increased by the addition of photocatalysts  
77 or some chemical agents (Dunlop et al. 2010, Ferro et al. 2015, Fiorentino et al. 2015). In a recent  
78 study (Becerra-Castro et al. 2016), we have shown that oxidation processes (such as  $\text{UV}_{254 \text{ nm}}$ ,

79 ozonation and photocatalytic ozonation) might have the potential to act selectively over some  
80 bacterial groups. In that bench-scale study, irrespective of the type of treatment used, after 3-days  
81 storage the bacterial community was characterized by higher proportions of *Proteobacteria*  
82 (*Gamma*- and *Betaproteobacteria*) than those observed in non-treated wastewater. This is an  
83 example of bacterial community disturbance induced by disinfection, which may affect negatively  
84 the biological quality of the final stream.

85 The present study aims at comparing the efficiency of different solar-driven AOPs on the removal  
86 of micropollutants and disinfection of a secondary effluent of an UWWTP. Solar-H<sub>2</sub>O<sub>2</sub>,  
87 heterogeneous photocatalysis (with and/or without the addition of H<sub>2</sub>O<sub>2</sub>) and the photo-Fenton  
88 process were tested using a pilot-plant compound parabolic collector (CPC) solar photoreactor. For  
89 the first time, the performance of each process was assessed based on the removal efficiency of  
90 organic contaminants and ARB&ARG, as well as on the changes of the bacterial community  
91 composition. Microbial characterization was performed before treatment, after 5 hours of treatment  
92 and after 3-days storage of treated wastewater at room temperature. Thus, the simultaneous removal  
93 of chemical and biological contaminants by using solar-driven AOPs at pilot-scale and considering  
94 possible changes in the bacterial community, which can affect the water quality, is the main novelty  
95 of the present work.

96

## 97 **2. Materials and methods**

### 98 *2.1 Chemicals and materials*

99 Degussa (Aeroxide) TiO<sub>2</sub>-P25 catalyst from Evonik Corporation (hereafter referred to as P25) and  
100 a composite consisting of TiO<sub>2</sub> and 4.0 wt.% of graphene oxide (GO-TiO<sub>2</sub>) – respective preparation  
101 method and characterization described elsewhere (Pastrana-Martínez et al. 2012) – were used to  
102 conduct heterogeneous photocatalytic experiments. For comparative purposes, bare TiO<sub>2</sub> was also  
103 prepared following the same method used for GO-TiO<sub>2</sub>, but without the addition of GO (hereafter  
104 referred to as TiO<sub>2</sub>). The H<sub>2</sub>O<sub>2</sub> 30% (w/v) solution, analytical grade sulphuric acid (H<sub>2</sub>SO<sub>4</sub>, 98%),

105 bovine liver catalase and ferrous sulphate heptahydrate ( $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$ ) were obtained from Riedel-  
106 de Haën (Germany), Merk (Germany), Sigma-Aldrich (USA) and PANREAC (Spain),  
107 respectively. Carbamazepine (CBZ), sulfamethoxazole (SMX) and diclofenac (DFC) were all high-  
108 purity grade (>99%), and purchased from Sigma-Aldrich (Spain). Stock solutions of each  
109 individual compound were dissolved at  $2.5 \text{ g L}^{-1}$  in methanol due to water solubility limitations.  
110 Required amounts of stock solutions were directly loaded into the CPC pilot reactor to obtain the  
111 initial concentration of  $100 \mu\text{g L}^{-1}$  of each organic micropollutant in urban wastewater samples.  
112 Methanol (J.T.Baker) and acetonitrile (Sigma-Aldrich) were HPLC-grade. Ultrapure water was  
113 supplied by a Milli-Q water system.

114

## 115 *2.2 Municipal wastewater treatment plant samples*

116 All solar-driven treatments were carried out using urban wastewater samples collected (every day  
117 in batches of 60 L) after the secondary treatment from the UWWTP of El Bobar (Almería, Spain),  
118 and stored at  $4 \text{ }^\circ\text{C}$  no more than 2 hours before solar experiments. The UWWTP uses conventional  
119 activated sludge plus decantation as secondary treatment. In 2015, it produced an average of  
120 secondary effluent daily flow of ca.  $33,000 \text{ m}^3$ , with a capacity of 100,000 inhabitant-equivalent.  
121 The main physicochemical characteristics of the effluent, including turbidity, pH, conductivity,  
122 dissolved inorganic carbon (DIC), dissolved organic carbon (DOC) and inorganic ions  
123 concentration, are listed in Table 1.

124

## 125 *2.3 Pilot scale CPC photoreactor and solar experiments*

126 All the solar-driven oxidation processes were performed in a pilot-scale CPC photoreactor, at  
127 Plataforma Solar de Almeria (PSA), Spain ( $37^\circ 84' \text{N}$  and  $2^\circ 34' \text{W}$ ), on sunny days between June  
128 and August 2015, with a duration of 5 hours. The configuration of the CPC photoreactor is  
129 described elsewhere (Rodríguez-Chueca et al. 2014); it allows the optimal collection of solar  
130 radiation (direct and diffuse) and high removal rates of chemical and biological contaminants

131 (Booshehri et al. 2017, Malato et al. 2013). The CPC photoreactor tube module, tilted at an angle  
 132 of 37° relative to the horizontal plane, is connected to a recirculation tank and a centrifugal pump.  
 133 The water flow rate was set at 10 L min<sup>-1</sup>. The total volume of the photoreactor was 20 L, while  
 134 the illuminated volume and the irradiated collector surface area were 15 L and 1 m<sup>2</sup>, respectively.  
 135 In heterogeneous photocatalysis (P25, TiO<sub>2</sub> and GO-TiO<sub>2</sub>) a catalyst load of 200 mg L<sup>-1</sup> was used.  
 136 In photo-Fenton (Fe<sup>2+</sup>/H<sub>2</sub>O<sub>2</sub>), ferrous sulphate heptahydrate was used as source of 10 mg L<sup>-1</sup> of  
 137 Fe<sup>2+</sup>. In H<sub>2</sub>O<sub>2</sub> assisted processes (H<sub>2</sub>O<sub>2</sub>, Fe<sup>2+</sup>/H<sub>2</sub>O<sub>2</sub>, P25/H<sub>2</sub>O<sub>2</sub> and GO-TiO<sub>2</sub>/H<sub>2</sub>O<sub>2</sub>), the initial  
 138 concentration of H<sub>2</sub>O<sub>2</sub> was 20 mg L<sup>-1</sup> reached by adding the H<sub>2</sub>O<sub>2</sub> 30% (w/v) solution. The catalyst  
 139 and H<sub>2</sub>O<sub>2</sub> concentrations were selected considering the optimization done in our previous studies  
 140 with solar-driven oxidation processes (Polo-López et al. 2014, Fernández-Ibáñez et al. 1999).  
 141 Immediately after the collection of a sample, residual H<sub>2</sub>O<sub>2</sub> was eliminated by adding a freshly  
 142 prepared solution of bovine liver catalase (0.1 g L<sup>-1</sup>) in a ratio 0.1/5.0 (v/v). Photolysis (Blank)  
 143 assays were performed to study the effect of solar radiation without the addition of any catalyst or  
 144 reactant.  
 145 Besides the experimental time, the accumulated solar UVA energy received in the solar reactor per  
 146 unit of treated water volume (Q<sub>UV</sub>, kJ L<sup>-1</sup>) was considered for comparison of the treatment  
 147 efficiencies (Malato et al. 2009) and calculated using Eq. (1):

$$148 \quad Q_{UV,n} = Q_{UV,n-1} + \frac{\Delta t_n \overline{UV_{G,n}} A_r}{V_t}; \Delta t_n = t_n - t_{n-1} \quad (1)$$

149 where  $UV_{G,n}$  is the global UV irradiance (W m<sup>-2</sup>) averaged during exposure time;  $t_n$  the exposure  
 150 time (s);  $V_t$  the total reactor volume (L); and  $A_r$  the illuminated reactor surface (m<sup>2</sup>).

151 A global UVA (300–400 nm, Model CUV4, Kipp & Zonen, Netherlands) pyranometer tilted 37°  
 152 was used to measure the solar radiant UVA rate incident (W m<sup>-2</sup>) as described elsewhere  
 153 (Rodríguez-Chueca et al. 2014). The average value of solar UVA irradiance was 40 W m<sup>-2</sup>.

154 Before treatment, the CPC photoreactor was covered by an opaque sheet while the wastewater and  
 155 reagents were added to the reactor and recirculated during 15 min to guarantee homogenisation.

156 Samples were collected each hour over the treatment. During this period, temperature (Checktemp,  
 157 Hanna instruments, Spain) and pH (multi 720, WTW, Germany) were measured. The temperature  
 158 of the wastewater varied from  $16.6 \pm 1.5$  °C to  $43.2 \pm 4.2$  °C.

159

160 **Table 1.** Chemical characterization of the secondary wastewater from the UWWTP. Values are the average  
 161 of four independent assays and errors represents standard deviations.

<b>Secondary effluent characterisation</b>				
Turbidity (NTU)	$5.3 \pm 3.1$		$\text{PO}_4^{3-}$ (mg L <sup>-1</sup> )	$12.4 \pm 6.1$
pH	$7.5 \pm 0.2$		$\text{SO}_4^{2-}$ (mg L <sup>-1</sup> )	$74.5 \pm 15.6$
Conductivity ( $\mu\text{S cm}^{-1}$ )	$1780 \pm 63$		$\text{Br}^-$ (mg L <sup>-1</sup> )	$2.7 \pm 0.7$
$\text{DIC}^\dagger$ (mg L <sup>-1</sup> )	$55 \pm 30$		$\text{Na}^+$ (mg L <sup>-1</sup> )	$199 \pm 6$
$\text{DOC}^\dagger$ (mg L <sup>-1</sup> )	$19 \pm 4$		$\text{Cl}^-$ (mg L <sup>-1</sup> )	$368 \pm 18$
$\text{NH}_4^+$ (mg L <sup>-1</sup> )	$35.3 \pm 25.5$		$\text{K}^+$ (mg L <sup>-1</sup> )	$25.6 \pm 1.8$
$\text{NO}_3^-$ (mg L <sup>-1</sup> )	$68.6 \pm 64.5$		$\text{Mg}^{2+}$ (mg L <sup>-1</sup> )	$35.3 \pm 2.6$
$\text{NO}_2^-$ (mg L <sup>-1</sup> )	$1.8 \pm 1.0$		$\text{Ca}^{2+}$ (mg L <sup>-1</sup> )	$74.9 \pm 3.2$

162 <sup>†</sup>DIC: dissolved inorganic carbon; <sup>†</sup>DOC: dissolved organic carbon.

163

#### 164 2.4 Chemical analysis

165 High performance liquid chromatography (HPLC) was used to analyze the evolution of the target  
 166 organic micropollutants using an apparatus from Agilent Technologies (series 1260, Palo Alto, CA,  
 167 USA), equipped with a diode array detector (UV-DAD) and a C-18 column. The mobile phase  
 168 consisted of 90% formic acid aqueous solution at 25 mM and 10% acetonitrile. A linear gradient  
 169 was used from 10% to 85% of acetonitrile during 13 min at a flow rate of 1 mL min<sup>-1</sup>. The injection  
 170 volume was set at 100  $\mu\text{L}$ . Before injection, samples were filtered through a 0.2  $\mu\text{m}$  syringe-driven  
 171 filter, and afterwards the filter was washed with 1 mL of acetonitrile to remove the adsorbed  
 172 compounds. CBZ, SMX and DFC were selected as model organic micropollutants since they have  
 173 been frequently found in aquatic environments, including wastewater, surface and groundwater and

174 even in drinking water (Barbosa et al. 2016). The detection wavelength values were 267 nm for  
175 both CBZ and SMX, and 273 nm for DFC. The limits of quantification (LOQ) were 4.7, 6.2 and  
176 4.1  $\mu\text{g L}^{-1}$  for CBZ, SMX and DFC, respectively.

177 DX-600 and DX-120 (Dionex Corporation, Sunnyvale, CA) equipments were used for  
178 quantification of anions and cations, respectively. DOC and DIC measurements were performed  
179 using a 5050 A TOC (Shimadzu Corporation, Kyoto, Japan) analyzer, after sample filtration using  
180 0.2  $\mu\text{m}$  syringe filters. Turbidity was determined with a turbidimeter (Model 2100N, Hach, USA).  
181 The  $\text{H}_2\text{O}_2$  concentration was determined by a spectrometric method, as described elsewhere (Polo-  
182 López et al. 2011), and the  $\text{Fe}^{2+}$  concentration by the ISO 6332 method. Natural iron was not  
183 detected in any of the urban wastewater samples by using this method, i.e. a spectrophotometric  
184 technique with phenanthroline/acetic acid (UV–Vis measurements, limit of detection 0.05  $\text{mg L}^{-1}$ ).

185

### 186 *2.5 Microbiological cultivation, DNA extraction, qPCR and bacterial community analysis*

187 To assess the disinfection efficiency, cultivable indicator bacteria, targeted ARB and selected  
188 ARGs were quantified before and immediately after the 5 hours-treatment, and after 3-days of  
189 storage of treated wastewater at room temperature. The abundance of faecal coliforms, enterococci  
190 and their tetracycline and ciprofloxacin resistant counterparts was assessed by using the membrane  
191 filtration method. After serial 10-fold dilutions in sterile saline solution (0.85% NaCl), 100 mL of  
192 each dilution was filtered through cellulose membrane filters (0.22  $\mu\text{m}$  porosity; Whatman, UK).  
193 The filtering membranes were incubated on selective media for each target bacterial groups -  
194 membrane Faecal Coliforms agar (m-FC) (Difco, 30 °C, 24 hours) for faecal coliforms, and Slanetz  
195 & Bartley agar (S&B) (Difco, 30 °C, 48 hours) for enterococci. In addition, the prevalence of ARB  
196 was assessed in the same media supplemented with tetracycline (Fluka, 16  $\text{mg L}^{-1}$ ) and  
197 ciprofloxacin (Applichem, 1  $\text{mg L}^{-1}$ ). The antibiotic stock solutions were sterilized by filtration  
198 (0.2  $\mu\text{m}$  syringe driven-filters). For culture-independent assays, a volume of 250 mL of each sample  
199 (before and after the 5 hours treatment, and after 3-days of storage) was filtered through



200 polycarbonate membranes (0.22  $\mu\text{m}$  porosity; Whatman, UK). DNA was extracted using the  
 201 commercial kit PowerWater® DNA Isolation (MO BIO Laboratories, Inc., USA) and stored at -20  
 202 °C. These extracts were used for quantitative PCR (qPCR) of samples collected before treatment,  
 203 immediately after treatment, and after 3-days of storage, and also for the bacterial community  
 204 analysis, in this case except for the samples collected immediately after treatment, due to DNA  
 205 scarcity. The qPCR (StepOne™ Real-Time PCR System; Life Technologies, Carlsbad, CA) assays  
 206 were performed according to the conditions shown in Table 2, as described elsewhere (Narciso-da-  
 207 Rocha et al. 2014).

208

209 **Table 2.** Target genes and conditions used in qPCR assays.

Target gene	Primers (sequence) Reference	Conditions	Efficiency (%)	Reference
16S rRNA	1114F (CGGCAACGAGCGCAACCC) 1275R (CCATTGTAGCACGTGTGTAGCC) <i>Escherichia coli</i> - ATCC 25922	95 °C for 10 min (1 cycle) 95 °C for 15 s, 55 °C for 20 s and 72 °C for 10 s (35 cycles)	100	(Denman and McSweeney 2006)
<i>bla</i> <sub>TEM</sub>	<i>bla</i> <sub>TEM</sub> -F (TTCCTGTTTTTGCTCACCCAG) <i>bla</i> <sub>TEM</sub> -R (CTCAAGGATCTTACCGCTGTTG) <i>Escherichia coli</i> - A2FCC14	95 °C for 10 min (1 cycle) 95 °C for 15 s, 60 °C for 30 s and 72 °C for 10 s (40 cycles)	96	(Bibbal et al. 2007)
<i>int</i> <sub>I1</sub>	<i>int</i> <sub>I1</sub> -F (CCTCCCGCACGATGATC) <i>int</i> <sub>I1</sub> -R (TCCACGCATCGTCAGGC) <i>Escherichia coli</i> - A2FCC14	95 °C for 10 min (1 cycle) 95 °C for 15 s, 55 °C for 30 s and 72 °C for 10 s (40 cycles)	94	(Goldstein et al. 2001)
<i>qnr</i> <sub>S</sub>	<i>qnr</i> <sub>S</sub> rF11 (GACGTGCTAACTTGCGTGAT) <i>qnr</i> <sub>S</sub> rR11 (TGGCATTGTTGGAAACTTG) <i>Enterobacter cloacae</i> - S1+	95 °C for 5 min (1 cycle) 95 °C for 15 s and 60 °C for 1 min (40 cycles)	95	(Marti and Balcázar 2013)
<i>sul</i> <sub>1</sub>	<i>sul</i> <sub>1</sub> -FW (CGCACCGGAAACATCGCTGCAC) <i>sul</i> <sub>1</sub> -RV (TGAAGTTCCGCCGAAGGCTCG) <i>Achromobacter sp.</i>	95 °C for 5 min (1 cycle) 95 °C for 15 s and 60 °C for 1 min (40 cycles)	94	(Pei et al. 2006)
<i>van</i> <sub>A</sub>	<i>van</i> <sub>A</sub> 3FP (CTGTGAGGTCGGTTGTGCG) <i>van</i> <sub>A</sub> 3RP (TTTGGTCCACCTCGCCA) <i>Enterococcus faecalis</i> - H1EV23	95 °C for 5 min (1 cycle) 95 °C for 15 s and 60 °C for 1 min (40 cycles)	98	(Volkman et al. 2004)
<i>bla</i> <sub>CTX-M</sub>	<i>CTXM</i> -FW (CTATGGCACCACCAACGATA) <i>CTXM</i> -RV (ACGGCTTTCTGCCTTAGGTT) <i>Escherichia coli</i> - A2FC14	95 °C for 10 min (1 cycle), 95 °C for 15 s and 60 °C for 1 min (40 cycles)	94	(Marti et al. 2014)

210

211 The bacterial community composition was analysed based on the hypervariable V3/V4 region  
212 (forward primer Bakt\_341F 5'-CCTACGGGNGGCWGCAG-3' and reverse Bakt\_805R 5'-  
213 GACTACHVGGGTATCTAATCC-3') of 16S rRNA gene Illumina sequencing (Genoinseq,  
214 Cantanhede, Portugal). Nucleotide sequencing data were processed and analyzed using the QIIME  
215 pipeline (Caporaso et al. 2010a). Briefly, sequences shorter than 250 bp and with average quality  
216 scores lower than 25 were eliminated, and bases with average quality lower than 25 in a window  
217 of 5 bases, were trimmed using the software PRINSEQ (Schmieder and Edwards 2011). Chimeric  
218 sequences were identified and removed using USEARCH v6.1 (Edgar 2010). Free-chimeric  
219 sequences were further grouped into operational taxonomic units (OTUs) using USEARCH v6.1  
220 (Edgar 2010) with a phylotype threshold of  $\geq 97\%$  sequence similarity and were taxonomically  
221 assigned using QIIME default values. The sequences comprising each OTU were aligned using  
222 PyNAST (Caporaso et al. 2010b) and were taxonomically classified using Greengenes Database  
223 version 13\_8 (updated: August 2013) (DeSantis *et al.* 2006). As a variable number of sequences  
224 was obtained between samples, the alpha diversity indices Shannon, phylogenetic diversity (PD)  
225 whole tree, and Simpson were calculated after rarefying to 54,771 sequences *per* sample (value of  
226 the smallest dataset) (Shannon and Weaver 1963, Simpson 1949, Faith 1992). The cumulative sum  
227 scaling (CSS) normalization procedure was applied to the sequence data to assess the beta diversity  
228 patterns (Paulson et al. 2013). The weighted UniFrac metric distances (Lozupone and Knight 2005)  
229 were calculated in the QIIME pipeline and the results shown as Principal Coordinates Analysis  
230 (PCoA) biplots that include the position of the ten most prevalent bacterial classes. Correlations  
231 between the relative abundance of populations at different taxonomic levels were analyzed using  
232 the statistics software STAMP v2.1.3 (Parks et al. 2014).

233

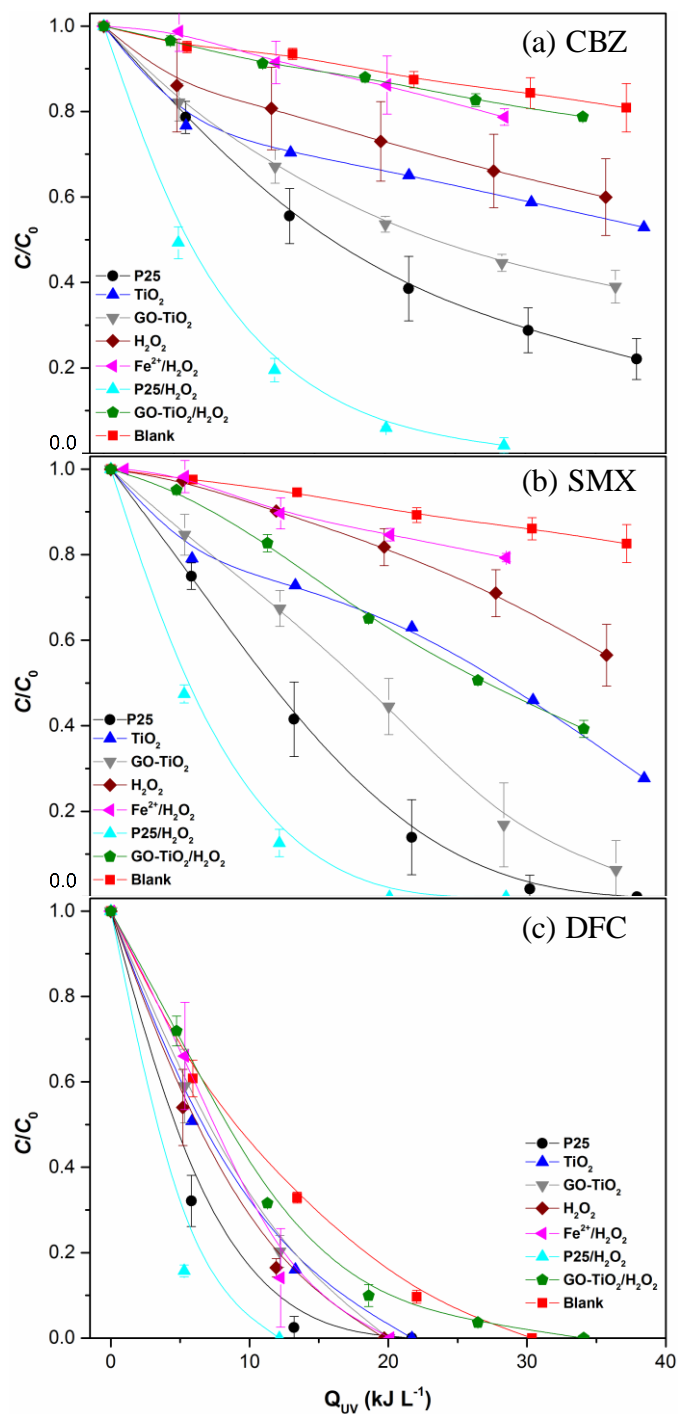
### 234 **3. Results and discussion**

#### 235 *3.1 Degradation of organic micropollutants*

236 The evolution of the concentrations of targeted organic micropollutants (i.e., CBZ, SMX and DFC)  
237 spiked in urban wastewater under solar-driven oxidation treatments are shown in Fig. 1. Control  
238 experiments were performed under the same conditions but without the addition of any catalyst or  
239 H<sub>2</sub>O<sub>2</sub> (i.e., Blank = photolysis). CBZ and SMX were very resistant upon irradiation in the absence  
240 of a catalyst or H<sub>2</sub>O<sub>2</sub> (only 20 ± 6% and 17 ± 4% of removal, respectively). In contrast, DFC was  
241 efficiently removed by photolysis after 4 hours. This result was expected since the DFC absorption  
242 spectrum (not shown) has a tail entering well above the 300 nm (Moreira et al. 2015).

243 The heterogeneous photocatalysts (P25, TiO<sub>2</sub> and GO-TiO<sub>2</sub>), without the addition of H<sub>2</sub>O<sub>2</sub>,  
244 converted the targeted organic micropollutants in the following order of decreasing efficiency: P25  
245 > GO-TiO<sub>2</sub> > TiO<sub>2</sub> (Fig. 1). The high photocatalytic efficiency of GO-TiO<sub>2</sub> on the degradation of  
246 different types of organic micropollutants in synthetic matrices has been already demonstrated in  
247 our previous studies, namely for: diuron, alachlor, isoproturon and atrazine – pesticides classified  
248 by the EU as priority pollutants (Cruz et al. 2017); microcystin-LA – cyanotoxin (Sampaio et al.  
249 2015); bisphenol A – xenoestrogen (Maroga Mboula et al. 2013); diphenhydramine – antihistamine  
250 pharmaceutical, and methyl orange – azo dye (Pastrana-Martínez et al. 2012). The GO-TiO<sub>2</sub>  
251 photocatalyst has been much more active than P25 under Vis-light illumination, but under near  
252 UV-Vis radiation the activity of this type of photocatalysts depends on the target pollutant. The  
253 better performance of GO-TiO<sub>2</sub> in comparison to bare TiO<sub>2</sub> has been attributed to the good  
254 assembly and interfacial coupling between TiO<sub>2</sub> and GO sheets as well as the respective quenching  
255 of photoluminescence (inhibiting charge recombination) (Pastrana-Martínez et al. 2012). In the  
256 present study, P25 was the most efficient photocatalyst for the degradation of the organic  
257 micropollutants, most probably, because of the large fraction of UV radiation entering the photo-  
258 reactor by the CPC mirror configuration.

259



260

261 **Figure 1.** Normalized concentration ( $C/C_0$ ) of (a) CBZ, (b) SMX and (c) DFC spiked in urban wastewater  
 262 as function of accumulated energy ( $Q_{UV}$ ) using different solar-driven treatments. Except for  $TiO_2$ , values are  
 263 the average of four (P25), three ( $GO-TiO_2$ ,  $H_2O_2$ ,  $Fe^{2+}/H_2O_2$ ) and two ( $P25/H_2O_2$ ,  $GO-TiO_2/H_2O_2$ , Blank)  
 264 independent assays. Error bars represent standard deviations.

265

266 The addition of  $H_2O_2$  to the P25 photocatalytic system, significantly increased the degradation  
 267 efficiency of the targeted micropollutants.  $H_2O_2$  captures and reacts with the photoinduced surface

268 electrons (suppressing the electron/hole recombination) and it also reacts with the superoxide radical  
269 anions, both pathways leading to the formation of additional hydroxyl radicals (Pablos et al. 2013,  
270 Kositzi et al. 2004). There is markedly a competing process, which may be mediated by the active  
271 surface of the photocatalyst. Interestingly, the removal efficiency decreased when H<sub>2</sub>O<sub>2</sub> was added  
272 to the photocatalytic system containing GO-TiO<sub>2</sub>. Degradation of GO-TiO<sub>2</sub> may eventually occur,  
273 for instance by the H<sub>2</sub>O<sub>2</sub> attack to the underlying C-C bonds in the superficial defect sites of GO  
274 (Xing et al. 2014).

275 Solar-H<sub>2</sub>O<sub>2</sub> and photo-Fenton processes (Fe<sup>2+</sup>/H<sub>2</sub>O<sub>2</sub>) also led to very modest removals of CBZ and  
276 SMX (Figs. 1a and b, respectively). One of the downsides of photo-Fenton is the formation of iron  
277 sludge due to the precipitation of iron hydroxide at neutral pH. In this work, the pH was adjusted  
278 to a circumneutral value (5.5) by using H<sub>2</sub>SO<sub>4</sub>, which could explain the low photo-Fenton  
279 efficiency that is known to be maximized at pH values around 3 (Giannakis et al. 2017, Agulló-  
280 Barceló et al. 2013, García-Fernández et al. 2012).

281 Overall, P25/H<sub>2</sub>O<sub>2</sub> followed by the P25 and GO-TiO<sub>2</sub> photocatalytic processes were the best  
282 performing treatments for removal of the targeted organic micropollutants. Regarding the  
283 mineralization of the organic matter present in the urban wastewater, P25/H<sub>2</sub>O<sub>2</sub>, solar-H<sub>2</sub>O<sub>2</sub>, and  
284 the photo-Fenton process were the most efficient treatments (DOC removals always around 23 ±  
285 3%), other processes removing no more than ca. 10% of the initial DOC.

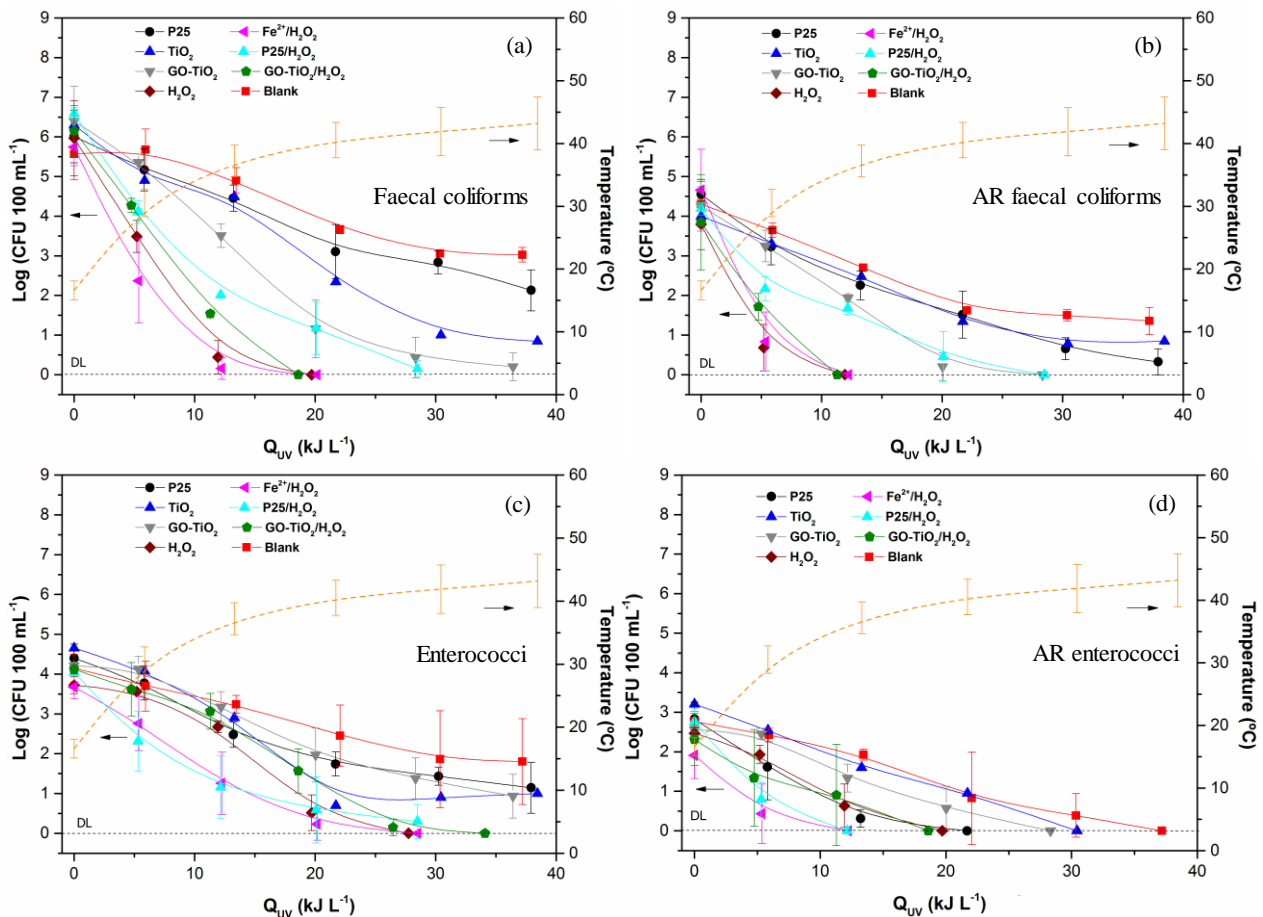
286

### 287 *3.2 Bacteria inactivation and reactivation*

288 The performance of different solar-driven processes was assessed before and over the treatment, in  
289 terms of removal of total faecal coliforms (Fig. 2a), enterococci (Fig. 2c) and respective fraction  
290 of resistant populations (Figs. 2b and d). The reduction of the bacterial indicators loads was  
291 observed in all the treatments, with the highest inactivation rates leading to values below or close  
292 the LOD (1 CFU 100 mL<sup>-1</sup>), for the processes where H<sub>2</sub>O<sub>2</sub> was used (H<sub>2</sub>O<sub>2</sub>, Fe<sup>2+</sup>/H<sub>2</sub>O<sub>2</sub>, P25/H<sub>2</sub>O<sub>2</sub>,  
293 GO-TiO<sub>2</sub>/ H<sub>2</sub>O<sub>2</sub>). Among these, and despite the iron precipitation, the photo-Fenton process was

294 the most efficient treatment on the reduction of resistant and non-resistant faecal coliforms and  
 295 enterococci (Figs. 2a-d, Fe<sup>2+</sup>/H<sub>2</sub>O<sub>2</sub>). However, photo-Fenton showed similar disinfection profiles  
 296 to solar-H<sub>2</sub>O<sub>2</sub> for faecal coliforms (Figs. 2a and b) and to P25/H<sub>2</sub>O<sub>2</sub> for enterococci (Figs. 2c and  
 297 d). These three processes removed those bacteria for Q<sub>UV</sub> < 30 kJ L<sup>-1</sup>, suggesting that H<sub>2</sub>O<sub>2</sub> plays  
 298 a major role on disinfection. In contrast, moderate inactivation rates were observed for photolysis  
 299 (Figs. 2a, b and c, Blank), except for antibiotic resistant enterococci that were reduced to values  
 300 below the LOD (Fig. 2d, Blank). Interestingly, P25/H<sub>2</sub>O<sub>2</sub> showed high efficiency on the removal  
 301 of organic micropollutants and resistant and non-resistant enterococci, whereas the efficiency for  
 302 inactivation of resistant and non-resistant faecal coliforms was not so high in comparison with  
 303 solar-H<sub>2</sub>O<sub>2</sub>.

304



305

306 **Figure 2.** Faecal coliforms (a) and enterococci (c) and their antibiotic resistant counterparts (b, d)  
 307 inactivation in urban wastewater as function of accumulated energy (Q<sub>UV</sub>) using different solar-driven  
 308 treatments. Except for TiO<sub>2</sub>, values are the average of four (P25), three (GO-TiO<sub>2</sub>, H<sub>2</sub>O<sub>2</sub>, Fe<sup>2+</sup>/H<sub>2</sub>O<sub>2</sub>) and two

309 (P25/H<sub>2</sub>O<sub>2</sub>, GO-TiO<sub>2</sub>/H<sub>2</sub>O<sub>2</sub>, Blank) independent assays. Error bars represent standard deviations. DL,  
310 detection limit.

311

312 The accepted hypotheses and mechanism that explain the inactivation of microorganisms by  
313 exposure to solar-H<sub>2</sub>O<sub>2</sub> is based on the accumulated damages inside cells by internal cellular injuries  
314 occurring under sunlight and accelerated in the presence of H<sub>2</sub>O<sub>2</sub>. It is well accepted that solar  
315 radiation produces internal damages affecting different intracellular vital components leading to  
316 bacterial death or lack of viability (Aguas et al. 2017). A recent study attributed bacterial  
317 inactivation during solar photolysis to the combined effect of intracellular production of reactive  
318 oxygen species by UV photons absorption and water temperature increase (Castro-Alfárez et al.  
319 2017). When H<sub>2</sub>O<sub>2</sub> is added, it may diffuse inside bacteria cells promoting additional internal  
320 photo-reactions with naturally present iron and other metals via Fenton and Fenton-like reactions,  
321 activating, thus, a photo-Fenton cycle under sunlight at intracellular level also (Aguas et al., 2017).  
322 Both photo-effects act jointly producing an accelerated disinfection that has been also reported to  
323 be very efficient for other types of bacteria, viral indicators, and fungi including, *Escherichia coli*  
324 and *Enterococcus faecalis* (Rodríguez-Chueca et al. 2014), *Legionella jordanis* (Polo-López et al.  
325 2017), F-specific RNA bacteriophage (Agulló-Barceló et al. 2013), *Fusarium* sp. (Polo-López et  
326 al. 2014, Sichel et al. 2009), *Phytophthora capsici* (Polo-López et al. 2013), *Curvularia* sp. (Aguas  
327 et al. 2017), and several antimicrobial resistant bacteria (Fiorentino et al. 2015).

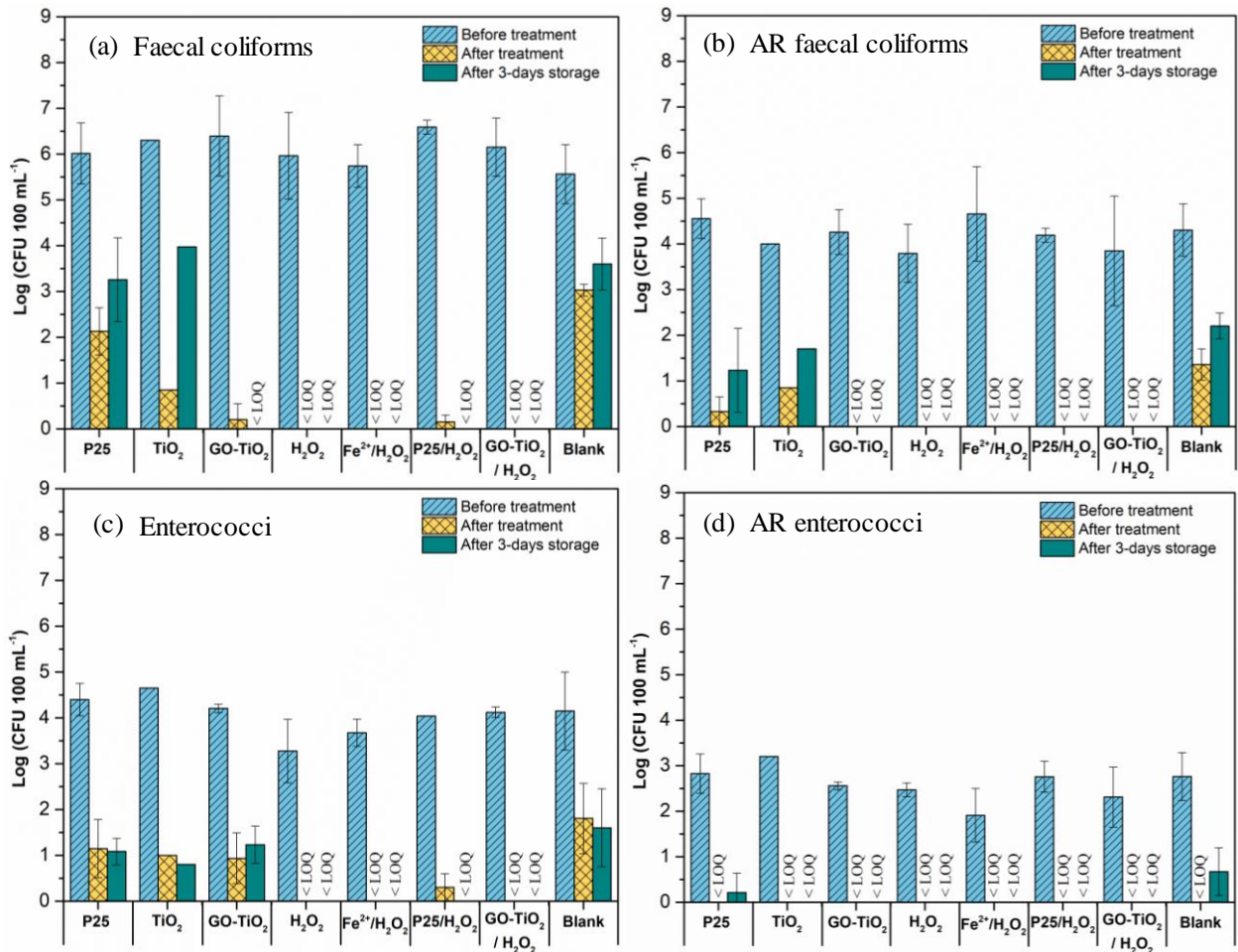
328 Although H<sub>2</sub>O<sub>2</sub> assisted processes performed better than non-assisted ones, heterogeneous  
329 photocatalysis without H<sub>2</sub>O<sub>2</sub> also performed quite well on the inactivation of total or resistant  
330 populations of faecal coliforms and enterococci. Among these processes, the GO-TiO<sub>2</sub> composite  
331 was the most efficient catalyst for the removal of total and resistant populations of faecal coliforms  
332 (Figs. 2a and b, GO-TiO<sub>2</sub>). The good efficiency of photocatalytic disinfection using this type of  
333 composites (but prepared by other methods) and under visible radiation only was already shown in  
334 previous studies (Cruz-Ortiz et al. 2017, Fernández-Ibáñez et al. 2015). Its high photocatalytic

335 activity has been attributed to the improvement in charge separation since GO may promote the  
336 electron transfer with TiO<sub>2</sub> particles, acting as an electron bridge, and to the decrease of the bandgap  
337 energy of the composite catalysts as well as to an enhancement of the adsorptive properties. These  
338 authors concluded that hydrogen peroxide, hydroxyl radicals, and singlet oxygen were the main  
339 species involved in the disinfection process under UV–Vis irradiation and only singlet oxygen  
340 under visible illumination (Cruz-Ortiz et al. 2017).

341 Since microbial inactivation, monitored via culture-based methods, can be a transient (Moreira et  
342 al. 2016, Zhao et al. 2014, Spuhler et al. 2010), further assays testing the regrowth capacity after  
343 3-days storage of the treated wastewater at room temperature were performed. Bacterial  
344 reactivation is influenced by factors such as the storage conditions, temperature, availability of  
345 nutrients and the UV dose, among others (Ubomba-Jaswa et al. 2009, Giannakis et al. 2014). The  
346 bacterial loads before, after the treatment and after 3-days storage at room temperature are shown  
347 in Figs. 3a and b for faecal coliforms and Figs. 3c and d for enterococci. No regrowth was observed  
348 in stored wastewater treated by the H<sub>2</sub>O<sub>2</sub>-assisted processes, and total faecal coliforms and  
349 enterococci as well as their ARB counterparts were kept below the detection limit. In stored  
350 wastewater treated by heterogeneous photocatalysis without H<sub>2</sub>O<sub>2</sub> (P25, TiO<sub>2</sub> and GO-TiO<sub>2</sub>), the  
351 bacterial loads of these groups were 2 to 3 log values lower than before the treatment. Similar  
352 observations were registered for the control (Blank = photolysis). These results indicate the  
353 inability of these bacterial groups to recover after the tested solar-driven processes. This can be  
354 also explained by the mode of action attributed to solar-H<sub>2</sub>O<sub>2</sub> disinfection, where oxidative  
355 damages alter the bacteria viability at intracellular level, as proven in the literature (Castro-Alfárez  
356 et al. 2017). In such report, an EMA-qPCR method for the detection of membrane integrity  
357 damages of *L. jordanis* cells under two photo-oxidative processes was used, i.e. solar-H<sub>2</sub>O<sub>2</sub> and  
358 P25 with solar radiation. It was confirmed the well-accepted mechanism of heterogeneous (P25)  
359 photocatalysis via oxidative attacks of the external cell membrane, whereas the mechanism for  
360 solar-H<sub>2</sub>O<sub>2</sub> was based on internal photochemical reactions. This may explain the results reported



361 in Fig. 3 on the non-recover capability of ARB when H<sub>2</sub>O<sub>2</sub> was used in these treatment processes,  
 362 while they can regrow when H<sub>2</sub>O<sub>2</sub> is not in the media (Castro-Alf rez et al. 2017).  
 363



364  
 365 **Figure 3.** (a) Faecal coliforms and (c) enterococci and their (b, d) antibiotic resistant counterparts counts  
 366 before, after treatment and after 3-days storage using different solar-driven treatments. Except for TiO<sub>2</sub>,  
 367 values are the average of four (P25), three (GO-TiO<sub>2</sub>, H<sub>2</sub>O<sub>2</sub>, Fe<sup>2+</sup>/H<sub>2</sub>O<sub>2</sub>) and two (P25/H<sub>2</sub>O<sub>2</sub>, GO-TiO<sub>2</sub>/H<sub>2</sub>O<sub>2</sub>,  
 368 Blank) independent assays. Error bars represent standard deviations.

369  
 370 *3.3 Effect of disinfection on ARGs and bacterial community composition*

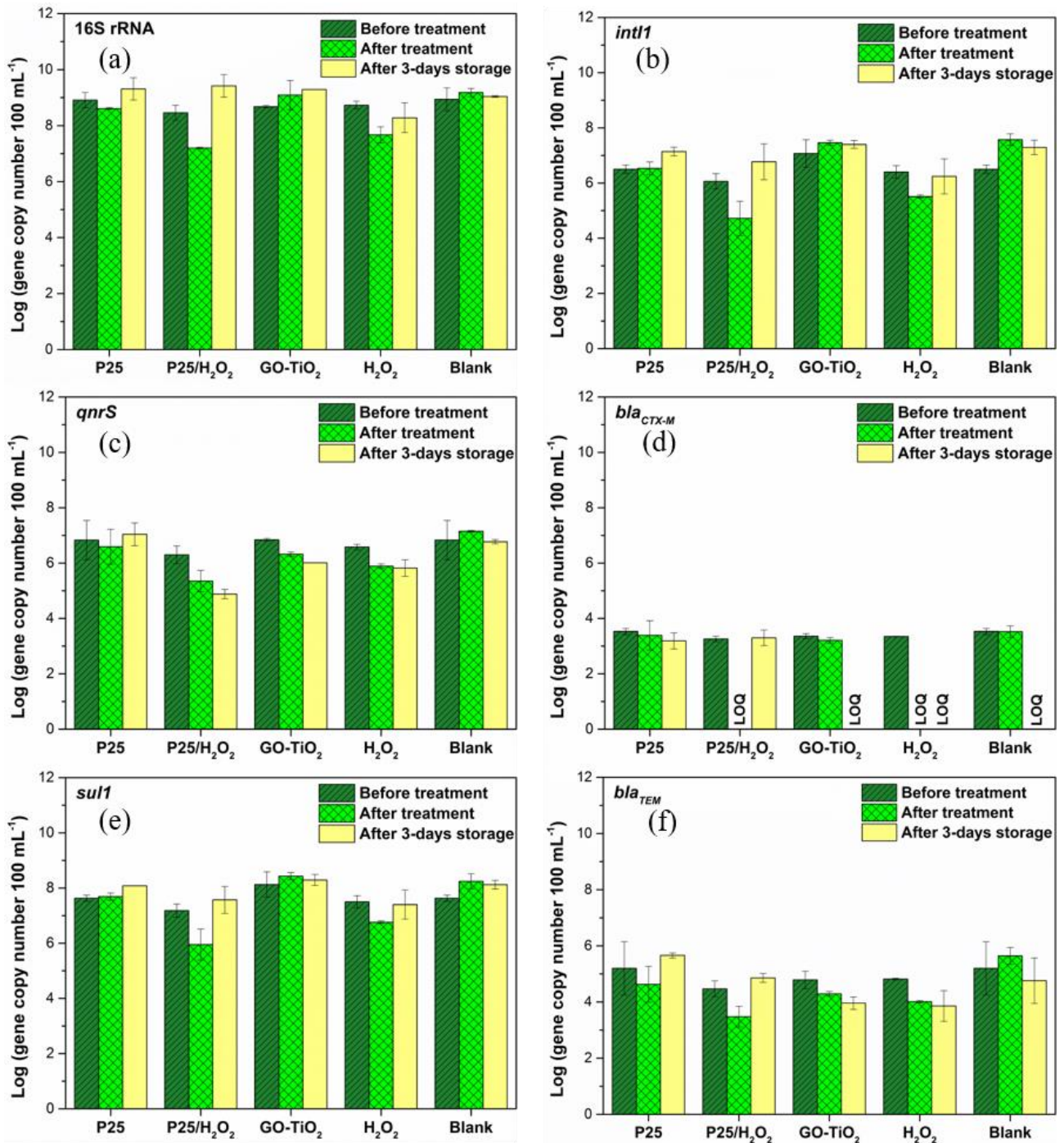
371 Considering that the bacterial community is much more diversified and complex than that assessed  
 372 based on the cultivation methods used, and that some bacteria may be injured and hence, unable to  
 373 grow, culture-independent methods were carried out to give additional insights on the disinfection  
 374 effectiveness of the studied solar-driven processes. Since the main purpose of this work was the

375 simultaneous removal of chemical and biological contaminants, the processes showing better  
376 performance on the degradation of organic micropollutants (i.e., P25/H<sub>2</sub>O<sub>2</sub>, P25 and GO-TiO<sub>2</sub>  
377 photocatalytic processes), disinfection (P25/H<sub>2</sub>O<sub>2</sub> and solar-H<sub>2</sub>O<sub>2</sub>), and also the reference process  
378 (photolysis), were selected for further investigation based on culture-independent methods -  
379 Fe<sup>2+</sup>/H<sub>2</sub>O<sub>2</sub> was not chosen due to its bad performance for degradation of the organic micropollutants  
380 (Fig. 1).

381 Among the analysed genes (i.e., 16S rRNA, *intI1*, *qnrS*, *bla<sub>CTX-M</sub>*, *sul1*, *bla<sub>TEM</sub>* and *vanA*), only  
382 *vanA* was below the LOD before treatment (not shown). For the other genes, P25/H<sub>2</sub>O<sub>2</sub>  
383 photocatalysis and solar-H<sub>2</sub>O<sub>2</sub> were the most efficient processes (i.e., lower abundance after  
384 treatment), both leading to log average reductions of 1-2 values (Fig. 4). However, after 3-days  
385 storage, regardless the treatment used, the abundance of 16S rRNA gene, a house keeping gene of  
386 prokaryotes, was close or even higher (up to 1 log for P25/H<sub>2</sub>O<sub>2</sub>) than those found before treatment,  
387 suggesting the ability of bacteria to recover after the treatment. Similar results were observed for  
388 *intI1* and *sul1* genes, encoding integrase and conferring resistance to sulphonamides, respectively.  
389 Among the studied processes, only solar-H<sub>2</sub>O<sub>2</sub> and GO-TiO<sub>2</sub> prevented the reactivation of *bla<sub>CTX-M</sub>*  
390 and *bla<sub>TEM</sub>* genes (encoding resistance to beta-lactams) above the pre-treatment levels. For the *qnrS*  
391 gene (encoding resistance to quinolones), besides these two processes, also P25/H<sub>2</sub>O<sub>2</sub> prevented its  
392 reactivation.

393 The effect of the different treatment processes on the bacterial communities was another aim of  
394 this study. Out of the 49 phyla found in freshly collected wastewater (before treatment),  
395 *Proteobacteria* (62 ± 7%) and *Bacteroidetes* (10 ± 2%) were, in average (n=8), the most abundant  
396 (Fig. 5a, t<sub>0</sub>). *Proteobacteria* comprised mainly members of the classes *Beta-* (28 ± 2%), *Gamma-*  
397 (20 ± 10%), *Alpha-* (7 ± 2%) and *Deltaproteobacteria* (5 ± 1%). Beside these groups, several other  
398 bacterial classes were detected in the freshly collected wastewater in relative abundances ranging  
399 from 2 ± 1% to 4 ± 2%; examples are “*Saprospirae*”, *Flavobacteriia* and *Bacteroidia*

400 (*Bacteroidetes*), *Clostridia* (*Firmicutes*), *Planctomycetia* (*Planctomycetes*), *ZB2* (*OD1*) and *TM7-*  
 401 *I* (*TM7*). The other identified classes had abundances < 1.3% (Fig. 5a, t<sub>0</sub>).  
 402



403  
 404 **Figure 4.** Abundance of target genes before and after treatment, and after 3-days storage at room  
 405 temperature using different solar-driven treatments: (a) 16S rRNA, (b) *intI1*, (c) *qnrS*, (d) *bla*<sub>CTX-M</sub>, (e) *sul1*  
 406 and (f) *bla*<sub>TEM</sub>. Values are the average of two independent assays. Error bars represent standard deviations.

407

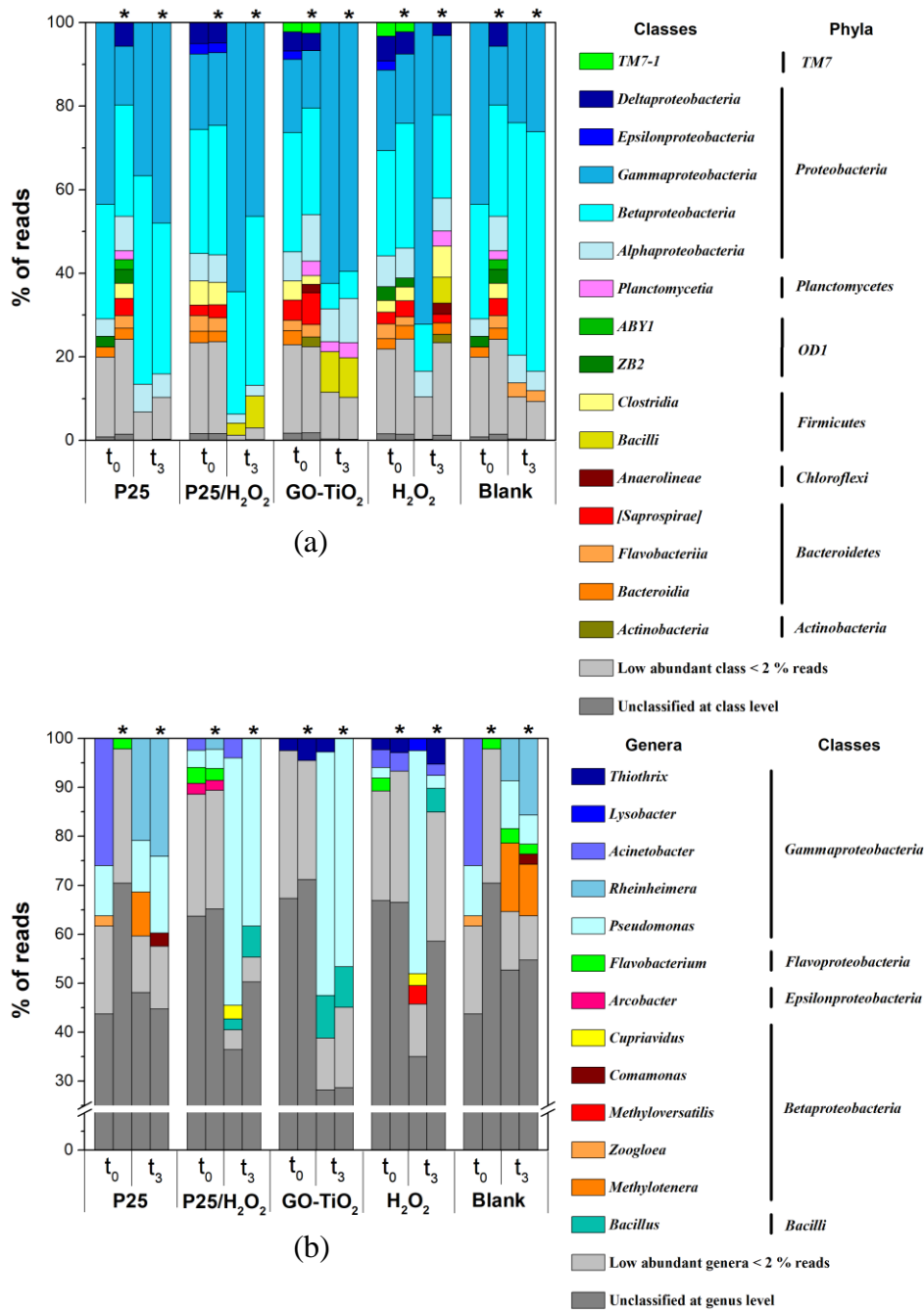
408 Regardless of the solar driven process, treatment followed by storage at room conditions led to  
 409 important bacterial community rearrangements that, in general, had the same pattern. The relative  
 410 abundance of the members of *Proteobacteria* was higher ( $p < 0.01$ ) in the stored treated wastewater  
 411 than in the freshly collected wastewater samples, whereas it was lower ( $p < 0.01$ ) for the majority  
 412 of members of the other phyla (Fig. 5a,  $t_3$ ). The lower values of the alpha-diversity indices of the  
 413 stored treated wastewater samples when compared with those of freshly collected wastewater  
 414 samples (Table 3) corroborate this loss of diversity and equitability. These rearrangements are well  
 415 depicted in the PCoA biplot, where the bacterial communities of the freshly collected wastewater  
 416 samples were separated from those treated and stored over axis PC1 (31.0%) (Fig. 6, squares and  
 417 stars, respectively).

418

419 **Table 3.** Alpha diversity indices of the wastewater samples before ( $t_0$ ) and after 3-days storage after  
 420 treatment ( $t_3$ ) calculated based on the average of 10 rarefaction OTU tables.

Experiment	Time	OTUs No.	Shannon	Simpson	PD whole tree
P25	$t_0$	3023	7.0	0.95	125.6
P25 *	$t_0$	3181	8.3	0.99	158.7
<b>P25</b>	<b><math>t_3</math></b>	3267	6.6	0.92	101.8
<b>P25 *</b>	<b><math>t_3</math></b>	4010	7.3	0.96	123.8
P25 / H <sub>2</sub> O <sub>2</sub>	$t_0$	3353	7.8	0.98	156.7
P25 / H <sub>2</sub> O <sub>2</sub> *	$t_0$	3586	8.1	0.98	164.8
<b>P25 / H<sub>2</sub>O<sub>2</sub></b>	<b><math>t_3</math></b>	2462	6.1	0.93	59.4
<b>P25 / H<sub>2</sub>O<sub>2</sub> *</b>	<b><math>t_3</math></b>	2047	4.9	0.84	58.7
GO-TiO <sub>2</sub>	$t_0$	3425	8.1	0.98	161.2
GO-TiO <sub>2</sub> *	$t_0$	2687	7.6	0.97	136.3
<b>GO-TiO<sub>2</sub></b>	<b><math>t_3</math></b>	2419	5.6	0.83	95.5
<b>GO-TiO<sub>2</sub> *</b>	<b><math>t_3</math></b>	2628	6.5	0.93	92.7
H <sub>2</sub> O <sub>2</sub>	$t_0$	3161	7.8	0.98	155.7
H <sub>2</sub> O <sub>2</sub> *	$t_0$	3567	8.1	0.98	166.3
<b>H<sub>2</sub>O<sub>2</sub></b>	<b><math>t_3</math></b>	2179	5.2	0.84	79.4
<b>H<sub>2</sub>O<sub>2</sub> *</b>	<b><math>t_3</math></b>	2595	6.3	0.91	136.2
Blank	$t_0$	3023	7.0	0.95	125.6
Blank *	$t_0$	3181	8.3	0.99	158.7
<b>Blank</b>	<b><math>t_3</math></b>	3487	6.7	0.94	112.4
<b>Blank *</b>	<b><math>t_3</math></b>	3511	6.3	0.91	111.0

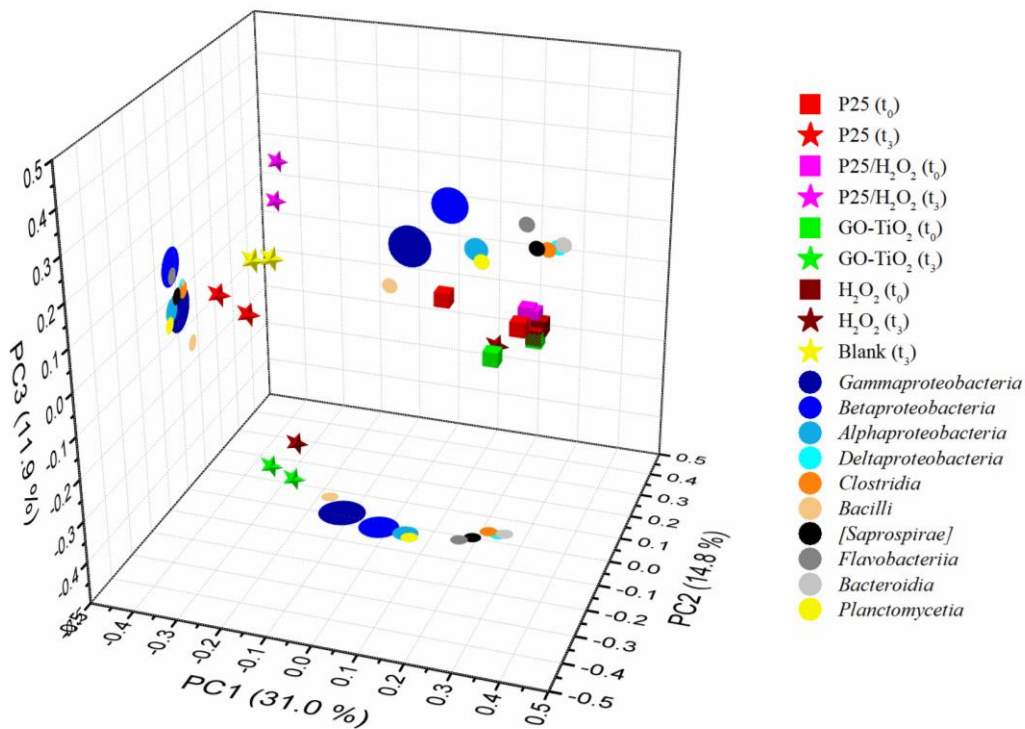
421 \*, indicates a second independent assay.



422  
 423 **Figure 5.** Relative abundance of (a) classes and (b) genera before (t<sub>0</sub>) and 3-days after treatment (t<sub>3</sub>). \*,  
 424 indicates a second independent assay.

425  
 426 Differences on the structure and composition of the bacterial community of the stored treated  
 427 wastewater samples were mainly based on the relative abundance of *Beta*- and  
 428 *Gammaproteobacteria* and *Bacilli* (Figs. 5a, t<sub>3</sub> and Fig. 6, stars). The structure and composition of  
 429 the bacterial communities of treated stored water was similar. Nevertheless, the relative abundance

430 of *Betaproteobacteria* was higher for photolysis > P25 > P25/H<sub>2</sub>O<sub>2</sub> > solar-H<sub>2</sub>O<sub>2</sub> > GO-TiO<sub>2</sub>,  
 431 varying between 57% and 7% (Fig. 5a). Consequently, the relative abundance of  
 432 *Gammaproteobacteria* followed a kind of inverse order (i.e., GO-TiO<sub>2</sub> > P25/H<sub>2</sub>O<sub>2</sub> > P25 > solar-  
 433 H<sub>2</sub>O<sub>2</sub> > photolysis), varying between 61% and 25% (Fig 5a). Despite the lower values when  
 434 compared with these proteobacterial classes, the relative abundance of *Bacilli* followed a similar  
 435 order (8% and 0%) (Fig.5a). The ability of members of class *Bacilli* to survive under harsh stressful  
 436 conditions through the production of resistance forms (endospores) as well as their ability to  
 437 withstand oxidative stress (Battistuzzi and Hedges 2009, Mols and Abee 2011) may explain their  
 438 survival upon these treatments.  
 439



440  
 441 **Figure 6.** Biplot of principal coordinates analysis (PCoA) based on weighted Unifrac distances of samples  
 442 before (t<sub>0</sub> - squares) and 3-days storage after treatment (t<sub>3</sub> – stars).  
 443  
 444 Altogether, the results obtained suggest that the solar-driven AOPs inactivated less efficiently *Beta*-  
 445 and/or *Gammaproteobacteria* or that bacteria belonging to these classes have higher capacity to

446 regrow. Curiously, at the genus level, it is possible to observe that members of the ubiquitous genus  
447 *Pseudomonas* were, in general, the group with the sharpest increase during storage, followed by  
448 *Rheinheimera* and *Methilotenera* (Fig. 5b). Alterations in the composition and structure of the  
449 bacterial communities leading to higher proportions of *Proteobacteria* in stored water than before  
450 the treatment were reported in the literature for photolysis, ozonation and photocatalytic ozonation  
451 (Becerra-Castro et al. 2016), coupled biological and photocatalysis treatments (Chen et al. 2013)  
452 and ozonation coupled to a sequencing batch biofilm reactor (Esplugas et al. 2013).

453

## 454 **Conclusions**

455 Among the tested solar-driven oxidation processes, photo-Fenton at circumneutral pH was the  
456 worst performing one (quite similar to photolysis), whereas the combination of P25 and H<sub>2</sub>O<sub>2</sub> was  
457 the most efficient approach for the removal of organic micropollutants in the urban wastewater  
458 sampled.

459 Regarding the biological indicators, a decrease in the abundance of total faecal coliforms and  
460 enterococci and their antibiotic resistant counterparts was found for all the processes employing  
461 H<sub>2</sub>O<sub>2</sub>, which was permanent after 3-days storage of the treated wastewater. P25/H<sub>2</sub>O<sub>2</sub> and solar-  
462 H<sub>2</sub>O<sub>2</sub> were also able to reduce the total bacterial load, assessed based on the abundance of the 16S  
463 rRNA gene. Nevertheless, the abundance of total bacterial load increased after 3-days storage to  
464 values close or higher than those verified before treatment. Similar observations were found for the  
465 genes *intI1* and *sull*. Hence, none of the studied processes was able to prevent bacterial  
466 reactivation, including antibiotic resistant populations.

467 Thus, among all the studied processes, P25/H<sub>2</sub>O<sub>2</sub> seemed to be that achieving the best compromise  
468 for the removal of both organic micropollutants and biological contaminants, although not able to  
469 prevent bacterial reactivation.

470 Interestingly, regardless of the oxidation process studied, higher relative abundance of the phylum  
471 *Proteobacteria* (*Beta-* and *Gammaproteobacteria*), namely of genera *Pseudomonas*, *Rheinheimera*

472 and *Methylothermobacter*, was observed in treated wastewater after 3-days storage. Since within the  
473 phylum *Proteobacteria*, in particular of the classes *Beta-* and *Gammaproteobacteria*, it is possible  
474 to find diverse multidrug-resistant bacteria, the increase of this group of organisms in stored treated  
475 water may deserve further investigation. Moreover, the potential disturbance of the water bacterial  
476 communities may have relevant ecology implication and should be considered in the design of  
477 advanced oxidation technologies.

478

## 479 **Acknowledgments**

480 This work was financially supported by Project nº P1404290052 under the SFERA Program  
481 (EC/FP7 - Integrating Activities), Project POCI-01-0145-FEDER-006984 – Associate Laboratory  
482 LSRE-LCM (UID/EQU/50020/2013) and POCI-01-0145-FEDER-006939 (LEPABE –  
483 UID/EQU/00511/2013), funded by FEDER through COMPETE2020 - Programa Operacional  
484 Competitividade e Internacionalização (POCI) – and by national funds through FCT - Fundação  
485 para a Ciência e a Tecnologia; UID/Multi/50016/2013-CBQF and Water JPI/0001/2013 STARE,  
486 and partially co-financed by QREN, ON2, FCT and FEDER through project AIProcMat@N2020  
487 NORTE-01-0145-FEDER-000006, NORTE-07-0162-FEDER-000050 and NORTE-01-0145-  
488 FEDER-000005 (LEPABE-2-ECO-INNOVATION), supported by North Portugal Regional  
489 Operational Program (NORTE 2020), under the Portugal 2020 Partnership Agreement, through the  
490 ERDF. NFFM, LMPM and AMTS acknowledge PD/BD/114318/2016, IF/01248/2014 and  
491 IF/01501/2013, respectively. The authors would like to acknowledge the financial support provided  
492 by COST-European Cooperation in Science and Technology, to the COST Action ES1403: New  
493 and emerging challenges and opportunities in wastewater reuse (NEREUS). Disclaimer: The  
494 content of this article is the authors' responsibility and neither COST nor any person acting on its  
495 behalf is responsible for the use, which might be made of the information contained in it.

496

## 497 **References**



498 Fatta-Kassinou, D., Kalavrouzioti, I.K., Koukoulaki, P.H. and Vasquez, M.I. (2011) The risks  
499 associated with wastewater reuse and xenobiotics in the agroecological environment. *Science of*  
500 *The Total Environment* 409, 3555-3563.

501 Manaia, C.M., Macedo, G., Fatta-Kassinou, D. and Nunes, O.C. (2016) Antibiotic resistance in  
502 urban aquatic environments: can it be controlled? *Applied Microbiology and Biotechnology* 100,  
503 1543-1557.

504 Rizzo, L., Manaia, C., Merlin, C., Schwartz, T., Dagot, C., Ploy, M.C., Michael, I. and Fatta-  
505 Kassinou, D. (2013) Urban wastewater treatment plants as hotspots for antibiotic resistant bacteria  
506 and genes spread into the environment: A review. *Science of The Total Environment* 447, 345-360.

507 Ferro, G., Guarino, F., Castiglione, S. and Rizzo, L. (2016) Antibiotic resistance spread potential  
508 in urban wastewater effluents disinfected by UV/H<sub>2</sub>O<sub>2</sub> process. *Science of The Total Environment*  
509 560–561, 29-35.

510 Davison, J. (1999) Genetic Exchange between Bacteria in the Environment. *Plasmid* 42, 73-91.

511 Ribeiro, A.R., Nunes, O.C., Pereira, M.F.R. and Silva, A.M.T. (2015) An overview on the  
512 advanced oxidation processes applied for the treatment of water pollutants defined in the recently  
513 launched Directive 2013/39/EU. *Environment International* 75, 33-51.

514 Moreira, N.F.F., Sousa, J.M., Macedo, G., Ribeiro, A.R., Barreiros, L., Pedrosa, M., Faria, J.L.,  
515 Pereira, M.F.R., Castro-Silva, S., Segundo, M.A., Manaia, C.M., Nunes, O.C. and Silva, A.M.T.  
516 (2016) Photocatalytic ozonation of urban wastewater and surface water using immobilized TiO<sub>2</sub>  
517 with LEDs: Micropollutants, antibiotic resistance genes and estrogenic activity. *Water Research*  
518 94, 10-22.

519 Malato, S., Fernández-Ibáñez, P., Maldonado, M.I., Blanco, J. and Gernjak, W. (2009)  
520 Decontamination and disinfection of water by solar photocatalysis: Recent overview and trends.  
521 *Catalysis Today* 147, 1-59.

522 Sousa, J.M., Macedo, G., Pedrosa, M., Becerra-Castro, C., Castro-Silva, S., Pereira, M.F.R., Silva,  
523 A.M.T., Nunes, O.C. and Manaia, C.M. (2017) Ozonation and UV<sub>254 nm</sub> radiation for the removal

524 of microorganisms and antibiotic resistance genes from urban wastewater. *Journal of Hazardous*  
525 *Materials* 323, 434-441.

526 Polo-López, M.I., Castro-Alferez, M., Oller, I. and Fernández-Ibáñez, P. (2014) Assessment of  
527 solar photo-Fenton, photocatalysis, and H<sub>2</sub>O<sub>2</sub> for removal of phytopathogen fungi spores in  
528 synthetic and real effluents of urban wastewater. *Chemical Engineering Journal* 257, 122-130.

529 Pablos, C., Marugán, J., van Grieken, R. and Serrano, E. (2013) Emerging micropollutant oxidation  
530 during disinfection processes using UV-C, UV-C/H<sub>2</sub>O<sub>2</sub>, UV-A/TiO<sub>2</sub> and UV-A/TiO<sub>2</sub>/H<sub>2</sub>O<sub>2</sub>. *Water*  
531 *Research* 47, 1237-1245.

532 Yang, W., Zhou, H. and Cicek, N. (2014) Treatment of organic micropollutants in water and  
533 wastewater by UV-based processes: a literature review. *Critical Reviews in Environmental Science*  
534 *and Technology* 44, 1443-1476.

535 Dunlop, P.S.M., Sheeran, C.P., Byrne, J.A., McMahon, M.A.S., Boyle, M.A. and McGuigan, K.G.  
536 (2010) Inactivation of clinically relevant pathogens by photocatalytic coatings. *Journal of*  
537 *Photochemistry and Photobiology A: Chemistry* 216, 303-310.

538 Ferro, G., Polo-López, M.I., Martínez-Piernas, A.B., Fernández-Ibáñez, P., Agüera, A. and Rizzo,  
539 L. (2015) Cross-Contamination of Residual Emerging Contaminants and Antibiotic Resistant  
540 Bacteria in Lettuce Crops and Soil Irrigated with Wastewater Treated by Sunlight/H<sub>2</sub>O<sub>2</sub>.  
541 *Environmental Science & Technology* 49, 11096-11104.

542 Fiorentino, A., Ferro, G., Alferez, M.C., Polo-López, M.I., Fernández-Ibáñez, P. and Rizzo, L.  
543 (2015) Inactivation and regrowth of multidrug resistant bacteria in urban wastewater after  
544 disinfection by solar-driven and chlorination processes. *Journal of Photochemistry and*  
545 *Photobiology B: Biology* 148, 43-50.

546 Becerra-Castro, C., Macedo, G., Silva, A.M.T., Manaia, C.M. and Nunes, O.C. (2016)  
547 Proteobacteria become predominant during regrowth after water disinfection. *Science of The Total*  
548 *Environment* 573, 313-323.

549 Pastrana-Martínez, L.M., Morales-Torres, S., Likodimos, V., Figueiredo, J.L., Faria, J.L., Falaras,  
550 P. and Silva, A.M.T. (2012) Advanced nanostructured photocatalysts based on reduced graphene  
551 oxide–TiO<sub>2</sub> composites for degradation of diphenhydramine pharmaceutical and methyl orange  
552 dye. *Applied Catalysis B: Environmental* 123, 241-256.

553 Rodríguez-Chueca, J., Polo-López, M.I., Mosteo, R., Ormad, M.P. and Fernández-Ibáñez, P.  
554 (2014) Disinfection of real and simulated urban wastewater effluents using a mild solar photo-  
555 Fenton. *Applied Catalysis B: Environmental* 150–151, 619-629.

556 Booshehri, A.Y., Polo-Lopez, M.I., Castro-Alfárez, M., He, P., Xu, R., Rong, W., Malato, S. and  
557 Fernández-Ibáñez, P. (2017) Assessment of solar photocatalysis using Ag/BiVO<sub>4</sub> at pilot solar  
558 Compound Parabolic Collector for inactivation of pathogens in well water and secondary effluents.  
559 *Catalysis Today* 281, 124-134.

560 Malato, S., Fernández-Ibáñez, P., Maldonado, M.I. and Oller, I. (2013) New and Future  
561 Developments in Catalysis, pp. 371-393, Elsevier, Amsterdam.

562 Fernández-Ibáñez, P., Malato, S. and de las Nieves, F.J. (1999) Relationship between TiO<sub>2</sub> particle  
563 size and reactor diameter in solar photoreactors efficiency. *Catalysis Today* 54(2), 195-204.

564 Barbosa, M.O., Moreira, N.F.F., Ribeiro, A.R., Pereira, M.F.R. and Silva, A.M.T. (2016)  
565 Occurrence and removal of organic micropollutants: An overview of the watch list of EU Decision  
566 2015/495. *Water Research* 94, 257-279.

567 Polo-López, M.I., Fernández-Ibáñez, P., Ubomba-Jaswa, E., Navntoft, C., García-Fernández, I.,  
568 Dunlop, P.S.M., Schmid, M., Byrne, J.A. and McGuigan, K.G. (2011) Elimination of water  
569 pathogens with solar radiation using an automated sequential batch CPC reactor. *Journal of*  
570 *Hazardous Materials* 196, 16-21.

571 Narciso-da-Rocha, C., Varela, A.R., Schwartz, T., Nunes, O.C. and Manaia, C.M. (2014) *bla*TEM  
572 and *vanA* as indicator genes of antibiotic resistance contamination in a hospital–urban wastewater  
573 treatment plant system. *Journal of Global Antimicrobial Resistance* 2, 309-315.

574 Denman, S.E. and McSweeney, C.S. (2006) Development of a real-time PCR assay for monitoring  
575 anaerobic fungal and cellulolytic bacterial populations within the rumen. *FEMS Microbiology*  
576 *Ecology* 58, 572-582.

577 Bibbal, D., Dupouy, V., Ferré, J.P., Toutain, P.L., Fayet, O., Prère, M.F. and Bousquet-Mélou, A.  
578 (2007) Impact of three ampicillin dosage regimens on selection of ampicillin resistance in  
579 Enterobacteriaceae and excretion of blaTEM genes in swine feces. *Applied and Environmental*  
580 *Microbiology* 73, 4785-4790.

581 Goldstein, C., Lee, M.D., Sanchez, S., Hudson, C., Phillips, B., Register, B., Grady, M., Liebert,  
582 C., Summers, A.O., White, D.G. and Maurer, J.J. (2001) Incidence of class 1 and 2 integrases in  
583 clinical and commensal bacteria from livestock, companion animals, and exotics. *Antimicrobial*  
584 *Agents and Chemotherapy* 45, 723-726.

585 Marti, E. and Balcázar, J.L. (2013) Real-time PCR assays for quantification of qnr genes in  
586 environmental water samples and chicken feces. *Applied and Environmental Microbiology* 79,  
587 1743-1745.

588 Pei, R., Kim, S.C., Carlson, K.H. and Pruden, A. (2006) Effect of River Landscape on the sediment  
589 concentrations of antibiotics and corresponding antibiotic resistance genes (ARG). *Water Research*  
590 40, 2427-2435.

591 Volkmann, H., Schwartz, T., Bischoff, P., Kirchen, S. and Obst, U. (2004) Detection of clinically  
592 relevant antibiotic-resistance genes in municipal wastewater using real-time PCR (TaqMan).  
593 *Journal of Microbiological Methods* 56, 277-286.

594 Marti, E., Variatza, E. and Balcázar, J.L. (2014) Bacteriophages as a reservoir of extended-  
595 spectrum  $\beta$ -lactamase and fluoroquinolone resistance genes in the environment. *Clinical*  
596 *Microbiology and Infection* 20, O456-O459.

597 Caporaso, J.G., Kuczynski, J., Stombaugh, J., Bittinger, K., Bushman, F.D., Costello, E.K., Fierer,  
598 N., Peña, A.G., Goodrich, J.K., Gordon, J.I., Huttley, G.A., Kelley, S.T., Knights, D., Koenig, J.E.,  
599 Ley, R.E., Lozupone, C.A., McDonald, D., Muegge, B.D., Pirrung, M., Reeder, J., Sevinsky, J.R.,

600 Turnbaugh, P.J., Walters, W.A., Widmann, J., Yatsunenko, T., Zaneveld, J. and Knight, R. (2010a)  
601 QIIME allows analysis of high-throughput community sequencing data. *Nature methods* 7, 335-  
602 336.

603 Schmieder, R. and Edwards, R. (2011) Quality control and preprocessing of metagenomic datasets.  
604 *Bioinformatics* 27, 863-864.

605 Edgar, R.C. (2010) Search and clustering orders of magnitude faster than BLAST. *Bioinformatics*  
606 26, 2460-2461.

607 Caporaso, J.G., Bittinger, K., Bushman, F.D., DeSantis, T.Z., Andersen, G.L. and Knight, R.  
608 (2010b) PyNAST: a flexible tool for aligning sequences to a template alignment. *Bioinformatics*  
609 26, 266-267.

610 Shannon, C.E. and Weaver, W. (1963) *The mathematical theory of communication*, University of  
611 Illinois Press, Urbana.

612 Simpson, E.H. (1949) Measurement of diversity. *Nature* 163, 688.

613 Faith, D.P. (1992) Conservation evaluation and phylogenetic diversity. *Biological Conservation*  
614 61, 1-10.

615 Paulson, J.N., Stine, O.C., Bravo, H.C. and Pop, M. (2013) Differential abundance analysis for  
616 microbial marker-gene surveys. *Nat Meth* 10, 1200-1202.

617 Lozupone, C. and Knight, R. (2005) UniFrac: a New Phylogenetic Method for Comparing  
618 Microbial Communities. *Applied and Environmental Microbiology* 71, 8228-8235.

619 Parks, D.H., Tyson, G.W., Hugenholtz, P. and Beiko, R.G. (2014) STAMP: statistical analysis of  
620 taxonomic and functional profiles. *Bioinformatics* 30, 3123-3124.

621 Moreira, N.F.F., Orge, C.A., Ribeiro, A.R., Faria, J.L., Nunes, O.C., Pereira, M.F.R. and Silva,  
622 A.M.T. (2015) Fast mineralization and detoxification of amoxicillin and diclofenac by  
623 photocatalytic ozonation and application to an urban wastewater. *Water Research* 87, 87-96.

624 Cruz, M., Gomez, C., Duran-Valle, C.J., Pastrana-Martínez, L.M., Faria, J.L., Silva, A.M.T.,  
625 Faraldos, M. and Bahamonde, A. (2017) Bare TiO<sub>2</sub> and graphene oxide TiO<sub>2</sub> photocatalysts on the

626 degradation of selected pesticides and influence of the water matrix. *Applied Surface Science* 416,  
627 1013-1021.

628 Sampaio, M.J., Silva, C.G., Silva, A.M.T., Pastrana-Martínez, L.M., Han, C., Morales-Torres, S.,  
629 Figueiredo, J.L., Dionysiou, D.D. and Faria, J.L. (2015) Carbon-based TiO<sub>2</sub> materials for the  
630 degradation of Microcystin-LA. *Applied Catalysis B: Environmental* 170–171, 74-82.

631 Maroga Mboula, V., Héquet, V., Andrès, Y., Pastrana-Martínez, L.M., Doña-Rodríguez, J.M.,  
632 Silva, A.M.T. and Falaras, P. (2013) Photocatalytic degradation of endocrine disruptor compounds  
633 under simulated solar light. *Water Research* 47, 3997-4005.

634 Kositzi, M., Poullos, I., Malato, S., Caceres, J. and Campos, A. (2004) Solar photocatalytic  
635 treatment of synthetic municipal wastewater. *Water Research* 38, 1147-1154.

636 Xing, W., Lalwani, G., Rusakova, I. and Sitharaman, B. (2014) Degradation of graphene by  
637 hydrogen peroxide. *Particle & Particle Systems Characterization* 31, 745-750.

638 Giannakis, S., Hendaoui, I., Rtimi, S., Fürbringer, J.-M. and Pulgarin, C. (2017) Modeling and  
639 treatment optimization of pharmaceutically active compounds by the photo-Fenton process: The  
640 case of the antidepressant Venlafaxine. *Journal of Environmental Chemical Engineering* 5, 818-  
641 828.

642 Agulló-Barceló, M., Polo-López, M.I., Lucena, F., Jofre, J. and Fernández-Ibáñez, P. (2013) Solar  
643 advanced oxidation processes as disinfection tertiary treatments for real wastewater: implications  
644 for water reclamation. *Applied Catalysis B: Environmental* 136, 341-350.

645 García-Fernández, I., Polo-López, M.I., Oller, I. and Fernández-Ibáñez, P. (2012) Bacteria and  
646 fungi inactivation using Fe<sup>3+</sup>/sunlight, H<sub>2</sub>O<sub>2</sub>/sunlight and near neutral photo-Fenton: A  
647 comparative study. *Applied Catalysis B: Environmental* 121, 20-29.

648 Aguas, Y., Hincapie, M., Fernández-Ibáñez, P. and Polo-López, M.I. (2017) Solar photocatalytic  
649 disinfection of agricultural pathogenic fungi (*Curvularia* sp.) in real urban wastewater. *Science of*  
650 *The Total Environment* 607, 1213-1224.

651 Castro-Alfárez, M., Polo-López, M.I., Marugán, J. and Fernández-Ibáñez, P. (2017) Mechanistic  
652 modeling of UV and mild-heat synergistic effect on solar water disinfection. *Chemical Engineering*  
653 *Journal* 316, 111-120.

654 Polo-López, M.I., Castro-Alfárez, M., Nahim-Granados, S., Malato, S. and Fernández-Ibáñez, P.  
655 (2017) *Legionella jordanis* inactivation in water by solar driven processes: EMA-qPCR versus  
656 culture-based analyses for new mechanistic insights. *Catalysis Today* 287, 15-21.

657 Sichel, C., Fernández-Ibáñez, P., de Cara, M. and Tello, J. (2009) Lethal synergy of solar UV-  
658 radiation and H<sub>2</sub>O<sub>2</sub> on wild *Fusarium solani* spores in distilled and natural well water. *Water*  
659 *Research* 43, 1841-1850.

660 Polo-López, M.I., Oller, I. and Fernández-Ibáñez, P. (2013) Benefits of photo-Fenton at low  
661 concentrations for solar disinfection of distilled water. A case study: *Phytophthora capsici*.  
662 *Catalysis Today* 209, 181-187.

663 Cruz-Ortiz, B.R., Hamilton, J.W.J., Pablos, C., Díaz-Jiménez, L., Cortés-Hernández, D.A.,  
664 Sharma, P.K., Castro-Alfárez, M., Fernández-Ibáñez, P., Dunlop, P.S.M. and Byrne, J.A. (2017)  
665 Mechanism of photocatalytic disinfection using titania-graphene composites under UV and visible  
666 irradiation. *Chemical Engineering Journal* 316, 179-186.

667 Fernández-Ibáñez, P., Polo-López, M.I., Malato, S., Wadhwa, S., Hamilton, J.W.J., Dunlop,  
668 P.S.M., D'Sa, R., Magee, E., O'Shea, K., Dionysiou, D.D. and Byrne, J.A. (2015) Solar  
669 photocatalytic disinfection of water using titanium dioxide graphene composites. *Chemical*  
670 *Engineering Journal* 261, 36-44.

671 Zhao, X., Hu, H.Y., Yu, T., Su, C., Jiang, H. and Liu, S. (2014) Effect of different molecular weight  
672 organic components on the increase of microbial growth potential of secondary effluent by  
673 ozonation. *Journal of Environmental Sciences (China)* 26, 2190-2197.

674 Spuhler, D., Andrés Rengifo-Herrera, J. and Pulgarin, C. (2010) The effect of Fe<sup>2+</sup>, Fe<sup>3+</sup>, H<sub>2</sub>O<sub>2</sub> and  
675 the photo-Fenton reagent at near neutral pH on the solar disinfection (SODIS) at low temperatures  
676 of water containing *Escherichia coli* K12. *Applied Catalysis B: Environmental* 96(1–2), 126-141.

677 Ubomba-Jaswa, E., Navntoft, C., Polo-López, M.I., Fernandez-Ibáñez, P. and McGuigan, K.G.  
678 (2009) Solar disinfection of drinking water (SODIS): An investigation of the effect of UV-A dose  
679 on inactivation efficiency. *Photochemical and Photobiological Sciences* 8, 587-595.

680 Giannakis, S., Merino Gamo, A.I., Darakas, E., Escalas-Cañellas, A. and Pulgarin, C. (2014)  
681 Monitoring the post-irradiation *E. coli* survival patterns in environmental water matrices:  
682 Implications in handling solar disinfected wastewater. *Chemical Engineering Journal* 253, 366-  
683 376.

684 Battistuzzi, F.U. and Hedges, S.B. (2009) A Major Clade of Prokaryotes with Ancient Adaptations  
685 to Life on Land. *Molecular Biology and Evolution* 26, 335-343.

686 Mols, M. and Abee, T. (2011) Primary and secondary oxidative stress in *Bacillus*. *Environmental*  
687 *Microbiology* 13, 1387-1394.

688 Chen, C.-Y., Kuo, J.-T., Yang, H.-A. and Chung, Y.-C. (2013) A coupled biological and  
689 photocatalysis pretreatment system for the removal of crystal violet from wastewater.  
690 *Chemosphere* 92, 695-701.

691 Esplugas, M., González, O. and Sans, C. (2013) Bacterial community characterization of a  
692 sequencing batch reactor treating pre-ozonized sulfamethoxazole in water. *Environmental*  
693 *Technology* 34, 1583-1591.

694